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(54) Title: COMPOSITIONS AND METHODS FOR THE THERAPY AND DIAGNOSIS OF OVARIAN AND ENDOMETRIAL CANCER

(57) Abstract: Compositions and methods for the therapy and diagnosis of cancer, such as ovarian or endometrial cancer, are disclosed. Compositions may comprise one or more ovarian carcinoma proteins, immunogenic portions thereof, or polynucleotides that encode such portions. Alternatively, a therapeutic composition may comprise an antigen presenting cell that expresses such an antigen, or a T cell that is specific for cells expressing such an antigen. Such compositions may be used, for example, for the prevention and treatment of diseases such as ovarian and endometrial cancer. Diagnostic methods based on detecting an ovarian carcinoma protein, or mRNA encoding such an antigen, in a sample are also provided.

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COMPOSITIONS AND METHODS FOR THE THERAPY AND DIAGNOSIS OF OVARIAN AND ENDOMETRIAL CANCER

TECHNICAL FIELD OF THE INVENTION

The present invention relates generally to therapy and diagnosis of cancer, such as ovarian or endometrial cancer. The invention is more specifically related to polypeptides comprising at least a portion of an ovarian carcinoma protein, and to polynucleotides encoding such polypeptides. Such polypeptides and polynucleotides may be used in vaccines and pharmaceutical compositions for prevention and treatment of cancers such as ovarian and endometrial cancer, and for the diagnosis and monitoring of such cancers.

BACKGROUND OF THE INVENTION

Ovarian cancer is a significant health problem for women in the United States and throughout the world. Although advances have been made in detection and therapy of this cancer, no vaccine or other universally successful method for prevention or treatment is currently available. Management of the disease currently relies on a combination of early diagnosis and aggressive treatment, which may include one or more of a variety of treatments such as surgery, radiotherapy, chemotherapy and hormone therapy. The course of treatment for a particular cancer is often selected based on a variety of prognostic parameters, including an analysis of specific tumor markers. However, the use of established markers often leads to a result that is difficult to interpret, and high mortality continues to be observed in many cancer patients.

Immunotherapies have the potential to substantially improve cancer treatment and survival. Such therapies may involve the generation or enhancement of an immune response to an ovarian carcinoma protein. However, to date, relatively few ovarian carcinoma proteins are known and the generation of an immune response against such antigens has not been shown to be therapeutically beneficial.

Accordingly, there is a need in the art for improved methods for detecting and treating cancers such as ovarian cancer. The present invention fulfills these needs and further provides other related advantages.

SUMMARY OF THE INVENTION

Briefly stated, the present invention provides compositions and methods for the diagnosis and therapy of cancer, such as ovarian and endometrial cancer. In one aspect, the present invention provides polypeptides comprising at least a portion of an ovarian carcinoma protein, or a variant thereof. Certain portions and other variants are immunogenic, such that the ability of the variant to react with antigen-specific antisera is not substantially diminished. Within certain embodiments, the polypeptide comprises a sequence that is encoded by a polynucleotide sequence selected from the group consisting of: (a) sequences recited in SEQ ID NO:1-222; (b) variants of a sequence recited in SEQ ID NO:1-222 and (c) complements of a sequence of (a) or (b).

The present invention further provides polynucleotides that encode a polypeptide as described above, or a portion thereof (such as a portion encoding at least 15 amino acid residues of an ovarian carcinoma protein), expression vectors comprising such polynucleotides and host cells transformed or transfected with such expression vectors.

Within other aspects, the present invention provides pharmaceutical compositions comprising a polypeptide or polynucleotide as described above and a physiologically acceptable carrier.

Within a related aspect of the present invention, vaccines for prophylactic or therapeutic use are provided. Such vaccines comprise a polypeptide or polynucleotide as described above and an immunostimulant.

The present invention further provides pharmaceutical compositions that comprise: (a) an antibody or antigen-binding fragment thereof that specifically binds to an ovarian carcinoma protein; and (b) a physiologically acceptable carrier.

Within further aspects, the present invention provides pharmaceutical compositions comprising: (a) an antigen presenting cell that expresses a polypeptide as described above and (b) a pharmaceutically acceptable carrier or excipient. Antigen presenting cells include dendritic cells, macrophages, monocytes, fibroblasts and B cells.

Within related aspects, vaccines are provided that comprise: (a) an antigen presenting cell that expresses a polypeptide as described above and (b) an immunostimulant.

The present invention further provides, in other aspects, fusion proteins that comprise at least one polypeptide as described above, as well as polynucleotides encoding such fusion proteins.

Within related aspects, pharmaceutical compositions comprising a fusion
5 protein, or a polynucleotide encoding a fusion protein, in combination with a physiologically acceptable carrier are provided.

Vaccines are further provided, within other aspects, that comprise a fusion protein, or a polynucleotide encoding a fusion protein, in combination with an immunostimulant.

10 Within further aspects, the present invention provides methods for inhibiting the development of a cancer in a patient, comprising administering to a patient a pharmaceutical composition or vaccine as recited above. The patient may be afflicted with ovarian or endometrial cancer, in which case the methods provide treatment for the disease, or patient considered at risk for such a disease may be treated
15 prophylactically.

The present invention further provides, within other aspects, methods for removing tumor cells from a biological sample, comprising contacting a biological sample with T cells that specifically react with an ovarian carcinoma protein, wherein the step of contacting is performed under conditions and for a time sufficient to permit
20 the removal of cells expressing the protein from the sample.

Within related aspects, methods are provided for inhibiting the development of a cancer in a patient, comprising administering to a patient a biological sample treated as described above.

Methods are further provided, within other aspects, for stimulating
25 and/or expanding T cells specific for an ovarian carcinoma protein, comprising contacting T cells with one or more of: (i) a polypeptide as described above; (ii) a polynucleotide encoding such a polypeptide; and/or (iii) an antigen presenting cell that expresses such a polypeptide; under conditions and for a time sufficient to permit the stimulation and/or expansion of T cells. Isolated T cell populations comprising T cells
30 prepared as described above are also provided.

Within further aspects, the present invention provides methods for inhibiting the development of a cancer in a patient, comprising administering to a patient an effective amount of a T cell population as described above.

The present invention further provides methods for inhibiting the development of a cancer in a patient, comprising the steps of: (a) incubating CD4⁺ and/or CD8⁺ T cells isolated from a patient with one or more of: (i) a polypeptide comprising at least an immunogenic portion of an ovarian carcinoma protein; (ii) a polynucleotide encoding such a polypeptide; and (iii) an antigen-presenting cell that expresses such a polypeptide; and (b) administering to the patient an effective amount of the proliferated T cells, and thereby inhibiting the development of a cancer in the patient. Proliferated cells may, but need not, be cloned prior to administration to the patient.

Within further aspects, the present invention provides methods for determining the presence or absence of a cancer in a patient, comprising: (a) contacting a biological sample obtained from a patient with a binding agent that binds to a polypeptide as recited above; (b) detecting in the sample an amount of polypeptide that binds to the binding agent; and (c) comparing the amount of polypeptide with a predetermined cut-off value, and therefrom determining the presence or absence of a cancer in the patient. Within preferred embodiments, the binding agent is an antibody, more preferably a monoclonal antibody. The cancer may be ovarian or endometrial cancer.

The present invention also provides, within other aspects, methods for monitoring the progression of a cancer in a patient. Such methods comprise the steps of: (a) contacting a biological sample obtained from a patient at a first point in time with a binding agent that binds to a polypeptide as recited above; (b) detecting in the sample an amount of polypeptide that binds to the binding agent; (c) repeating steps (a) and (b) using a biological sample obtained from the patient at a subsequent point in time; and (d) comparing the amount of polypeptide detected in step (c) with the amount detected in step (b) and therefrom monitoring the progression of the cancer in the patient.

The present invention further provides, within other aspects, methods for determining the presence or absence of a cancer in a patient, comprising the steps of: (a)

contacting a biological sample obtained from a patient with an oligonucleotide that hybridizes to a polynucleotide that encodes an ovarian carcinoma protein; (b) detecting in the sample a level of a polynucleotide, preferably mRNA, that hybridizes to the oligonucleotide; and (c) comparing the level of polynucleotide that hybridizes to the oligonucleotide with a predetermined cut-off value, and therefrom determining the presence or absence of a cancer in the patient. Within certain embodiments, the amount of mRNA is detected via polymerase chain reaction using, for example, at least one oligonucleotide primer that hybridizes to a polynucleotide encoding a polypeptide as recited above, or a complement of such a polynucleotide. Within other embodiments, the amount of mRNA is detected using a hybridization technique, employing an oligonucleotide probe that hybridizes to a polynucleotide that encodes a polypeptide as recited above, or a complement of such a polynucleotide.

In related aspects, methods are provided for monitoring the progression of a cancer in a patient, comprising the steps of: (a) contacting a biological sample obtained from a patient with an oligonucleotide that hybridizes to a polynucleotide that encodes an ovarian carcinoma protein; (b) detecting in the sample an amount of a polynucleotide that hybridizes to the oligonucleotide; (c) repeating steps (a) and (b) using a biological sample obtained from the patient at a subsequent point in time; and (d) comparing the amount of polynucleotide detected in step (c) with the amount detected in step (b) and therefrom monitoring the progression of the cancer in the patient.

Within further aspects, the present invention provides antibodies, such as monoclonal antibodies, that bind to a polypeptide as described above, as well as diagnostic kits comprising such antibodies. Diagnostic kits comprising one or more oligonucleotide probes or primers as described above are also provided.

These and other aspects of the present invention will become apparent upon reference to the following detailed description. All references disclosed herein are hereby incorporated by reference in their entirety as if each was incorporated individually.

BRIEF DESCRIPTION OF THE SEQUENCE IDENTIFIERS

SEQ ID NOS: 1-41 are identified in Example 1.

SEQ ID NO:42 is the determined cDNA sequence for clone R0198:A03
SEQ ID NO:43 is the determined cDNA sequence for clone R0198:A07
SEQ ID NO:44 is the determined cDNA sequence for clone R0198:A08
SEQ ID NO:45 is the determined cDNA sequence for clone R0198:A09
5 SEQ ID NO:46 is the determined cDNA sequence for clone R0198:B01
SEQ ID NO:47 is the determined cDNA sequence for clone R0198:B02
SEQ ID NO:48 is the determined cDNA sequence for clone R0198:B04
SEQ ID NO:49 is the determined cDNA sequence for clone R0198:B08
SEQ ID NO:50 is the determined cDNA sequence for clone R0198:B11
10 SEQ ID NO:51 is the determined cDNA sequence for clone R0198:C01
SEQ ID NO:52 is the determined cDNA sequence for clone R0198:C02
SEQ ID NO:53 is the determined cDNA sequence for clone R0198:C03
SEQ ID NO:54 is the determined cDNA sequence for clone R0198:C04
SEQ ID NO:55 is the determined cDNA sequence for clone R0198:C05
15 SEQ ID NO:56 is the determined cDNA sequence for clone R0198:C06
SEQ ID NO:57 is the determined cDNA sequence for clone R0198:C08
SEQ ID NO:58 is the determined cDNA sequence for clone R0198:C09
SEQ ID NO:59 is the determined cDNA sequence for clone R0198:C10
SEQ ID NO:60 is the determined cDNA sequence for clone R0198:C12
20 SEQ ID NO:61 is the determined cDNA sequence for clone R0198:D01
SEQ ID NO:62 is the determined cDNA sequence for clone R0198:D02
SEQ ID NO:63 is the determined cDNA sequence for clone R0198:D03
SEQ ID NO:64 is the determined cDNA sequence for clone R0198:D04
SEQ ID NO:65 is the determined cDNA sequence for clone R0198:D05
25 SEQ ID NO:66 is the determined cDNA sequence for clone R0198:D06
SEQ ID NO:67 is the determined cDNA sequence for clone R0198:D07
SEQ ID NO:68 is the determined cDNA sequence for clone R0198:D08
SEQ ID NO:69 is the determined cDNA sequence for clone R0198:D09
SEQ ID NO:70 is the determined cDNA sequence for clone R0198:D11
30 SEQ ID NO:71 is the determined cDNA sequence for clone R0198:E01
SEQ ID NO:72 is the determined cDNA sequence for clone R0198:E03
SEQ ID NO:73 is the determined cDNA sequence for clone R0198:E05

SEQ ID NO:74 is the determined cDNA sequence for clone R0198:E06
SEQ ID NO:75 is the determined cDNA sequence for clone R0198:E09
SEQ ID NO:76 is the determined cDNA sequence for clone R0198:E10
SEQ ID NO:77 is the determined cDNA sequence for clone R0198:E11
5 SEQ ID NO:78 is the determined cDNA sequence for clone R0198:E12
SEQ ID NO:79 is the determined cDNA sequence for clone R0198:F01
SEQ ID NO:80 is the determined cDNA sequence for clone R0198:F02
SEQ ID NO:81 is the determined cDNA sequence for clone R0198:F03
SEQ ID NO:82 is the determined cDNA sequence for clone R0198:F04
10 SEQ ID NO:83 is the determined cDNA sequence for clone R0198:F06
SEQ ID NO:84 is the determined cDNA sequence for clone R0198:F07
SEQ ID NO:85 is the determined cDNA sequence for clone R0198:F09
SEQ ID NO:86 is the determined cDNA sequence for clone R0198:F10
SEQ ID NO:87 is the determined cDNA sequence for clone R0198:F11
15 SEQ ID NO:88 is the determined cDNA sequence for clone R0198:F12
SEQ ID NO:89 is the determined cDNA sequence for clone R0198:G01
SEQ ID NO:90 is the determined cDNA sequence for clone R0198:G02
SEQ ID NO:91 is the determined cDNA sequence for clone R0198:G03
SEQ ID NO:92 is the determined cDNA sequence for clone R0198:G04
20 SEQ ID NO:93 is the determined cDNA sequence for clone R0198:G05
SEQ ID NO:94 is the determined cDNA sequence for clone R0198:G06
SEQ ID NO:95 is the determined cDNA sequence for clone R0198:G09
SEQ ID NO:96 is the determined cDNA sequence for clone R0198:G11
SEQ ID NO:97 is the determined cDNA sequence for clone R0198:G12
25 SEQ ID NO:98 is the determined cDNA sequence for clone R0198:H01
SEQ ID NO:99 is the determined cDNA sequence for clone R0198:H03
SEQ ID NO:100 is the determined cDNA sequence for clone R0198:H04
SEQ ID NO:101 is the determined cDNA sequence for clone R0198:H06
SEQ ID NO:102 is the determined cDNA sequence for clone R0198:H09
30 SEQ ID NO:103 is the determined cDNA sequence for clone R0198:H10
SEQ ID NO:104 is the determined cDNA sequence for clone R0199:A03
SEQ ID NO:105 is the determined cDNA sequence for clone R0199:A05

SEQ ID NO:106 is the determined cDNA sequence for clone R0199:A06
SEQ ID NO:107 is the determined cDNA sequence for clone R0199:A07
SEQ ID NO:108 is the determined cDNA sequence for clone R0199:A08
SEQ ID NO:109 is the determined cDNA sequence for clone R0199:A11
5 SEQ ID NO:110 is the determined cDNA sequence for clone R0199:B01
SEQ ID NO:111 is the determined cDNA sequence for clone R0199:B03
SEQ ID NO:112 is the determined cDNA sequence for clone R0199:B06
SEQ ID NO:113 is the determined cDNA sequence for clone R0199:B07
SEQ ID NO:114 is the determined cDNA sequence for clone R0199:B08
10 SEQ ID NO:115 is the determined cDNA sequence for clone R0199:B09
SEQ ID NO:116 is the determined cDNA sequence for clone R0199:B11
SEQ ID NO:117 is the determined cDNA sequence for clone R0199:C01
SEQ ID NO:118 is the determined cDNA sequence for clone R0199:C02
SEQ ID NO:119 is the determined cDNA sequence for clone R0199:C06
15 SEQ ID NO:120 is the determined cDNA sequence for clone R0199:C07
SEQ ID NO:121 is the determined cDNA sequence for clone R0199:C08
SEQ ID NO:122 is the determined cDNA sequence for clone R0199:C09
SEQ ID NO:123 is the determined cDNA sequence for clone R0199:C10
SEQ ID NO:124 is the determined cDNA sequence for clone R0199:C11
20 SEQ ID NO:125 is the determined cDNA sequence for clone R0199:C12
SEQ ID NO:126 is the determined cDNA sequence for clone R0199:D01
SEQ ID NO:127 is the determined cDNA sequence for clone R0199:D02
SEQ ID NO:128 is the determined cDNA sequence for clone R0199:D04
SEQ ID NO:129 is the determined cDNA sequence for clone R0199:D06
25 SEQ ID NO:130 is the determined cDNA sequence for clone R0199:D07
SEQ ID NO:131 is the determined cDNA sequence for clone R0199:D08
SEQ ID NO:132 is the determined cDNA sequence for clone R0199:D11
SEQ ID NO:133 is the determined cDNA sequence for clone R0199:E02
SEQ ID NO:134 is the determined cDNA sequence for clone R0199:E03
30 SEQ ID NO:135 is the determined cDNA sequence for clone R0199:E05
SEQ ID NO:136 is the determined cDNA sequence for clone R0199:E06
SEQ ID NO:137 is the determined cDNA sequence for clone R0199:E-8

SEQ ID NO:138 is the determined cDNA sequence for clone R0199:E09
SEQ ID NO:139 is the determined cDNA sequence for clone R0199:E10
SEQ ID NO:140 is the determined cDNA sequence for clone R0199:E12
SEQ ID NO:141 is the determined cDNA sequence for clone R0199:F01
5 SEQ ID NO:142 is the determined cDNA sequence for clone R0199:F03
SEQ ID NO:143 is the determined cDNA sequence for clone R0199:F04
SEQ ID NO:144 is the determined cDNA sequence for clone R0199:F06
SEQ ID NO:145 is the determined cDNA sequence for clone R0199:F09
SEQ ID NO:146 is the determined cDNA sequence for clone R0199:F10
10 SEQ ID NO:147 is the determined cDNA sequence for clone R0199:G01
SEQ ID NO:148 is the determined cDNA sequence for clone R0199:G05
SEQ ID NO:149 is the determined cDNA sequence for clone R0199:G06
SEQ ID NO:150 is the determined cDNA sequence for clone R0199:G08
SEQ ID NO:151 is the determined cDNA sequence for clone R0199:G11
15 SEQ ID NO:152 is the determined cDNA sequence for clone R0199:G12
SEQ ID NO:153 is the determined cDNA sequence for clone R0199:H02
SEQ ID NO:154 is the determined cDNA sequence for clone R0199:H03
SEQ ID NO:155 is the determined cDNA sequence for clone R0200:A05
SEQ ID NO:156 is the determined cDNA sequence for clone R0200:A06
20 SEQ ID NO:157 is the determined cDNA sequence for clone R0200:A10
SEQ ID NO:158 is the determined cDNA sequence for clone R0200:A12
SEQ ID NO:159 is the determined cDNA sequence for clone R0200:B03
SEQ ID NO:160 is the determined cDNA sequence for clone R0200:B04
SEQ ID NO:161 is the determined cDNA sequence for clone R0200:B07
25 SEQ ID NO:162 is the determined cDNA sequence for clone R0200:B08
SEQ ID NO:163 is the determined cDNA sequence for clone R0200:B12
SEQ ID NO:164 is the determined cDNA sequence for clone R0200:C02
SEQ ID NO:165 is the determined cDNA sequence for clone R0200:C07
SEQ ID NO:166 is the determined cDNA sequence for clone R0200:C09
30 SEQ ID NO:167 is the determined cDNA sequence for clone R0200:C10
SEQ ID NO:168 is the determined cDNA sequence for clone R0200:D01
SEQ ID NO:169 is the determined cDNA sequence for clone R0200:D03

SEQ ID NO:170 is the determined cDNA sequence for clone R0200:D05
SEQ ID NO:171 is the determined cDNA sequence for clone R0200:D06
SEQ ID NO:172 is the determined cDNA sequence for clone R0200:D07
SEQ ID NO:173 is the determined cDNA sequence for clone R0200:D08
5 SEQ ID NO:174 is the determined cDNA sequence for clone R0200:D09
SEQ ID NO:175 is the determined cDNA sequence for clone R0200:D11
SEQ ID NO:176 is the determined cDNA sequence for clone R0200:D12
SEQ ID NO:177 is the determined cDNA sequence for clone R0200:E03
SEQ ID NO:178 is the determined cDNA sequence for clone R0200:E04
10 SEQ ID NO:179 is the determined cDNA sequence for clone R0200:E06
SEQ ID NO:180 is the determined cDNA sequence for clone R0200:E07
SEQ ID NO:181 is the determined cDNA sequence for clone R0200:E08
SEQ ID NO:182 is the determined cDNA sequence for clone R0200:E09
SEQ ID NO:183 is the determined cDNA sequence for clone R0200:E12
15 SEQ ID NO:184 is the determined cDNA sequence for clone R0200:F01
SEQ ID NO:185 is the determined cDNA sequence for clone R0200:F04
SEQ ID NO:186 is the determined cDNA sequence for clone R0200:F05
SEQ ID NO:187 is the determined cDNA sequence for clone R0200:F07
SEQ ID NO:188 is the determined cDNA sequence for clone R0200:F08
20 SEQ ID NO:189 is the determined cDNA sequence for clone R0200:F09
SEQ ID NO:190 is the determined cDNA sequence for clone R0200:F10
SEQ ID NO:191 is the determined cDNA sequence for clone R0200:F11
SEQ ID NO:192 is the determined cDNA sequence for clone R0200:F12
SEQ ID NO:193 is the determined cDNA sequence for clone R0200:G02
25 SEQ ID NO:194 is the determined cDNA sequence for clone R0200:G07
SEQ ID NO:195 is the determined cDNA sequence for clone R0200:G08
SEQ ID NO:196 is the determined cDNA sequence for clone R0200:G09
SEQ ID NO:197 is the determined cDNA sequence for clone R0200:G10
SEQ ID NO:198 is the determined cDNA sequence for clone R0200:G12
30 SEQ ID NO:199 is the determined cDNA sequence for clone R0200:H03
SEQ ID NO:200 is the determined cDNA sequence for clone R0200:H05
SEQ ID NO:201 is the determined cDNA sequence for clone R0200:H07

- SEQ ID NO:202 is the determined cDNA sequence for clone R0200:H09
SEQ ID NO:203 is the determined cDNA sequence for clone R0200:H11
SEQ ID NO:204 is the determined cDNA sequence for clone 57877.2
SEQ ID NO:205 is the determined cDNA sequence for clone 57879.3
5 SEQ ID NO:206 is the determined cDNA sequence for clone 57881.2
SEQ ID NO:207 is the determined cDNA sequence for clone 57882.1
SEQ ID NO:208 is the determined cDNA sequence for clone 57884.2
SEQ ID NO:209 is the determined cDNA sequence for clone 57888.2
SEQ ID NO:210 is an extended cDNA sequence for clone RO198 C12
10 (SEQ ID NO: 60), also referred to as O593S
SEQ ID NO:211 is an extended cDNA sequence for clone RO198 F2
(SEQ ID NO: 80), also referred to as O594S
SEQ ID NO:212 is an extended cDNA sequence for clone RO199 A7
(SEQ ID NO: 107), also referred to as O595S
15 SEQ ID NO:213 is an extended cDNA sequence for clone RO199 C12
(SEQ ID NO: 125), also referred to as O596S
SEQ ID NO:214 is a full length cDNA sequence for HSPCO67, a
sequence having homology with O596S
SEQ ID NO:215 is an extended cDNA sequence for clone RO200 A10
20 (SEQ ID NO: 157), also referred to as O597S
SEQ ID NO:216 is an extended cDNA sequence for clone RO200 A12
(SEQ ID NO: 158), also referred to as O598S
SEQ ID NO:217 is a full length cDNA sequence for monocarboxylate
transporter (MCT3), a sequence having homology with O598S
25 SEQ ID NO:218 is an extended cDNA sequence for clone RO200 E10
(57881.2; SEQ ID NO: 206), also referred to as O599S
SEQ ID NO:219 is an extended cDNA sequence for clone RO200 G2
(SEQ ID NO: 193), also referred to as O600S
SEQ ID NO:220 is an extended cDNA sequence for clone RO200 B4
30 (57882.1; SEQ ID NO: 207), also referred to as O601S
SEQ ID NO:221 is a full length cDNA sequence for lysophospholipase I
(LYPLA1), a sequence having homology with O601S

SEQ ID NO:222 is an extended cDNA sequence for clone RO201 D1 (57884.2; SEQ ID NO: 208), also referred to as O602S

DETAILED DESCRIPTION OF THE INVENTION

As noted above, the present invention is generally directed to compositions and methods for using the compositions, for example in the therapy and diagnosis of cancer, such as ovarian and endometrial cancer. Certain illustrative compositions described herein include ovarian tumor polypeptides, polynucleotides encoding such polypeptides, binding agents such as antibodies, antigen presenting cells (APCs) and/or immune system cells (*e.g.*, T cells). An "ovarian tumor protein," as the term is used herein, refers generally to a protein that is expressed in ovarian tumor cells at a level that is at least two fold, and preferably at least five fold, greater than the level of expression in a normal tissue, as determined using a representative assay provided herein. Certain ovarian tumor proteins are tumor proteins that react detectably (within an immunoassay, such as an ELISA or Western blot) with antisera of a patient afflicted with ovarian cancer.

Therefore, in accordance with the above, and as described further below, the present invention provides illustrative polynucleotide compositions having sequences set forth in SEQ ID NO:1-222, antibody compositions capable of binding polypeptides encoded by the polynucleotides, and numerous additional embodiments employing such compositions, for example in the detection, diagnosis and/or therapy of human ovarian cancer.

POLYNUCLEOTIDE COMPOSITIONS

As used herein, the terms "DNA segment" and "polynucleotide" refer to a DNA molecule that has been isolated free of total genomic DNA of a particular species. Therefore, a DNA segment encoding a polypeptide refers to a DNA segment that contains one or more coding sequences yet is substantially isolated away from, or purified free from, total genomic DNA of the species from which the DNA segment is obtained. Included within the terms "DNA segment" and "polynucleotide" are DNA segments and smaller fragments of such segments, and also recombinant vectors, including, for example, plasmids, cosmids, phagemids, phage, viruses, and the like.

As will be understood by those skilled in the art, the DNA segments of this invention can include genomic sequences, extra-genomic and plasmid-encoded sequences and smaller engineered gene segments that express, or may be adapted to express, proteins, polypeptides, peptides and the like. Such segments may be naturally
5 isolated, or modified synthetically by the hand of man.

"Isolated," as used herein, means that a polynucleotide is substantially away from other coding sequences, and that the DNA segment does not contain large portions of unrelated coding DNA, such as large chromosomal fragments or other functional genes or polypeptide coding regions. Of course, this refers to the DNA
10 segment as originally isolated, and does not exclude genes or coding regions later added to the segment by the hand of man.

As will be recognized by the skilled artisan, polynucleotides may be single-stranded (coding or antisense) or double-stranded, and may be DNA (genomic, cDNA or synthetic) or RNA molecules. RNA molecules include HnRNA molecules,
15 which contain introns and correspond to a DNA molecule in a one-to-one manner, and mRNA molecules, which do not contain introns. Additional coding or non-coding sequences may, but need not, be present within a polynucleotide of the present invention, and a polynucleotide may, but need not, be linked to other molecules and/or support materials.

20 Polynucleotides may comprise a native sequence (*i.e.*, an endogenous sequence that encodes an ovarian tumor protein or a portion thereof) or may comprise a variant, or a biological or antigenic functional equivalent of such a sequence. Polynucleotide variants may contain one or more substitutions, additions, deletions and/or insertions, as further described below, preferably such that the immunogenicity
25 of the encoded polypeptide is not diminished, relative to a native tumor protein. The effect on the immunogenicity of the encoded polypeptide may generally be assessed as described herein. The term "variants" also encompasses homologous genes of xenogenic origin.

When comparing polynucleotide or polypeptide sequences, two
30 sequences are said to be "identical" if the sequence of nucleotides or amino acids in the two sequences is the same when aligned for maximum correspondence, as described below. Comparisons between two sequences are typically performed by comparing the

sequences over a comparison window to identify and compare local regions of sequence similarity. A "comparison window" as used herein, refers to a segment of at least about 20 contiguous positions, usually 30 to about 75, 40 to about 50, in which a sequence may be compared to a reference sequence of the same number of contiguous positions
5 after the two sequences are optimally aligned.

Optimal alignment of sequences for comparison may be conducted using the Megalign program in the Lasergene suite of bioinformatics software (DNASTAR, Inc., Madison, WI), using default parameters. This program embodies several alignment schemes described in the following references: Dayhoff, M.O. (1978) A
10 model of evolutionary change in proteins – Matrices for detecting distant relationships. In Dayhoff, M.O. (ed.) *Atlas of Protein Sequence and Structure*, National Biomedical Research Foundation, Washington DC Vol. 5, Suppl. 3, pp. 345-358; Hein J. (1990) *Unified Approach to Alignment and Phylogenies* pp. 626-645 *Methods in Enzymology* vol. 183, Academic Press, Inc., San Diego, CA; Higgins, D.G. and Sharp, P.M. (1989)
15 *CABIOS* 5:151-153; Myers, E.W. and Muller W. (1988) *CABIOS* 4:11-17; Robinson, E.D. (1971) *Comb. Theor* 11:105; Santou, N. Nes, M. (1987) *Mol. Biol. Evol.* 4:406-425; Sneath, P.H.A. and Sokal, R.R. (1973) *Numerical Taxonomy – the Principles and Practice of Numerical Taxonomy*, Freeman Press, San Francisco, CA; Wilbur, W.J. and Lipman, D.J. (1983) *Proc. Natl. Acad. Sci. USA* 80:726-730.

20 Alternatively, optimal alignment of sequences for comparison may be conducted by the local identity algorithm of Smith and Waterman (1981) *Add. APL. Math* 2:482, by the identity alignment algorithm of Needleman and Wunsch (1970) *J. Mol. Biol.* 48:443, by the search for similarity methods of Pearson and Lipman (1988) *Proc. Natl. Acad. Sci. USA* 85: 2444, by computerized implementations of these
25 algorithms (GAP, BESTFIT, BLAST, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group (GCG), 575 Science Dr., Madison, WI), or by inspection.

One preferred example of algorithms that are suitable for determining percent sequence identity and sequence similarity are the BLAST and BLAST 2.0
30 algorithms, which are described in Altschul *et al.* (1977) *Nucl. Acids Res.* 25:3389-3402 and Altschul *et al.* (1990) *J. Mol. Biol.* 215:403-410, respectively. BLAST and BLAST 2.0 can be used, for example with the parameters described herein, to determine percent

sequence identity for the polynucleotides and polypeptides of the invention. Software for performing BLAST analyses is publicly available through the National Center for Biotechnology Information. In one illustrative example, cumulative scores can be calculated using, for nucleotide sequences, the parameters M (reward score for a pair of
5 matching residues; always >0) and N (penalty score for mismatching residues; always <0). For amino acid sequences, a scoring matrix can be used to calculate the cumulative score. Extension of the word hits in each direction are halted when: the cumulative alignment score falls off by the quantity X from its maximum achieved value; the cumulative score goes to zero or below, due to the accumulation of one or
10 more negative-scoring residue alignments; or the end of either sequence is reached. The BLAST algorithm parameters W, T and X determine the sensitivity and speed of the alignment. The BLASTN program (for nucleotide sequences) uses as defaults a wordlength (W) of 11, and expectation (E) of 10, and the BLOSUM62 scoring matrix (see Henikoff and Henikoff (1989) *Proc. Natl. Acad. Sci. USA* 89:10915) alignments,
15 (B) of 50, expectation (E) of 10, M=5, N=-4 and a comparison of both strands.

Preferably, the "percentage of sequence identity" is determined by comparing two optimally aligned sequences over a window of comparison of at least 20 positions, wherein the portion of the polynucleotide or polypeptide sequence in the comparison window may comprise additions or deletions (*i.e.*, gaps) of 20 percent or
20 less, usually 5 to 15 percent, or 10 to 12 percent, as compared to the reference sequences (which does not comprise additions or deletions) for optimal alignment of the two sequences. The percentage is calculated by determining the number of positions at which the identical nucleic acid bases or amino acid residue occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the
25 total number of positions in the reference sequence (*i.e.*, the window size) and multiplying the results by 100 to yield the percentage of sequence identity.

Therefore, the present invention encompasses polynucleotide and polypeptide sequences having substantial identity to the sequences disclosed herein, for example those comprising at least 50% sequence identity, preferably at least 55%, 60%,
30 65%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% or higher, sequence identity compared to a polynucleotide or polypeptide sequence of this invention using the methods described herein, (*e.g.*, BLAST analysis using standard parameters, as

described below). One skilled in this art will recognize that these values can be appropriately adjusted to determine corresponding identity of proteins encoded by two nucleotide sequences by taking into account codon degeneracy, amino acid similarity, reading frame positioning and the like.

5 In additional embodiments, the present invention provides isolated polynucleotides and polypeptides comprising various lengths of contiguous stretches of sequence identical to or complementary to one or more of the sequences disclosed herein. For example, polynucleotides are provided by this invention that comprise at least about 15, 20, 30, 40, 50, 75, 100, 150, 200, 300, 400, 500 or 1000 or more
10 contiguous nucleotides of one or more of the sequences disclosed herein as well as all intermediate lengths there between. It will be readily understood that "intermediate lengths", in this context, means any length between the quoted values, such as 16, 17, 18, 19, *etc.*; 21, 22, 23, *etc.*; 30, 31, 32, *etc.*; 50, 51, 52, 53, *etc.*; 100, 101, 102, 103, *etc.*; 150, 151, 152, 153, *etc.*; including all integers through 200-500; 500-1,000, and the
15 like.

The polynucleotides of the present invention, or fragments thereof, regardless of the length of the coding sequence itself, may be combined with other DNA sequences, such as promoters, polyadenylation signals, additional restriction enzyme sites, multiple cloning sites, other coding segments, and the like, such that their
20 overall length may vary considerably. It is therefore contemplated that a nucleic acid fragment of almost any length may be employed, with the total length preferably being limited by the ease of preparation and use in the intended recombinant DNA protocol. For example, illustrative DNA segments with total lengths of about 10,000, about 5000, about 3000, about 2,000, about 1,000, about 500, about 200, about 100, about 50 base
25 pairs in length, and the like, (including all intermediate lengths) are contemplated to be useful in many implementations of this invention.

In other embodiments, the present invention is directed to polynucleotides that are capable of hybridizing under moderately stringent conditions to a polynucleotide sequence provided herein, or a fragment thereof, or a complementary
30 sequence thereof. Hybridization techniques are well known in the art of molecular biology. For purposes of illustration, suitable moderately stringent conditions for testing the hybridization of a polynucleotide of this invention with other

polynucleotides include prewashing in a solution of 5 X SSC, 0.5% SDS, 1.0 mM EDTA (pH 8.0); hybridizing at 50°C-65°C, 5 X SSC, overnight; followed by washing twice at 65°C for 20 minutes with each of 2X, 0.5X and 0.2X SSC containing 0.1% SDS.

5 Moreover, it will be appreciated by those of ordinary skill in the art that, as a result of the degeneracy of the genetic code, there are many nucleotide sequences that encode a polypeptide as described herein. Some of these polynucleotides bear minimal homology to the nucleotide sequence of any native gene. Nonetheless, polynucleotides that vary due to differences in codon usage are specifically
10 contemplated by the present invention. Further, alleles of the genes comprising the polynucleotide sequences provided herein are within the scope of the present invention. Alleles are endogenous genes that are altered as a result of one or more mutations, such as deletions, additions and/or substitutions of nucleotides. The resulting mRNA and protein may, but need not, have an altered structure or function. Alleles may be
15 identified using standard techniques (such as hybridization, amplification and/or database sequence comparison).

PROBES AND PRIMERS

In other embodiments of the present invention, the polynucleotide sequences provided herein can be advantageously used as probes or primers for nucleic
20 acid hybridization. As such, it is contemplated that nucleic acid segments that comprise a sequence region of at least about 15 nucleotide long contiguous sequence that has the same sequence as, or is complementary to, a 15 nucleotide long contiguous sequence disclosed herein will find particular utility. Longer contiguous identical or complementary sequences, *e.g.*, those of about 20, 30, 40, 50, 100, 200, 500, 1000
25 (including all intermediate lengths) and even up to full length sequences will also be of use in certain embodiments.

The ability of such nucleic acid probes to specifically hybridize to a sequence of interest will enable them to be of use in detecting the presence of complementary sequences in a given sample. However, other uses are also envisioned,
30 such as the use of the sequence information for the preparation of mutant species primers, or primers for use in preparing other genetic constructions.

Polynucleotide molecules having sequence regions consisting of contiguous nucleotide stretches of 10-14, 15-20, 30, 50, or even of 100-200 nucleotides or so (including intermediate lengths as well), identical or complementary to a polynucleotide sequence disclosed herein, are particularly contemplated as
5 hybridization probes for use in, *e.g.*, Southern and Northern blotting. This would allow a gene product, or fragment thereof, to be analyzed, both in diverse cell types and also in various bacterial cells. The total size of fragment, as well as the size of the complementary stretch(es), will ultimately depend on the intended use or application of the particular nucleic acid segment. Smaller fragments will generally find use in
10 hybridization embodiments, wherein the length of the contiguous complementary region may be varied, such as between about 15 and about 100 nucleotides, but larger contiguous complementarity stretches may be used, according to the length complementary sequences one wishes to detect.

The use of a hybridization probe of about 15-25 nucleotides in length
15 allows the formation of a duplex molecule that is both stable and selective. Molecules having contiguous complementary sequences over stretches greater than 15 bases in length are generally preferred, though, in order to increase stability and selectivity of the hybrid, and thereby improve the quality and degree of specific hybrid molecules obtained. One will generally prefer to design nucleic acid molecules having gene-
20 complementary stretches of 15 to 25 contiguous nucleotides, or even longer where desired.

Hybridization probes may be selected from any portion of any of the sequences disclosed herein. All that is required is to review the sequence set forth in SEQ ID NO:1-222, or to any continuous portion of the sequence, from about 15-25
25 nucleotides in length up to and including the full length sequence, that one wishes to utilize as a probe or primer. The choice of probe and primer sequences may be governed by various factors. For example, one may wish to employ primers from towards the termini of the total sequence.

Small polynucleotide segments or fragments may be readily prepared by,
30 for example, directly synthesizing the fragment by chemical means, as is commonly practiced using an automated oligonucleotide synthesizer. Also, fragments may be obtained by application of nucleic acid reproduction technology, such as the PCR™

technology of U. S. Patent 4,683,202 (incorporated herein by reference), by introducing selected sequences into recombinant vectors for recombinant production, and by other recombinant DNA techniques generally known to those of skill in the art of molecular biology.

5 The nucleotide sequences of the invention may be used for their ability to selectively form duplex molecules with complementary stretches of the entire gene or gene fragments of interest. Depending on the application envisioned, one will typically desire to employ varying conditions of hybridization to achieve varying degrees of selectivity of probe towards target sequence. For applications requiring high
10 selectivity, one will typically desire to employ relatively stringent conditions to form the hybrids, *e.g.*, one will select relatively low salt and/or high temperature conditions, such as provided by a salt concentration of from about 0.02 M to about 0.15 M salt at temperatures of from about 50°C to about 70°C. Such selective conditions tolerate little, if any, mismatch between the probe and the template or target strand, and would
15 be particularly suitable for isolating related sequences.

Of course, for some applications, for example, where one desires to prepare mutants employing a mutant primer strand hybridized to an underlying template, less stringent (reduced stringency) hybridization conditions will typically be needed in order to allow formation of the heteroduplex. In these circumstances, one
20 may desire to employ salt conditions such as those of from about 0.15 M to about 0.9 M salt, at temperatures ranging from about 20°C to about 55°C. Cross-hybridizing species can thereby be readily identified as positively hybridizing signals with respect to control hybridizations. In any case, it is generally appreciated that conditions can be rendered more stringent by the addition of increasing amounts of formamide, which serves to
25 destabilize the hybrid duplex in the same manner as increased temperature. Thus, hybridization conditions can be readily manipulated, and thus will generally be a method of choice depending on the desired results.

POLYNUCLEOTIDE IDENTIFICATION AND CHARACTERIZATION

Polynucleotides may be identified, prepared and/or manipulated using
30 any of a variety of well established techniques. For example, a polynucleotide may be identified, as described in more detail below, by screening a microarray of cDNAs for

tumor-associated expression (*i.e.*, expression that is at least two fold greater in a tumor than in normal tissue, as determined using a representative assay provided herein). Such screens may be performed, for example, using a Synteni microarray (Palo Alto, CA) according to the manufacturer's instructions (and essentially as described by Schena *et al.*, *Proc. Natl. Acad. Sci. USA* 93:10614-10619, 1996 and Heller *et al.*, *Proc. Natl. Acad. Sci. USA* 94:2150-2155, 1997). Alternatively, polynucleotides may be amplified from cDNA prepared from cells expressing the proteins described herein, such as ovarian tumor cells. Such polynucleotides may be amplified via polymerase chain reaction (PCR). For this approach, sequence-specific primers may be designed based on the sequences provided herein, and may be purchased or synthesized.

An amplified portion of a polynucleotide of the present invention may be used to isolate a full length gene from a suitable library (*e.g.*, an ovarian tumor cDNA library) using well known techniques. Within such techniques, a library (cDNA or genomic) is screened using one or more polynucleotide probes or primers suitable for amplification. Preferably, a library is size-selected to include larger molecules. Random primed libraries may also be preferred for identifying 5' and upstream regions of genes. Genomic libraries are preferred for obtaining introns and extending 5' sequences.

For hybridization techniques, a partial sequence may be labeled (*e.g.*, by nick-translation or end-labeling with ^{32}P) using well known techniques. A bacterial or bacteriophage library is then generally screened by hybridizing filters containing denatured bacterial colonies (or lawns containing phage plaques) with the labeled probe (*see* Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratories, Cold Spring Harbor, NY, 1989). Hybridizing colonies or plaques are selected and expanded, and the DNA is isolated for further analysis. cDNA clones may be analyzed to determine the amount of additional sequence by, for example, PCR using a primer from the partial sequence and a primer from the vector. Restriction maps and partial sequences may be generated to identify one or more overlapping clones. The complete sequence may then be determined using standard techniques, which may involve generating a series of deletion clones. The resulting overlapping sequences can then be assembled into a single contiguous sequence. A full length cDNA molecule can be generated by ligating suitable fragments, using well known techniques.

Alternatively, there are numerous amplification techniques for obtaining a full length coding sequence from a partial cDNA sequence. Within such techniques, amplification is generally performed via PCR. Any of a variety of commercially available kits may be used to perform the amplification step. Primers may be designed
5 using, for example, software well known in the art. Primers are preferably 22-30 nucleotides in length, have a GC content of at least 50% and anneal to the target sequence at temperatures of about 68°C to 72°C. The amplified region may be sequenced as described above, and overlapping sequences assembled into a contiguous sequence.

10 One such amplification technique is inverse PCR (see Triglia *et al.*, *Nucl. Acids Res.* 16:8186, 1988), which uses restriction enzymes to generate a fragment in the known region of the gene. The fragment is then circularized by intramolecular ligation and used as a template for PCR with divergent primers derived from the known region. Within an alternative approach, sequences adjacent to a partial sequence may
15 be retrieved by amplification with a primer to a linker sequence and a primer specific to a known region. The amplified sequences are typically subjected to a second round of amplification with the same linker primer and a second primer specific to the known region. A variation on this procedure, which employs two primers that initiate extension in opposite directions from the known sequence, is described in WO
20 96/38591. Another such technique is known as "rapid amplification of cDNA ends" or RACE. This technique involves the use of an internal primer and an external primer, which hybridizes to a polyA region or vector sequence, to identify sequences that are 5' and 3' of a known sequence. Additional techniques include capture PCR (Lagerstrom *et al.*, *PCR Methods Applic.* 1:111-19, 1991) and walking PCR (Parker *et al.*, *Nucl. Acids.*
25 *Res.* 19:3055-60, 1991). Other methods employing amplification may also be employed to obtain a full length cDNA sequence.

In certain instances, it is possible to obtain a full length cDNA sequence by analysis of sequences provided in an expressed sequence tag (EST) database, such as that available from GenBank. Searches for overlapping ESTs may generally be
30 performed using well known programs (*e.g.*, NCBI BLAST searches), and such ESTs may be used to generate a contiguous full length sequence. Full length DNA sequences may also be obtained by analysis of genomic fragments.

POLYNUCLEOTIDE EXPRESSION IN HOST CELLS

In other embodiments of the invention, polynucleotide sequences or fragments thereof which encode polypeptides of the invention, or fusion proteins or functional equivalents thereof, may be used in recombinant DNA molecules to direct
5 expression of a polypeptide in appropriate host cells. Due to the inherent degeneracy of the genetic code, other DNA sequences that encode substantially the same or a functionally equivalent amino acid sequence may be produced and these sequences may be used to clone and express a given polypeptide.

As will be understood by those of skill in the art, it may be advantageous
10 in some instances to produce polypeptide-encoding nucleotide sequences possessing non-naturally occurring codons. For example, codons preferred by a particular prokaryotic or eukaryotic host can be selected to increase the rate of protein expression or to produce a recombinant RNA transcript having desirable properties, such as a half-life which is longer than that of a transcript generated from the naturally occurring
15 sequence.

Moreover, the polynucleotide sequences of the present invention can be engineered using methods generally known in the art in order to alter polypeptide encoding sequences for a variety of reasons, including but not limited to, alterations which modify the cloning, processing, and/or expression of the gene product. For
20 example, DNA shuffling by random fragmentation and PCR reassembly of gene fragments and synthetic oligonucleotides may be used to engineer the nucleotide sequences. In addition, site-directed mutagenesis may be used to insert new restriction sites, alter glycosylation patterns, change codon preference, produce splice variants, or introduce mutations, and so forth.

25 In another embodiment of the invention, natural, modified, or recombinant nucleic acid sequences may be ligated to a heterologous sequence to encode a fusion protein. For example, to screen peptide libraries for inhibitors of polypeptide activity, it may be useful to encode a chimeric protein that can be recognized by a commercially available antibody. A fusion protein may also be
30 engineered to contain a cleavage site located between the polypeptide-encoding sequence and the heterologous protein sequence, so that the polypeptide may be cleaved and purified away from the heterologous moiety.

Sequences encoding a desired polypeptide may be synthesized, in whole or in part, using chemical methods well known in the art (see Caruthers, M. H. *et al.* (1980) *Nucl. Acids Res. Symp. Ser.* 215-223, Horn, T. *et al.* (1980) *Nucl. Acids Res. Symp. Ser.* 225-232). Alternatively, the protein itself may be produced using chemical
5 methods to synthesize the amino acid sequence of a polypeptide, or a portion thereof. For example, peptide synthesis can be performed using various solid-phase techniques (Roberge, J. Y. *et al.* (1995) *Science* 269:202-204) and automated synthesis may be achieved, for example, using the ABI 431A Peptide Synthesizer (Perkin Elmer, Palo Alto, CA).

10 A newly synthesized peptide may be substantially purified by preparative high performance liquid chromatography (*e.g.*, Creighton, T. (1983) *Proteins, Structures and Molecular Principles*, WH Freeman and Co., New York, N.Y.) or other comparable techniques available in the art. The composition of the synthetic peptides may be confirmed by amino acid analysis or sequencing (*e.g.*, the Edman
15 degradation procedure). Additionally, the amino acid sequence of a polypeptide, or any part thereof, may be altered during direct synthesis and/or combined using chemical methods with sequences from other proteins, or any part thereof, to produce a variant polypeptide.

In order to express a desired polypeptide, the nucleotide sequences
20 encoding the polypeptide, or functional equivalents, may be inserted into appropriate expression vector, *i.e.*, a vector which contains the necessary elements for the transcription and translation of the inserted coding sequence. Methods which are well known to those skilled in the art may be used to construct expression vectors containing sequences encoding a polypeptide of interest and appropriate transcriptional and
25 translational control elements. These methods include in vitro recombinant DNA techniques, synthetic techniques, and in vivo genetic recombination. Such techniques are described in Sambrook, J. *et al.* (1989) *Molecular Cloning, A Laboratory Manual*, Cold Spring Harbor Press, Plainview, N.Y., and Ausubel, F. M. *et al.* (1989) *Current Protocols in Molecular Biology*, John Wiley & Sons, New York, N.Y.

30 A variety of expression vector/host systems may be utilized to contain and express polynucleotide sequences. These include, but are not limited to, microorganisms such as bacteria transformed with recombinant bacteriophage, plasmid,

or cosmid DNA expression vectors; yeast transformed with yeast expression vectors; insect cell systems infected with virus expression vectors (e.g., baculovirus); plant cell systems transformed with virus expression vectors (e.g., cauliflower mosaic virus, CaMV; tobacco mosaic virus, TMV) or with bacterial expression vectors (e.g., Ti or pBR322 plasmids); or animal cell systems.

The "control elements" or "regulatory sequences" present in an expression vector are those non-translated regions of the vector--enhancers, promoters, 5' and 3' untranslated regions--which interact with host cellular proteins to carry out transcription and translation. Such elements may vary in their strength and specificity. Depending on the vector system and host utilized, any number of suitable transcription and translation elements, including constitutive and inducible promoters, may be used. For example, when cloning in bacterial systems, inducible promoters such as the hybrid lacZ promoter of the PBLUESCRIPT phagemid (Stratagene, La Jolla, Calif.) or PSFORT1 plasmid (Gibco BRL, Gaithersburg, MD) and the like may be used. In mammalian cell systems, promoters from mammalian genes or from mammalian viruses are generally preferred. If it is necessary to generate a cell line that contains multiple copies of the sequence encoding a polypeptide, vectors based on SV40 or EBV may be advantageously used with an appropriate selectable marker.

In bacterial systems, a number of expression vectors may be selected depending upon the use intended for the expressed polypeptide. For example, when large quantities are needed, for example for the induction of antibodies, vectors which direct high level expression of fusion proteins that are readily purified may be used. Such vectors include, but are not limited to, the multifunctional *E. coli* cloning and expression vectors such as BLUESCRIPT (Stratagene), in which the sequence encoding the polypeptide of interest may be ligated into the vector in frame with sequences for the amino-terminal Met and the subsequent 7 residues of β -galactosidase so that a hybrid protein is produced; pIN vectors (Van Heeke, G. and S. M. Schuster (1989) *J. Biol. Chem.* 264:5503-5509); and the like. pGEX Vectors (Promega, Madison, Wis.) may also be used to express foreign polypeptides as fusion proteins with glutathione S-transferase (GST). In general, such fusion proteins are soluble and can easily be purified from lysed cells by adsorption to glutathione-agarose beads followed by elution in the presence of free glutathione. Proteins made in such systems may be designed to

include heparin, thrombin, or factor XA protease cleavage sites so that the cloned polypeptide of interest can be released from the GST moiety at will.

In the yeast, *Saccharomyces cerevisiae*, a number of vectors containing constitutive or inducible promoters such as alpha factor, alcohol oxidase, and PGH may be used. For reviews, see Ausubel *et al.* (supra) and Grant *et al.* (1987) *Methods Enzymol.* 153:516-544.

In cases where plant expression vectors are used, the expression of sequences encoding polypeptides may be driven by any of a number of promoters. For example, viral promoters such as the 35S and 19S promoters of CaMV may be used alone or in combination with the omega leader sequence from TMV (Takamatsu, N. (1987) *EMBO J.* 6:307-311. Alternatively, plant promoters such as the small subunit of RUBISCO or heat shock promoters may be used (Coruzzi, G. *et al.* (1984) *EMBO J.* 3:1671-1680; Broglie, R. *et al.* (1984) *Science* 224:838-843; and Winter, J. *et al.* (1991) *Results Probl. Cell Differ.* 17:85-105). These constructs can be introduced into plant cells by direct DNA transformation or pathogen-mediated transfection. Such techniques are described in a number of generally available reviews (see, for example, Hobbs, S. or Murry, L. E. in McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York, N.Y.; pp. 191-196).

An insect system may also be used to express a polypeptide of interest. For example, in one such system, *Autographa californica* nuclear polyhedrosis virus (AcNPV) is used as a vector to express foreign genes in *Spodoptera frugiperda* cells or in *Trichoplusia* larvae. The sequences encoding the polypeptide may be cloned into a non-essential region of the virus, such as the polyhedrin gene, and placed under control of the polyhedrin promoter. Successful insertion of the polypeptide-encoding sequence will render the polyhedrin gene inactive and produce recombinant virus lacking coat protein. The recombinant viruses may then be used to infect, for example, *S. frugiperda* cells or *Trichoplusia* larvae in which the polypeptide of interest may be expressed (Engelhard, E. K. *et al.* (1994) *Proc. Natl. Acad. Sci.* 91 :3224-3227).

In mammalian host cells, a number of viral-based expression systems are generally available. For example, in cases where an adenovirus is used as an expression vector, sequences encoding a polypeptide of interest may be ligated into an adenovirus transcription/translation complex consisting of the late promoter and tripartite leader

sequence. Insertion in a non-essential E1 or E3 region of the viral genome may be used to obtain a viable virus which is capable of expressing the polypeptide in infected host cells (Logan, J. and Shenk, T. (1984) *Proc. Natl. Acad. Sci.* 81:3655-3659). In addition, transcription enhancers, such as the Rous sarcoma virus (RSV) enhancer, may be used
5 to increase expression in mammalian host cells.

Specific initiation signals may also be used to achieve more efficient translation of sequences encoding a polypeptide of interest. Such signals include the ATG initiation codon and adjacent sequences. In cases where sequences encoding the polypeptide, its initiation codon, and upstream sequences are inserted into the
10 appropriate expression vector, no additional transcriptional or translational control signals may be needed. However, in cases where only coding sequence, or a portion thereof, is inserted, exogenous translational control signals including the ATG initiation codon should be provided. Furthermore, the initiation codon should be in the correct reading frame to ensure translation of the entire insert. Exogenous translational
15 elements and initiation codons may be of various origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of enhancers which are appropriate for the particular cell system which is used, such as those described in the literature (Scharf, D. *et al.* (1994) *Results Probl. Cell Differ.* 20:125-162).

In addition, a host cell strain may be chosen for its ability to modulate
20 the expression of the inserted sequences or to process the expressed protein in the desired fashion. Such modifications of the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" form of the protein may also be used to facilitate correct insertion, folding and/or function. Different host cells such as
25 CHO, HeLa, MDCK, HEK293, and WI38, which have specific cellular machinery and characteristic mechanisms for such post-translational activities, may be chosen to ensure the correct modification and processing of the foreign protein.

For long-term, high-yield production of recombinant proteins, stable expression is generally preferred. For example, cell lines which stably express a
30 polynucleotide of interest may be transformed using expression vectors which may contain viral origins of replication and/or endogenous expression elements and a selectable marker gene on the same or on a separate vector. Following the introduction

of the vector, cells may be allowed to grow for 1-2 days in an enriched media before they are switched to selective media. The purpose of the selectable marker is to confer resistance to selection, and its presence allows growth and recovery of cells which successfully express the introduced sequences. Resistant clones of stably transformed
5 cells may be proliferated using tissue culture techniques appropriate to the cell type.

Any number of selection systems may be used to recover transformed cell lines. These include, but are not limited to, the herpes simplex virus thymidine kinase (Wigler, M. *et al.* (1977) *Cell* 11:223-32) and adenine phosphoribosyltransferase (Lowy, I. *et al.* (1990) *Cell* 22:817-23) genes which can be employed in tk.sup.- or
10 aprt.sup.- cells, respectively. Also, antimetabolite, antibiotic or herbicide resistance can be used as the basis for selection; for example, dhfr which confers resistance to methotrexate (Wigler, M. *et al.* (1980) *Proc. Natl. Acad. Sci.* 77:3567-70); npt, which confers resistance to the aminoglycosides, neomycin and G-418 (Colbere-Garapin, F. *et al.* (1981) *J. Mol. Biol.* 150:1-14); and als or pat, which confer resistance to
15 chlorsulfuron and phosphinotricin acetyltransferase, respectively (Murry, *supra*). Additional selectable genes have been described, for example, trpB, which allows cells to utilize indole in place of tryptophan, or hisD, which allows cells to utilize histinol in place of histidine (Hartman, S. C. and R. C. Mulligan (1988) *Proc. Natl. Acad. Sci.* 85:8047-51). Recently, the use of visible markers has gained popularity with such
20 markers as anthocyanins, beta-glucuronidase and its substrate GUS, and luciferase and its substrate luciferin, being widely used not only to identify transformants, but also to quantify the amount of transient or stable protein expression attributable to a specific vector system (Rhodes, C. A. *et al.* (1995) *Methods Mol. Biol.* 55:121-131).

Although the presence/absence of marker gene expression suggests that
25 the gene of interest is also present, its presence and expression may need to be confirmed. For example, if the sequence encoding a polypeptide is inserted within a marker gene sequence, recombinant cells containing sequences can be identified by the absence of marker gene function. Alternatively, a marker gene can be placed in tandem with a polypeptide-encoding sequence under the control of a single promoter.
30 Expression of the marker gene in response to induction or selection usually indicates expression of the tandem gene as well.

Alternatively, host cells which contain and express a desired polynucleotide sequence may be identified by a variety of procedures known to those of skill in the art. These procedures include, but are not limited to, DNA-DNA or DNA-RNA hybridizations and protein bioassay or immunoassay techniques which include
5 membrane, solution, or chip based technologies for the detection and/or quantification of nucleic acid or protein.

A variety of protocols for detecting and measuring the expression of polynucleotide-encoded products, using either polyclonal or monoclonal antibodies specific for the product are known in the art. Examples include enzyme-linked
10 immunosorbent assay (ELISA), radioimmunoassay (RIA), and fluorescence activated cell sorting (FACS). A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering epitopes on a given polypeptide may be preferred for some applications, but a competitive binding assay may also be employed. These and other assays are described, among other places, in Hampton, R. *et al.* (1990;
15 Serological Methods, a Laboratory Manual, APS Press, St Paul, Minn.) and Maddox, D. E. *et al.* (1983; *J. Exp. Med.* 158:1211-1216).

A wide variety of labels and conjugation techniques are known by those skilled in the art and may be used in various nucleic acid and amino acid assays. Means for producing labeled hybridization or PCR probes for detecting sequences related to
20 polynucleotides include oligolabeling, nick translation, end-labeling or PCR amplification using a labeled nucleotide. Alternatively, the sequences, or any portions thereof may be cloned into a vector for the production of an mRNA probe. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes in vitro by addition of an appropriate RNA polymerase such as T7, T3, or SP6
25 and labeled nucleotides. These procedures may be conducted using a variety of commercially available kits. Suitable reporter molecules or labels, which may be used include radionuclides, enzymes, fluorescent, chemiluminescent, or chromogenic agents as well as substrates, cofactors, inhibitors, magnetic particles, and the like.

Host cells transformed with a polynucleotide sequence of interest may be
30 cultured under conditions suitable for the expression and recovery of the protein from cell culture. The protein produced by a recombinant cell may be secreted or contained intracellularly depending on the sequence and/or the vector used. As will be understood

by those of skill in the art, expression vectors containing polynucleotides of the invention may be designed to contain signal sequences which direct secretion of the encoded polypeptide through a prokaryotic or eukaryotic cell membrane. Other recombinant constructions may be used to join sequences encoding a polypeptide of interest to nucleotide sequence encoding a polypeptide domain which will facilitate purification of soluble proteins. Such purification facilitating domains include, but are not limited to, metal chelating peptides such as histidine-tryptophan modules that allow purification on immobilized metals, protein A domains that allow purification on immobilized immunoglobulin, and the domain utilized in the FLAGS extension/affinity purification system (Immunex Corp., Seattle, Wash.). The inclusion of cleavable linker sequences such as those specific for Factor XA or enterokinase (Invitrogen, San Diego, Calif.) between the purification domain and the encoded polypeptide may be used to facilitate purification. One such expression vector provides for expression of a fusion protein containing a polypeptide of interest and a nucleic acid encoding 6 histidine residues preceding a thioredoxin or an enterokinase cleavage site. The histidine residues facilitate purification on IMIAC (immobilized metal ion affinity chromatography) as described in Porath, J. *et al.* (1992, *Prot. Exp. Purif.* 3:263-281) while the enterokinase cleavage site provides a means for purifying the desired polypeptide from the fusion protein. A discussion of vectors which contain fusion proteins is provided in Kroll, D. J. *et al.* (1993; *DNA Cell Biol.* 12:441-453).

In addition to recombinant production methods, polypeptides of the invention, and fragments thereof, may be produced by direct peptide synthesis using solid-phase techniques (Merrifield J. (1963) *J. Am. Chem. Soc.* 85:2149-2154). Protein synthesis may be performed using manual techniques or by automation. Automated synthesis may be achieved, for example, using Applied Biosystems 431A Peptide Synthesizer (Perkin Elmer). Alternatively, various fragments may be chemically synthesized separately and combined using chemical methods to produce the full length molecule.

SITE-SPECIFIC MUTAGENESIS

Site-specific mutagenesis is a technique useful in the preparation of individual peptides, or biologically functional equivalent polypeptides, through specific

mutagenesis of the underlying polynucleotides that encode them. The technique, well-known to those of skill in the art, further provides a ready ability to prepare and test sequence variants, for example, incorporating one or more of the foregoing considerations, by introducing one or more nucleotide sequence changes into the DNA.

- 5 Site-specific mutagenesis allows the production of mutants through the use of specific oligonucleotide sequences which encode the DNA sequence of the desired mutation, as well as a sufficient number of adjacent nucleotides, to provide a primer sequence of sufficient size and sequence complexity to form a stable duplex on both sides of the deletion junction being traversed. Mutations may be employed in a selected
- 10 polynucleotide sequence to improve, alter, decrease, modify, or otherwise change the properties of the polynucleotide itself, and/or alter the properties, activity, composition, stability, or primary sequence of the encoded polypeptide.

In certain embodiments of the present invention, the inventors contemplate the mutagenesis of the disclosed polynucleotide sequences to alter one or

15 more properties of the encoded polypeptide, such as the antigenicity of a polypeptide vaccine. The techniques of site-specific mutagenesis are well-known in the art, and are widely used to create variants of both polypeptides and polynucleotides. For example, site-specific mutagenesis is often used to alter a specific portion of a DNA molecule. In such embodiments, a primer comprising typically about 14 to about 25 nucleotides or so

20 in length is employed, with about 5 to about 10 residues on both sides of the junction of the sequence being altered.

As will be appreciated by those of skill in the art, site-specific mutagenesis techniques have often employed a phage vector that exists in both a single stranded and double stranded form. Typical vectors useful in site-directed mutagenesis

25 include vectors such as the M13 phage. These phage are readily commercially-available and their use is generally well-known to those skilled in the art. Double-stranded plasmids are also routinely employed in site directed mutagenesis that eliminates the step of transferring the gene of interest from a plasmid to a phage.

In general, site-directed mutagenesis in accordance herewith is

30 performed by first obtaining a single-stranded vector or melting apart of two strands of a double-stranded vector that includes within its sequence a DNA sequence that encodes the desired peptide. An oligonucleotide primer bearing the desired mutated

sequence is prepared, generally synthetically. This primer is then annealed with the single-stranded vector, and subjected to DNA polymerizing enzymes such as *E. coli* polymerase I Klenow fragment, in order to complete the synthesis of the mutation-bearing strand. Thus, a heteroduplex is formed wherein one strand encodes the original
5 non-mutated sequence and the second strand bears the desired mutation. This heteroduplex vector is then used to transform appropriate cells, such as *E. coli* cells, and clones are selected which include recombinant vectors bearing the mutated sequence arrangement.

The preparation of sequence variants of the selected peptide-encoding
10 DNA segments using site-directed mutagenesis provides a means of producing potentially useful species and is not meant to be limiting as there are other ways in which sequence variants of peptides and the DNA sequences encoding them may be obtained. For example, recombinant vectors encoding the desired peptide sequence may be treated with mutagenic agents, such as hydroxylamine, to obtain sequence
15 variants. Specific details regarding these methods and protocols are found in the teachings of Maloy *et al.*, 1994; Segal, 1976; Prokop and Bajpai, 1991; Kuby, 1994; and Maniatis *et al.*, 1982, each incorporated herein by reference, for that purpose.

As used herein, the term "oligonucleotide directed mutagenesis procedure" refers to template-dependent processes and vector-mediated propagation
20 which result in an increase in the concentration of a specific nucleic acid molecule relative to its initial concentration, or in an increase in the concentration of a detectable signal, such as amplification. As used herein, the term "oligonucleotide directed mutagenesis procedure" is intended to refer to a process that involves the template-dependent extension of a primer molecule. The term template dependent
25 process refers to nucleic acid synthesis of an RNA or a DNA molecule wherein the sequence of the newly synthesized strand of nucleic acid is dictated by the well-known rules of complementary base pairing (see, for example, Watson, 1987). Typically, vector mediated methodologies involve the introduction of the nucleic acid fragment into a DNA or RNA vector, the clonal amplification of the vector, and the recovery of
30 the amplified nucleic acid fragment. Examples of such methodologies are provided by U. S. Patent No. 4,237,224, specifically incorporated herein by reference in its entirety.

POLYNUCLEOTIDE AMPLIFICATION TECHNIQUES

A number of template dependent processes are available to amplify the target sequences of interest present in a sample. One of the best known amplification methods is the polymerase chain reaction (PCRTM) which is described in detail in U.S. Patent Nos. 4,683,195, 4,683,202 and 4,800,159, each of which is incorporated herein by reference in its entirety. Briefly, in PCRTM, two primer sequences are prepared which are complementary to regions on opposite complementary strands of the target sequence. An excess of deoxynucleoside triphosphates is added to a reaction mixture along with a DNA polymerase (*e.g.*, *Taq* polymerase). If the target sequence is present in a sample, the primers will bind to the target and the polymerase will cause the primers to be extended along the target sequence by adding on nucleotides. By raising and lowering the temperature of the reaction mixture, the extended primers will dissociate from the target to form reaction products, excess primers will bind to the target and to the reaction product and the process is repeated. Preferably reverse transcription and PCRTM amplification procedure may be performed in order to quantify the amount of mRNA amplified. Polymerase chain reaction methodologies are well known in the art.

Another method for amplification is the ligase chain reaction (referred to as LCR), disclosed in Eur. Pat. Appl. Publ. No. 320,308 (specifically incorporated herein by reference in its entirety). In LCR, two complementary probe pairs are prepared, and in the presence of the target sequence, each pair will bind to opposite complementary strands of the target such that they abut. In the presence of a ligase, the two probe pairs will link to form a single unit. By temperature cycling, as in PCRTM, bound ligated units dissociate from the target and then serve as "target sequences" for ligation of excess probe pairs. U.S. Patent No. 4,883,750, incorporated herein by reference in its entirety, describes an alternative method of amplification similar to LCR for binding probe pairs to a target sequence.

Qbeta Replicase, described in PCT Intl. Pat. Appl. Publ. No. PCT/US87/00880, incorporated herein by reference in its entirety, may also be used as still another amplification method in the present invention. In this method, a replicative sequence of RNA that has a region complementary to that of a target is added to a

sample in the presence of an RNA polymerase. The polymerase will copy the replicative sequence that can then be detected.

An isothermal amplification method, in which restriction endonucleases and ligases are used to achieve the amplification of target molecules that contain nucleotide 5'-[α -thio]triphosphates in one strand of a restriction site (Walker *et al.*, 1992, incorporated herein by reference in its entirety), may also be useful in the amplification of nucleic acids in the present invention.

Strand Displacement Amplification (SDA) is another method of carrying out isothermal amplification of nucleic acids which involves multiple rounds of strand displacement and synthesis, *i.e.* nick translation. A similar method, called Repair Chain Reaction (RCR) is another method of amplification which may be useful in the present invention and is involves annealing several probes throughout a region targeted for amplification, followed by a repair reaction in which only two of the four bases are present. The other two bases can be added as biotinylated derivatives for easy detection. A similar approach is used in SDA.

Sequences can also be detected using a cyclic probe reaction (CPR). In CPR, a probe having a 3' and 5' sequences of non-target DNA and an internal or "middle" sequence of the target protein specific RNA is hybridized to DNA which is present in a sample. Upon hybridization, the reaction is treated with RNaseH, and the products of the probe are identified as distinctive products by generating a signal that is released after digestion. The original template is annealed to another cycling probe and the reaction is repeated. Thus, CPR involves amplifying a signal generated by hybridization of a probe to a target gene specific expressed nucleic acid.

Still other amplification methods described in Great Britain Pat. Appl. No. 2 202 328, and in PCT Intl. Pat. Appl. Publ. No. PCT/US89/01025, each of which is incorporated herein by reference in its entirety, may be used in accordance with the present invention. In the former application, "modified" primers are used in a PCR-like, template and enzyme dependent synthesis. The primers may be modified by labeling with a capture moiety (*e.g.*, biotin) and/or a detector moiety (*e.g.*, enzyme). In the latter application, an excess of labeled probes is added to a sample. In the presence of the target sequence, the probe binds and is cleaved catalytically. After cleavage, the

target sequence is released intact to be bound by excess probe. Cleavage of the labeled probe signals the presence of the target sequence.

Other nucleic acid amplification procedures include transcription-based amplification systems (TAS) (Kwoh *et al.*, 1989; PCT Intl. Pat. Appl. Publ. No. WO 88/10315, incorporated herein by reference in its entirety), including nucleic acid sequence based amplification (NASBA) and 3SR. In NASBA, the nucleic acids can be prepared for amplification by standard phenol/chloroform extraction, heat denaturation of a sample, treatment with lysis buffer and minispin columns for isolation of DNA and RNA or guanidinium chloride extraction of RNA. These amplification techniques involve annealing a primer that has sequences specific to the target sequence. Following polymerization, DNA/RNA hybrids are digested with RNase H while double stranded DNA molecules are heat-denatured again. In either case the single stranded DNA is made fully double stranded by addition of second target-specific primer, followed by polymerization. The double stranded DNA molecules are then multiply transcribed by a polymerase such as T7 or SP6. In an isothermal cyclic reaction, the RNAs are reverse transcribed into DNA, and transcribed once again with a polymerase such as T7 or SP6. The resulting products, whether truncated or complete, indicate target-specific sequences.

Eur. Pat. Appl. Publ. No. 329,822, incorporated herein by reference in its entirety, disclose a nucleic acid amplification process involving cyclically synthesizing single-stranded RNA ("ssRNA"), ssDNA, and double-stranded DNA (dsDNA), which may be used in accordance with the present invention. The ssRNA is a first template for a first primer oligonucleotide, which is elongated by reverse transcriptase (RNA-dependent DNA polymerase). The RNA is then removed from resulting DNA:RNA duplex by the action of ribonuclease H (RNase H, an RNase specific for RNA in a duplex with either DNA or RNA). The resultant ssDNA is a second template for a second primer, which also includes the sequences of an RNA polymerase promoter (exemplified by T7 RNA polymerase) 5' to its homology to its template. This primer is then extended by DNA polymerase (exemplified by the large "Klenow" fragment of *E. coli* DNA polymerase I), resulting as a double-stranded DNA ("dsDNA") molecule, having a sequence identical to that of the original RNA between the primers and having additionally, at one end, a promoter sequence. This promoter

sequence can be used by the appropriate RNA polymerase to make many RNA copies of the DNA. These copies can then re-enter the cycle leading to very swift amplification. With proper choice of enzymes, this amplification can be done isothermally without addition of enzymes at each cycle. Because of the cyclical nature of this process, the starting sequence can be chosen to be in the form of either DNA or RNA.

PCT Intl. Pat. Appl. Publ. No. WO 89/06700, incorporated herein by reference in its entirety, disclose a nucleic acid sequence amplification scheme based on the hybridization of a promoter/primer sequence to a target single-stranded DNA ("ssDNA") followed by transcription of many RNA copies of the sequence. This scheme is not cyclic; *i.e.* new templates are not produced from the resultant RNA transcripts. Other amplification methods include "RACE" (Frohman, 1990), and "one-sided PCR" (Ohara, 1989) which are well-known to those of skill in the art.

Methods based on ligation of two (or more) oligonucleotides in the presence of nucleic acid having the sequence of the resulting "di-oligonucleotide", thereby amplifying the di-oligonucleotide (Wu and Dean, 1996, incorporated herein by reference in its entirety), may also be used in the amplification of DNA sequences of the present invention.

BIOLOGICAL FUNCTIONAL EQUIVALENTS

Modification and changes may be made in the structure of the polynucleotides and polypeptides of the present invention and still obtain a functional molecule that encodes a polypeptide with desirable characteristics. As mentioned above, it is often desirable to introduce one or more mutations into a specific polynucleotide sequence. In certain circumstances, the resulting encoded polypeptide sequence is altered by this mutation, or in other cases, the sequence of the polypeptide is unchanged by one or more mutations in the encoding polynucleotide.

When it is desirable to alter the amino acid sequence of a polypeptide to create an equivalent, or even an improved, second-generation molecule, the amino acid changes may be achieved by changing one or more of the codons of the encoding DNA sequence, according to Table 1.

For example, certain amino acids may be substituted for other amino acids in a protein structure without appreciable loss of interactive binding capacity with structures such as, for example, antigen-binding regions of antibodies or binding sites on substrate molecules. Since it is the interactive capacity and nature of a protein that defines that protein's biological functional activity, certain amino acid sequence substitutions can be made in a protein sequence, and, of course, its underlying DNA coding sequence, and nevertheless obtain a protein with like properties. It is thus contemplated by the inventors that various changes may be made in the peptide sequences of the disclosed compositions, or corresponding DNA sequences which encode said peptides without appreciable loss of their biological utility or activity.

Table 1

Amino Acids			Codons						
Alanine	Ala	A	GCA	GCC	GCG	GCU			
Cysteine	Cys	C	UGC	UGU					
Aspartic acid	Asp	D	GAC	GAU					
Glutamic acid	Glu	E	GAA	GAG					
Phenylalanine	Phe	F	UUC	UUU					
Glycine	Gly	G	GGA	GGC	GGG	GGU			
Histidine	His	H	CAC	CAU					
Isoleucine	Ile	I	AUA	AUC	AUU				
Lysine	Lys	K	AAA	AAG					
Leucine	Leu	L	UUA	UUG	CUA	CUC	CUG	CUU	
Methionine	Met	M	AUG						
Asparagine	Asn	N	AAC	AAU					
Proline	Pro	P	CCA	CCC	CCG	CCU			
Glutamine	Gln	Q	CAA	CAG					
Arginine	Arg	R	AGA	AGG	CGA	CGC	CGG	CGU	
Serine	Ser	S	AGC	AGU	UCA	UCC	UCG	UCU	
Threonine	Thr	T	ACA	ACC	ACG	ACU			
Valine	Val	V	GUA	GUC	GUG	GUU			
Tryptophan	Trp	W	UGG						
Tyrosine	Tyr	Y	UAC	UAU					

In making such changes, the hydropathic index of amino acids may be considered. The importance of the hydropathic amino acid index in conferring interactive biologic function on a protein is generally understood in the art (Kyte and Doolittle, 1982, incorporated herein by reference). It is accepted that the relative hydropathic character of the amino acid contributes to the secondary structure of the resultant protein, which in turn defines the interaction of the protein with other molecules, for example, enzymes, substrates, receptors, DNA, antibodies, antigens, and the like. Each amino acid has been assigned a hydropathic index on the basis of its hydrophobicity and charge characteristics (Kyte and Doolittle, 1982). These values are: isoleucine (+4.5); valine (+4.2); leucine (+3.8); phenylalanine (+2.8); cysteine/cystine (+2.5); methionine (+1.9); alanine (+1.8); glycine (−0.4); threonine (−0.7); serine (−0.8); tryptophan (−0.9); tyrosine (−1.3); proline (−1.6); histidine (−3.2); glutamate (−3.5); glutamine (−3.5); aspartate (−3.5); asparagine (−3.5); lysine (−3.9); and arginine (−4.5).

It is known in the art that certain amino acids may be substituted by other amino acids having a similar hydropathic index or score and still result in a protein with similar biological activity, *i.e.* still obtain a biological functionally equivalent protein. In making such changes, the substitution of amino acids whose hydropathic indices are within ± 2 is preferred, those within ± 1 are particularly preferred, and those within ± 0.5 are even more particularly preferred. It is also understood in the art that the substitution of like amino acids can be made effectively on the basis of hydrophilicity. U. S. Patent 4,554,101 (specifically incorporated herein by reference in its entirety), states that the greatest local average hydrophilicity of a protein, as governed by the hydrophilicity of its adjacent amino acids, correlates with a biological property of the protein.

As detailed in U. S. Patent 4,554,101, the following hydrophilicity values have been assigned to amino acid residues: arginine (+3.0); lysine (+3.0); aspartate (+3.0 \pm 1); glutamate (+3.0 \pm 1); serine (+0.3); asparagine (+0.2); glutamine (+0.2); glycine (0); threonine (−0.4); proline (−0.5 \pm 1); alanine (−0.5); histidine (−0.5); cysteine (−1.0); methionine (−1.3); valine (−1.5); leucine (−1.8); isoleucine (−1.8); tyrosine (−2.3); phenylalanine (−2.5); tryptophan (−3.4). It is understood that an amino

acid can be substituted for another having a similar hydrophilicity value and still obtain a biologically equivalent, and in particular, an immunologically equivalent protein. In such changes, the substitution of amino acids whose hydrophilicity values are within ± 2 is preferred, those within ± 1 are particularly preferred, and those within ± 0.5 are even more particularly preferred.

As outlined above, amino acid substitutions are generally therefore based on the relative similarity of the amino acid side-chain substituents, for example, their hydrophobicity, hydrophilicity, charge, size, and the like. Exemplary substitutions that take various of the foregoing characteristics into consideration are well known to those of skill in the art and include: arginine and lysine; glutamate and aspartate; serine and threonine; glutamine and asparagine; and valine, leucine and isoleucine.

In addition, any polynucleotide may be further modified to increase stability *in vivo*. Possible modifications include, but are not limited to, the addition of flanking sequences at the 5' and/or 3' ends; the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages in the backbone; and/or the inclusion of nontraditional bases such as inosine, queosine and wybutosine, as well as acetyl-methyl-, thio- and other modified forms of adenine, cytidine, guanine, thymine and uridine.

IN VIVO POLYNUCLEOTIDE DELIVERY TECHNIQUES

In additional embodiments, genetic constructs comprising one or more of the polynucleotides of the invention are introduced into cells *in vivo*. This may be achieved using any of a variety of well known approaches, several of which are outlined below for the purpose of illustration.

1. ADENOVIRUS

One of the preferred methods for *in vivo* delivery of one or more nucleic acid sequences involves the use of an adenovirus expression vector. "Adenovirus expression vector" is meant to include those constructs containing adenovirus sequences sufficient to (a) support packaging of the construct and (b) to express a polynucleotide that has been cloned therein in a sense or antisense orientation. Of

course, in the context of an antisense construct, expression does not require that the gene product be synthesized.

The expression vector comprises a genetically engineered form of an adenovirus. Knowledge of the genetic organization of adenovirus, a 36 kb, linear, double-stranded DNA virus, allows substitution of large pieces of adenoviral DNA with foreign sequences up to 7 kb (Grunhaus and Horwitz, 1992). In contrast to retrovirus, the adenoviral infection of host cells does not result in chromosomal integration because adenoviral DNA can replicate in an episomal manner without potential genotoxicity. Also, adenoviruses are structurally stable, and no genome rearrangement has been detected after extensive amplification. Adenovirus can infect virtually all epithelial cells regardless of their cell cycle stage. So far, adenoviral infection appears to be linked only to mild disease such as acute respiratory disease in humans.

Adenovirus is particularly suitable for use as a gene transfer vector because of its mid-sized genome, ease of manipulation, high titer, wide target-cell range and high infectivity. Both ends of the viral genome contain 100-200 base pair inverted repeats (ITRs), which are *cis* elements necessary for viral DNA replication and packaging. The early (E) and late (L) regions of the genome contain different transcription units that are divided by the onset of viral DNA replication. The E1 region (E1A and E1B) encodes proteins responsible for the regulation of transcription of the viral genome and a few cellular genes. The expression of the E2 region (E2A and E2B) results in the synthesis of the proteins for viral DNA replication. These proteins are involved in DNA replication, late gene expression and host cell shut-off (Renan, 1990). The products of the late genes, including the majority of the viral capsid proteins, are expressed only after significant processing of a single primary transcript issued by the major late promoter (MLP). The MLP, (located at 16.8 m.u.) is particularly efficient during the late phase of infection, and all the mRNA's issued from this promoter possess a 5'-tripartite leader (TPL) sequence which makes them preferred mRNA's for translation.

In a current system, recombinant adenovirus is generated from homologous recombination between shuttle vector and provirus vector. Due to the possible recombination between two proviral vectors, wild-type adenovirus may be

generated from this process. Therefore, it is critical to isolate a single clone of virus from an individual plaque and examine its genomic structure.

Generation and propagation of the current adenovirus vectors, which are replication deficient, depend on a unique helper cell line, designated 293, which was transformed from human embryonic kidney cells by Ad5 DNA fragments and constitutively expresses E1 proteins (Graham *et al.*, 1977). Since the E3 region is dispensable from the adenovirus genome (Jones and Shenk, 1978), the current adenovirus vectors, with the help of 293 cells, carry foreign DNA in either the E1, the D3 or both regions (Graham and Prevec, 1991). In nature, adenovirus can package approximately 105% of the wild-type genome (Ghosh-Choudhury *et al.*, 1987), providing capacity for about 2 extra kB of DNA. Combined with the approximately 5.5 kB of DNA that is replaceable in the E1 and E3 regions, the maximum capacity of the current adenovirus vector is under 7.5 kB, or about 15% of the total length of the vector. More than 80% of the adenovirus viral genome remains in the vector backbone and is the source of vector-borne cytotoxicity. Also, the replication deficiency of the E1-deleted virus is incomplete. For example, leakage of viral gene expression has been observed with the currently available vectors at high multiplicities of infection (MOI) (Mulligan, 1993).

Helper cell lines may be derived from human cells such as human embryonic kidney cells, muscle cells, hematopoietic cells or other human embryonic mesenchymal or epithelial cells. Alternatively, the helper cells may be derived from the cells of other mammalian species that are permissive for human adenovirus. Such cells include, *e.g.*, Vero cells or other monkey embryonic mesenchymal or epithelial cells. As stated above, the currently preferred helper cell line is 293.

Recently, Racher *et al.* (1995) disclosed improved methods for culturing 293 cells and propagating adenovirus. In one format, natural cell aggregates are grown by inoculating individual cells into 1 liter siliconized spinner flasks (Technique, Cambridge, UK) containing 100-200 ml of medium. Following stirring at 40 rpm, the cell viability is estimated with trypan blue. In another format, Fibra-Cel microcarriers (Bibby Sterlin, Stone, UK) (5 g/l) is employed as follows. A cell inoculum, resuspended in 5 ml of medium, is added to the carrier (50 ml) in a 250 ml Erlenmeyer flask and left stationary, with occasional agitation, for 1 to 4 h. The medium is then

replaced with 50 ml of fresh medium and shaking initiated. For virus production, cells are allowed to grow to about 80% confluence, after which time the medium is replaced (to 25% of the final volume) and adenovirus added at an MOI of 0.05. Cultures are left stationary overnight, following which the volume is increased to 100% and shaking
5 commenced for another 72 h.

Other than the requirement that the adenovirus vector be replication defective, or at least conditionally defective, the nature of the adenovirus vector is not believed to be crucial to the successful practice of the invention. The adenovirus may be of any of the 42 different known serotypes or subgroups A-F. Adenovirus type 5 of
10 subgroup C is the preferred starting material in order to obtain a conditional replication-defective adenovirus vector for use in the present invention, since Adenovirus type 5 is a human adenovirus about which a great deal of biochemical and genetic information is known, and it has historically been used for most constructions employing adenovirus as a vector.

15 As stated above, the typical vector according to the present invention is replication defective and will not have an adenovirus E1 region. Thus, it will be most convenient to introduce the polynucleotide encoding the gene of interest at the position from which the E1-coding sequences have been removed. However, the position of insertion of the construct within the adenovirus sequences is not critical to the
20 invention. The polynucleotide encoding the gene of interest may also be inserted in lieu of the deleted E3 region in E3 replacement vectors as described by Karlsson *et al.* (1986) or in the E4 region where a helper cell line or helper virus complements the E4 defect.

Adenovirus is easy to grow and manipulate and exhibits broad host
25 range *in vitro* and *in vivo*. This group of viruses can be obtained in high titers, *e.g.*, 10^9 - 10^{11} plaque-forming units per ml, and they are highly infective. The life cycle of adenovirus does not require integration into the host cell genome. The foreign genes delivered by adenovirus vectors are episomal and, therefore, have low genotoxicity to host cells. No side effects have been reported in studies of vaccination with wild-type
30 adenovirus (Couch *et al.*, 1963; Top *et al.*, 1971), demonstrating their safety and therapeutic potential as *in vivo* gene transfer vectors.

Adenovirus vectors have been used in eukaryotic gene expression (Levrero *et al.*, 1991; Gomez-Foix *et al.*, 1992) and vaccine development (Grunhaus and Horwitz, 1992; Graham and Prevec, 1992). Recently, animal studies suggested that recombinant adenovirus could be used for gene therapy (Stratford-Perricaudet and Perricaudet, 1991; Stratford-Perricaudet *et al.*, 1990; Rich *et al.*, 1993). Studies in administering recombinant adenovirus to different tissues include trachea instillation (Rosenfeld *et al.*, 1991; Rosenfeld *et al.*, 1992), muscle injection (Ragot *et al.*, 1993), peripheral intravenous injections (Herz and Gerard, 1993) and stereotactic inoculation into the brain (Le Gal La Salle *et al.*, 1993).

10 2. RETROVIRUSES

The retroviruses are a group of single-stranded RNA viruses characterized by an ability to convert their RNA to double-stranded DNA in infected cells by a process of reverse-transcription (Coffin, 1990). The resulting DNA then stably integrates into cellular chromosomes as a provirus and directs synthesis of viral proteins. The integration results in the retention of the viral gene sequences in the recipient cell and its descendants. The retroviral genome contains three genes, gag, pol, and env that code for capsid proteins, polymerase enzyme, and envelope components, respectively. A sequence found upstream from the gag gene contains a signal for packaging of the genome into virions. Two long terminal repeat (LTR) sequences are present at the 5' and 3' ends of the viral genome. These contain strong promoter and enhancer sequences and are also required for integration in the host cell genome (Coffin, 1990).

In order to construct a retroviral vector, a nucleic acid encoding one or more oligonucleotide or polynucleotide sequences of interest is inserted into the viral genome in the place of certain viral sequences to produce a virus that is replication-defective. In order to produce virions, a packaging cell line containing the gag, pol, and env genes but without the LTR and packaging components is constructed (Mann *et al.*, 1983). When a recombinant plasmid containing a cDNA, together with the retroviral LTR and packaging sequences is introduced into this cell line (by calcium phosphate precipitation for example), the packaging sequence allows the RNA transcript of the recombinant plasmid to be packaged into viral particles, which are then secreted into

the culture media (Nicolas and Rubenstein, 1988; Temin, 1986; Mann *et al.*, 1983). The media containing the recombinant retroviruses is then collected, optionally concentrated, and used for gene transfer. Retroviral vectors are able to infect a broad variety of cell types. However, integration and stable expression require the division of
5 host cells (Paskind *et al.*, 1975).

A novel approach designed to allow specific targeting of retrovirus vectors was recently developed based on the chemical modification of a retrovirus by the chemical addition of lactose residues to the viral envelope. This modification could permit the specific infection of hepatocytes *via* sialoglycoprotein receptors.

10 A different approach to targeting of recombinant retroviruses was designed in which biotinylated antibodies against a retroviral envelope protein and against a specific cell receptor were used. The antibodies were coupled *via* the biotin components by using streptavidin (Roux *et al.*, 1989). Using antibodies against major histocompatibility complex class I and class II antigens, they demonstrated the infection
15 of a variety of human cells that bore those surface antigens with an ecotropic virus *in vitro* (Roux *et al.*, 1989).

3. ADENO-ASSOCIATED VIRUSES

AAV (Ridgeway, 1988; Hermonat and Muzyczka, 1984) is a parovirus, discovered as a contamination of adenoviral stocks. It is a ubiquitous virus (antibodies
20 are present in 85% of the US human population) that has not been linked to any disease. It is also classified as a dependovirus, because its replications is dependent on the presence of a helper virus, such as adenovirus. Five serotypes have been isolated, of which AAV-2 is the best characterized. AAV has a single-stranded linear DNA that is encapsidated into capsid proteins VP1, VP2 and VP3 to form an icosahedral virion of
25 20 to 24 nm in diameter (Muzyczka and McLaughlin, 1988).

The AAV DNA is approximately 4.7 kilobases long. It contains two open reading frames and is flanked by two ITRs (FIG. 2). There are two major genes in the AAV genome: *rep* and *cap*. The *rep* gene codes for proteins responsible for viral replications, whereas *cap* codes for capsid protein VP1-3. Each ITR forms a T-shaped
30 hairpin structure. These terminal repeats are the only essential *cis* components of the AAV for chromosomal integration. Therefore, the AAV can be used as a vector with

all viral coding sequences removed and replaced by the cassette of genes for delivery. Three viral promoters have been identified and named p5, p19, and p40, according to their map position. Transcription from p5 and p19 results in production of rep proteins, and transcription from p40 produces the capsid proteins (Hermonat and Muzyczka,
5 1984).

There are several factors that prompted researchers to study the possibility of using rAAV as an expression vector. One is that the requirements for delivering a gene to integrate into the host chromosome are surprisingly few. It is necessary to have the 145-bp ITRs, which are only 6% of the AAV genome. This
10 leaves room in the vector to assemble a 4.5-kb DNA insertion. While this carrying capacity may prevent the AAV from delivering large genes, it is amply suited for delivering the antisense constructs of the present invention.

AAV is also a good choice of delivery vehicles due to its safety. There is a relatively complicated rescue mechanism: not only wild type adenovirus but also
15 AAV genes are required to mobilize rAAV. Likewise, AAV is not pathogenic and not associated with any disease. The removal of viral coding sequences minimizes immune reactions to viral gene expression, and therefore, rAAV does not evoke an inflammatory response.

4. OTHER VIRAL VECTORS AS EXPRESSION CONSTRUCTS

20 Other viral vectors may be employed as expression constructs in the present invention for the delivery of oligonucleotide or polynucleotide sequences to a host cell. Vectors derived from viruses such as vaccinia virus (Ridgeway, 1988; Coupar *et al.*, 1988), lentiviruses, polio viruses and herpes viruses may be employed. They offer several attractive features for various mammalian cells (Friedmann, 1989;
25 Ridgeway, 1988; Coupar *et al.*, 1988; Horwich *et al.*, 1990).

With the recent recognition of defective hepatitis B viruses, new insight was gained into the structure-function relationship of different viral sequences. *In vitro* studies showed that the virus could retain the ability for helper-dependent packaging and reverse transcription despite the deletion of up to 80% of its genome (Horwich *et al.*, 1990). This suggested that large portions of the genome could be replaced with
30 foreign genetic material. The hepatotropism and persistence (integration) were

particularly attractive properties for liver-directed gene transfer. Chang *et al.* (1991) introduced the chloramphenicol acetyltransferase (CAT) gene into duck hepatitis B virus genome in the place of the polymerase, surface, and pre-surface coding sequences. It was cotransfected with wild-type virus into an avian hepatoma cell line. Culture media containing high titers of the recombinant virus were used to infect primary duckling hepatocytes. Stable CAT gene expression was detected for at least 24 days after transfection (Chang *et al.*, 1991).

5. NON-VIRAL VECTORS

In order to effect expression of the oligonucleotide or polynucleotide sequences of the present invention, the expression construct must be delivered into a cell. This delivery may be accomplished *in vitro*, as in laboratory procedures for transforming cells lines, or *in vivo* or *ex vivo*, as in the treatment of certain disease states. As described above, one preferred mechanism for delivery is *via* viral infection where the expression construct is encapsulated in an infectious viral particle.

Once the expression construct has been delivered into the cell the nucleic acid encoding the desired oligonucleotide or polynucleotide sequences may be positioned and expressed at different sites. In certain embodiments, the nucleic acid encoding the construct may be stably integrated into the genome of the cell. This integration may be in the specific location and orientation *via* homologous recombination (gene replacement) or it may be integrated in a random, non-specific location (gene augmentation). In yet further embodiments, the nucleic acid may be stably maintained in the cell as a separate, episomal segment of DNA. Such nucleic acid segments or "episomes" encode sequences sufficient to permit maintenance and replication independent of or in synchronization with the host cell cycle. How the expression construct is delivered to a cell and where in the cell the nucleic acid remains is dependent on the type of expression construct employed.

In certain embodiments of the invention, the expression construct comprising one or more oligonucleotide or polynucleotide sequences may simply consist of naked recombinant DNA or plasmids. Transfer of the construct may be performed by any of the methods mentioned above which physically or chemically permeabilize the cell membrane. This is particularly applicable for transfer *in vitro* but

it may be applied to *in vivo* use as well. Dubensky *et al.* (1984) successfully injected polyomavirus DNA in the form of calcium phosphate precipitates into liver and spleen of adult and newborn mice demonstrating active viral replication and acute infection. Benvenisty and Reshef (1986) also demonstrated that direct intraperitoneal injection of calcium phosphate-precipitated plasmids results in expression of the transfected genes. It is envisioned that DNA encoding a gene of interest may also be transferred in a similar manner *in vivo* and express the gene product.

Another embodiment of the invention for transferring a naked DNA expression construct into cells may involve particle bombardment. This method depends on the ability to accelerate DNA-coated microprojectiles to a high velocity allowing them to pierce cell membranes and enter cells without killing them (Klein *et al.*, 1987). Several devices for accelerating small particles have been developed. One such device relies on a high voltage discharge to generate an electrical current, which in turn provides the motive force (Yang *et al.*, 1990). The microprojectiles used have consisted of biologically inert substances such as tungsten or gold beads.

Selected organs including the liver, skin, and muscle tissue of rats and mice have been bombarded *in vivo* (Yang *et al.*, 1990; Zelenin *et al.*, 1991). This may require surgical exposure of the tissue or cells, to eliminate any intervening tissue between the gun and the target organ, *i.e. ex vivo* treatment. Again, DNA encoding a particular gene may be delivered *via* this method and still be incorporated by the present invention.

ANTISENSE OLIGONUCLEOTIDES

The end result of the flow of genetic information is the synthesis of protein. DNA is transcribed by polymerases into messenger RNA and translated on the ribosome to yield a folded, functional protein. Thus there are several steps along the route where protein synthesis can be inhibited. The native DNA segment coding for a polypeptide described herein, as all such mammalian DNA strands, has two strands: a sense strand and an antisense strand held together by hydrogen bonding. The messenger RNA coding for polypeptide has the same nucleotide sequence as the sense DNA strand except that the DNA thymidine is replaced by uridine. Thus, synthetic

antisense nucleotide sequences will bind to a mRNA and inhibit expression of the protein encoded by that mRNA.

The targeting of antisense oligonucleotides to mRNA is thus one mechanism to shut down protein synthesis, and, consequently, represents a powerful and targeted therapeutic approach. For example, the synthesis of polygalacturonase and the muscarine type 2 acetylcholine receptor are inhibited by antisense oligonucleotides directed to their respective mRNA sequences (U. S. Patent 5,739,119 and U. S. Patent 5,759,829, each specifically incorporated herein by reference in its entirety). Further, examples of antisense inhibition have been demonstrated with the nuclear protein cyclin, the multiple drug resistance gene (MDG1), ICAM-1, E-selectin, STK-1, striatal GABA_A receptor and human EGF (Jaskulski *et al.*, 1988; Vasanthakumar and Ahmed, 1989; Peris *et al.*, 1998; U. S. Patent 5,801,154; U. S. Patent 5,789,573; U. S. Patent 5,718,709 and U. S. Patent 5,610,288, each specifically incorporated herein by reference in its entirety). Antisense constructs have also been described that inhibit and can be used to treat a variety of abnormal cellular proliferations, *e.g.* cancer (U. S. Patent 5,747,470; U. S. Patent 5,591,317 and U. S. Patent 5,783,683, each specifically incorporated herein by reference in its entirety).

Therefore, in exemplary embodiments, the invention provides oligonucleotide sequences that comprise all, or a portion of, any sequence that is capable of specifically binding to polynucleotide sequence described herein, or a complement thereof. In one embodiment, the antisense oligonucleotides comprise DNA or derivatives thereof. In another embodiment, the oligonucleotides comprise RNA or derivatives thereof. In a third embodiment, the oligonucleotides are modified DNAs comprising a phosphorothioated modified backbone. In a fourth embodiment, the oligonucleotide sequences comprise peptide nucleic acids or derivatives thereof. In each case, preferred compositions comprise a sequence region that is complementary, and more preferably substantially-complementary, and even more preferably, completely complementary to one or more portions of polynucleotides disclosed herein.

Selection of antisense compositions specific for a given gene sequence is based upon analysis of the chosen target sequence (*i.e.* in these illustrative examples the rat and human sequences) and determination of secondary structure, T_m , binding energy, relative stability, and antisense compositions were selected based upon their

relative inability to form dimers, hairpins, or other secondary structures that would reduce or prohibit specific binding to the target mRNA in a host cell.

Highly preferred target regions of the mRNA, are those which are at or near the AUG translation initiation codon, and those sequences which were
5 substantially complementary to 5' regions of the mRNA. These secondary structure analyses and target site selection considerations were performed using v.4 of the OLIGO primer analysis software (Rychlik, 1997) and the BLASTN 2.0.5 algorithm software (Altschul *et al.*, 1997).

The use of an antisense delivery method employing a short peptide
10 vector, termed MPG (27 residues), is also contemplated. The MPG peptide contains a hydrophobic domain derived from the fusion sequence of HIV gp41 and a hydrophilic domain from the nuclear localization sequence of SV40 T-antigen (Morris *et al.*, 1997). It has been demonstrated that several molecules of the MPG peptide coat the antisense oligonucleotides and can be delivered into cultured mammalian cells in less than 1 hour
15 with relatively high efficiency (90%). Further, the interaction with MPG strongly increases both the stability of the oligonucleotide to nuclease and the ability to cross the plasma membrane (Morris *et al.*, 1997).

RIBOZYMES

Although proteins traditionally have been used for catalysis of nucleic
20 acids, another class of macromolecules has emerged as useful in this endeavor. Ribozymes are RNA-protein complexes that cleave nucleic acids in a site-specific fashion. Ribozymes have specific catalytic domains that possess endonuclease activity (Kim and Cech, 1987; Gerlach *et al.*, 1987; Forster and Symons, 1987). For example, a large number of ribozymes accelerate phosphoester transfer reactions with a high
25 degree of specificity, often cleaving only one of several phosphoesters in an oligonucleotide substrate (Cech *et al.*, 1981; Michel and Westhof, 1990; Reinhold-Hurek and Shub, 1992). This specificity has been attributed to the requirement that the substrate bind via specific base-pairing interactions to the internal guide sequence ("IGS") of the ribozyme prior to chemical reaction.

30 Ribozyme catalysis has primarily been observed as part of sequence-specific cleavage/ligation reactions involving nucleic acids (Joyce, 1989; Cech *et al.*,

1981). For example, U. S. Patent No. 5,354,855 (specifically incorporated herein by reference) reports that certain ribozymes can act as endonucleases with a sequence specificity greater than that of known ribonucleases and approaching that of the DNA restriction enzymes. Thus, sequence-specific ribozyme-mediated inhibition of gene
5 expression may be particularly suited to therapeutic applications (Scanlon *et al.*, 1991; Sarver *et al.*, 1990). Recently, it was reported that ribozymes elicited genetic changes in some cells lines to which they were applied; the altered genes included the oncogenes *H-ras*, *c-fos* and genes of HIV. Most of this work involved the modification of a target mRNA, based on a specific mutant codon that is cleaved by a specific ribozyme.

10 Six basic varieties of naturally-occurring enzymatic RNAs are known presently. Each can catalyze the hydrolysis of RNA phosphodiester bonds *in trans* (and thus can cleave other RNA molecules) under physiological conditions. In general, enzymatic nucleic acids act by first binding to a target RNA. Such binding occurs through the target binding portion of a enzymatic nucleic acid which is held in close
15 proximity to an enzymatic portion of the molecule that acts to cleave the target RNA. Thus, the enzymatic nucleic acid first recognizes and then binds a target RNA through complementary base-pairing, and once bound to the correct site, acts enzymatically to cut the target RNA. Strategic cleavage of such a target RNA will destroy its ability to direct synthesis of an encoded protein. After an enzymatic nucleic acid has bound and
20 cleaved its RNA target, it is released from that RNA to search for another target and can repeatedly bind and cleave new targets.

 The enzymatic nature of a ribozyme is advantageous over many technologies, such as antisense technology (where a nucleic acid molecule simply binds to a nucleic acid target to block its translation) since the concentration of ribozyme
25 necessary to affect a therapeutic treatment is lower than that of an antisense oligonucleotide. This advantage reflects the ability of the ribozyme to act enzymatically. Thus, a single ribozyme molecule is able to cleave many molecules of target RNA. In addition, the ribozyme is a highly specific inhibitor, with the specificity of inhibition depending not only on the base pairing mechanism of binding to the target
30 RNA, but also on the mechanism of target RNA cleavage. Single mismatches, or base-substitutions, near the site of cleavage can completely eliminate catalytic activity of a ribozyme. Similar mismatches in antisense molecules do not prevent their action

(Woolf *et al.*, 1992). Thus, the specificity of action of a ribozyme is greater than that of an antisense oligonucleotide binding the same RNA site.

The enzymatic nucleic acid molecule may be formed in a hammerhead, hairpin, a hepatitis δ virus, group I intron or RNaseP RNA (in association with an RNA
5 guide sequence) or Neurospora VS RNA motif. Examples of hammerhead motifs are described by Rossi *et al.* (1992). Examples of hairpin motifs are described by Hampel
et al. (Eur. Pat. Appl. Publ. No. EP 0360257), Hampel and Tritz (1989), Hampel *et al.* (1990) and U. S. Patent 5,631,359 (specifically incorporated herein by reference). An
example of the hepatitis δ virus motif is described by Perrotta and Been (1992); an
10 example of the RNaseP motif is described by Guerrier-Takada *et al.* (1983); Neurospora VS RNA ribozyme motif is described by Collins (Saville and Collins, 1990; Saville and Collins, 1991; Collins and Olive, 1993); and an example of the Group I intron is described in (U. S. Patent 4,987,071, specifically incorporated herein by
reference). All that is important in an enzymatic nucleic acid molecule of this invention
15 is that it has a specific substrate binding site which is complementary to one or more of the target gene RNA regions, and that it have nucleotide sequences within or surrounding that substrate binding site which impart an RNA cleaving activity to the molecule. Thus the ribozyme constructs need not be limited to specific motifs mentioned herein.

20 In certain embodiments, it may be important to produce enzymatic cleaving agents which exhibit a high degree of specificity for the RNA of a desired target, such as one of the sequences disclosed herein. The enzymatic nucleic acid molecule is preferably targeted to a highly conserved sequence region of a target mRNA. Such enzymatic nucleic acid molecules can be delivered exogenously to
25 specific cells as required. Alternatively, the ribozymes can be expressed from DNA or RNA vectors that are delivered to specific cells.

Small enzymatic nucleic acid motifs (*e.g.*, of the hammerhead or the hairpin structure) may also be used for exogenous delivery. The simple structure of these molecules increases the ability of the enzymatic nucleic acid to invade targeted
30 regions of the mRNA structure. Alternatively, catalytic RNA molecules can be expressed within cells from eukaryotic promoters (*e.g.*, Scanlon *et al.*, 1991; Kashani-Sabet *et al.*, 1992; Dropulic *et al.*, 1992; Weerasinghe *et al.*, 1991; Ojwang *et al.*, 1992;

Chen *et al.*, 1992; Sarver *et al.*, 1990). Those skilled in the art realize that any ribozyme can be expressed in eukaryotic cells from the appropriate DNA vector. The activity of such ribozymes can be augmented by their release from the primary transcript by a second ribozyme (Int. Pat. Appl. Publ. No. WO 93/23569, and Int. Pat. Appl. Publ. No. WO 94/02595, both hereby incorporated by reference; Ohkawa *et al.*, 1992; Taira *et al.*, 1991; and Ventura *et al.*, 1993).

Ribozymes may be added directly, or can be complexed with cationic lipids, lipid complexes, packaged within liposomes, or otherwise delivered to target cells. The RNA or RNA complexes can be locally administered to relevant tissues *ex vivo*, or *in vivo* through injection, aerosol inhalation, infusion pump or stent, with or without their incorporation in biopolymers.

Ribozymes may be designed as described in Int. Pat. Appl. Publ. No. WO 93/23569 and Int. Pat. Appl. Publ. No. WO 94/02595, each specifically incorporated herein by reference) and synthesized to be tested *in vitro* and *in vivo*, as described. Such ribozymes can also be optimized for delivery. While specific examples are provided, those in the art will recognize that equivalent RNA targets in other species can be utilized when necessary.

Hammerhead or hairpin ribozymes may be individually analyzed by computer folding (Jaeger *et al.*, 1989) to assess whether the ribozyme sequences fold into the appropriate secondary structure. Those ribozymes with unfavorable intramolecular interactions between the binding arms and the catalytic core are eliminated from consideration. Varying binding arm lengths can be chosen to optimize activity. Generally, at least 5 or so bases on each arm are able to bind to, or otherwise interact with, the target RNA.

Ribozymes of the hammerhead or hairpin motif may be designed to anneal to various sites in the mRNA message, and can be chemically synthesized. The method of synthesis used follows the procedure for normal RNA synthesis as described in Usman *et al.* (1987) and in Scaringe *et al.* (1990) and makes use of common nucleic acid protecting and coupling groups, such as dimethoxytrityl at the 5'-end, and phosphoramidites at the 3'-end. Average stepwise coupling yields are typically >98%. Hairpin ribozymes may be synthesized in two parts and annealed to reconstruct an active ribozyme (Chowrira and Burke, 1992). Ribozymes may be modified extensively

to enhance stability by modification with nuclease resistant groups, for example, 2'-amino, 2'-C-allyl, 2'-fluoro, 2'-o-methyl, 2'-H (for a review see *e.g.*, Usman and Cedergren, 1992). Ribozymes may be purified by gel electrophoresis using general methods or by high pressure liquid chromatography and resuspended in water.

5 Ribozyme activity can be optimized by altering the length of the ribozyme binding arms, or chemically synthesizing ribozymes with modifications that prevent their degradation by serum ribonucleases (see *e.g.*, Int. Pat. Appl. Publ. No. WO 92/07065; Perrault *et al.*, 1990; Pieken *et al.*, 1991; Usman and Cedergren, 1992; Int. Pat. Appl. Publ. No. WO 93/15187; Int. Pat. Appl. Publ. No. WO 91/03162; Eur. Pat. Appl. Publ. No. 92110298.4; U. S. Patent 5,334,711; and Int. Pat. Appl. Publ. No. WO 94/13688, which describe various chemical modifications that can be made to the sugar moieties of enzymatic RNA molecules), modifications which enhance their efficacy in cells, and removal of stem II bases to shorten RNA synthesis times and reduce chemical requirements.

15 Sullivan *et al.* (Int. Pat. Appl. Publ. No. WO 94/02595) describes the general methods for delivery of enzymatic RNA molecules. Ribozymes may be administered to cells by a variety of methods known to those familiar to the art, including, but not restricted to, encapsulation in liposomes, by iontophoresis, or by incorporation into other vehicles, such as hydrogels, cyclodextrins, biodegradable nanocapsules, and bioadhesive microspheres. For some indications, ribozymes may be directly delivered *ex vivo* to cells or tissues with or without the aforementioned vehicles. Alternatively, the RNA/vehicle combination may be locally delivered by direct inhalation, by direct injection or by use of a catheter, infusion pump or stent. Other routes of delivery include, but are not limited to, intravascular, intramuscular, 20 subcutaneous or joint injection, aerosol inhalation, oral (tablet or pill form), topical, systemic, ocular, intraperitoneal and/or intrathecal delivery. More detailed descriptions of ribozyme delivery and administration are provided in Int. Pat. Appl. Publ. No. WO 94/02595 and Int. Pat. Appl. Publ. No. WO 93/23569, each specifically incorporated herein by reference.

30 Another means of accumulating high concentrations of a ribozyme(s) within cells is to incorporate the ribozyme-encoding sequences into a DNA expression vector. Transcription of the ribozyme sequences are driven from a promoter for

eukaryotic RNA polymerase I (pol I), RNA polymerase II (pol II), or RNA polymerase III (pol III). Transcripts from pol II or pol III promoters will be expressed at high levels in all cells; the levels of a given pol II promoter in a given cell type will depend on the nature of the gene regulatory sequences (enhancers, silencers, *etc.*) present nearby.

- 5 Prokaryotic RNA polymerase promoters may also be used, providing that the prokaryotic RNA polymerase enzyme is expressed in the appropriate cells (Elroy-Stein and Moss, 1990; Gao and Huang, 1993; Lieber *et al.*, 1993; Zhou *et al.*, 1990). Ribozymes expressed from such promoters can function in mammalian cells (*e.g.* Kashani-Saber *et al.*, 1992; Ojwang *et al.*, 1992; Chen *et al.*, 1992; Yu *et al.*, 1993; 10 L'Huillier *et al.*, 1992; Lisiewicz *et al.*, 1993). Such transcription units can be incorporated into a variety of vectors for introduction into mammalian cells, including but not restricted to, plasmid DNA vectors, viral DNA vectors (such as adenovirus or adeno-associated vectors), or viral RNA vectors (such as retroviral, semliki forest virus, sindbis virus vectors).

- 15 Ribozymes may be used as diagnostic tools to examine genetic drift and mutations within diseased cells. They can also be used to assess levels of the target RNA molecule. The close relationship between ribozyme activity and the structure of the target RNA allows the detection of mutations in any region of the molecule which alters the base-pairing and three-dimensional structure of the target RNA. By using 20 multiple ribozymes, one may map nucleotide changes which are important to RNA structure and function *in vitro*, as well as in cells and tissues. Cleavage of target RNAs with ribozymes may be used to inhibit gene expression and define the role (essentially) of specified gene products in the progression of disease. In this manner, other genetic targets may be defined as important mediators of the disease. These studies will lead to 25 better treatment of the disease progression by affording the possibility of combinational therapies (*e.g.*, multiple ribozymes targeted to different genes, ribozymes coupled with known small molecule inhibitors, or intermittent treatment with combinations of ribozymes and/or other chemical or biological molecules). Other *in vitro* uses of ribozymes are well known in the art, and include detection of the presence of mRNA 30 associated with an IL-5 related condition. Such RNA is detected by determining the presence of a cleavage product after treatment with a ribozyme using standard methodology.

PEPTIDE NUCLEIC ACIDS

In certain embodiments, the inventors contemplate the use of peptide nucleic acids (PNAs) in the practice of the methods of the invention. PNA is a DNA mimic in which the nucleobases are attached to a pseudopeptide backbone (Good and
5 Nielsen, 1997). PNA is able to be utilized in a number methods that traditionally have used RNA or DNA. Often PNA sequences perform better in techniques than the corresponding RNA or DNA sequences and have utilities that are not inherent to RNA or DNA. A review of PNA including methods of making, characteristics of, and methods of using, is provided by Corey (1997) and is incorporated herein by reference.
10 As such, in certain embodiments, one may prepare PNA sequences that are complementary to one or more portions of the ACE mRNA sequence, and such PNA compositions may be used to regulate, alter, decrease, or reduce the translation of ACE-specific mRNA, and thereby alter the level of ACE activity in a host cell to which such PNA compositions have been administered.
15 PNAs have 2-aminoethyl-glycine linkages replacing the normal phosphodiester backbone of DNA (Nielsen *et al.*, 1991; Hanvey *et al.*, 1992; Hyrup and Nielsen, 1996; Neilsen, 1996). This chemistry has three important consequences: firstly, in contrast to DNA or phosphorothioate oligonucleotides, PNAs are neutral molecules; secondly, PNAs are achiral, which avoids the need to develop a
20 stereoselective synthesis; and thirdly, PNA synthesis uses standard Boc (Dueholm *et al.*, 1994) or Fmoc (Thomson *et al.*, 1995) protocols for solid-phase peptide synthesis, although other methods, including a modified Merrifield method, have been used (Christensen *et al.*, 1995).

PNA monomers or ready-made oligomers are commercially available
25 from PerSeptive Biosystems (Framingham, MA). PNA syntheses by either Boc or Fmoc protocols are straightforward using manual or automated protocols (Norton *et al.*, 1995). The manual protocol lends itself to the production of chemically modified PNAs or the simultaneous synthesis of families of closely related PNAs.

As with peptide synthesis, the success of a particular PNA synthesis will
30 depend on the properties of the chosen sequence. For example, while in theory PNAs can incorporate any combination of nucleotide bases, the presence of adjacent purines can lead to deletions of one or more residues in the product. In expectation of this

difficulty, it is suggested that, in producing PNAs with adjacent purines, one should repeat the coupling of residues likely to be added inefficiently. This should be followed by the purification of PNAs by reverse-phase high-pressure liquid chromatography (Norton *et al.*, 1995) providing yields and purity of product similar to those observed
5 during the synthesis of peptides.

Modifications of PNAs for a given application may be accomplished by coupling amino acids during solid-phase synthesis or by attaching compounds that contain a carboxylic acid group to the exposed N-terminal amine. Alternatively, PNAs can be modified after synthesis by coupling to an introduced lysine or cysteine. The
10 ease with which PNAs can be modified facilitates optimization for better solubility or for specific functional requirements. Once synthesized, the identity of PNAs and their derivatives can be confirmed by mass spectrometry. Several studies have made and utilized modifications of PNAs (Norton *et al.*, 1995; Haaima *et al.*, 1996; Stetsenko *et al.*, 1996; Petersen *et al.*, 1995; Ulmann *et al.*, 1996; Koch *et al.*, 1995; Orum *et al.*,
15 1995; Footer *et al.*, 1996; Griffith *et al.*, 1995; Kremsky *et al.*, 1996; Pardridge *et al.*, 1995; Boffa *et al.*, 1995; Landsdorp *et al.*, 1996; Gambacorti-Passerini *et al.*, 1996; Armitage *et al.*, 1997; Seeger *et al.*, 1997; Ruskowski *et al.*, 1997). U.S. Patent No. 5,700,922 discusses PNA-DNA-PNA chimeric molecules and their uses in diagnostics, modulating protein in organisms, and treatment of conditions susceptible to
20 therapeutics.

In contrast to DNA and RNA, which contain negatively charged linkages, the PNA backbone is neutral. In spite of this dramatic alteration, PNAs recognize complementary DNA and RNA by Watson-Crick pairing (Egholm *et al.*, 1993), validating the initial modeling by Nielsen *et al.* (1991). PNAs lack 3' to 5'
25 polarity and can bind in either parallel or antiparallel fashion, with the antiparallel mode being preferred (Egholm *et al.*, 1993).

Hybridization of DNA oligonucleotides to DNA and RNA is destabilized by electrostatic repulsion between the negatively charged phosphate backbones of the complementary strands. By contrast, the absence of charge repulsion
30 in PNA-DNA or PNA-RNA duplexes increases the melting temperature (T_m) and reduces the dependence of T_m on the concentration of mono- or divalent cations (Nielsen *et al.*, 1991). The enhanced rate and affinity of hybridization are significant

because they are responsible for the surprising ability of PNAs to perform strand invasion of complementary sequences within relaxed double-stranded DNA. In addition, the efficient hybridization at inverted repeats suggests that PNAs can recognize secondary structure effectively within double-stranded DNA. Enhanced
5 recognition also occurs with PNAs immobilized on surfaces, and Wang *et al.* have shown that support-bound PNAs can be used to detect hybridization events (Wang *et al.*, 1996).

One might expect that tight binding of PNAs to complementary sequences would also increase binding to similar (but not identical) sequences, reducing
10 the sequence specificity of PNA recognition. As with DNA hybridization, however, selective recognition can be achieved by balancing oligomer length and incubation temperature. Moreover, selective hybridization of PNAs is encouraged by PNA-DNA hybridization being less tolerant of base mismatches than DNA-DNA hybridization. For example, a single mismatch within a 16 bp PNA-DNA duplex can reduce the T_m by
15 up to 15°C (Egholm *et al.*, 1993). This high level of discrimination has allowed the development of several PNA-based strategies for the analysis of point mutations (Wang *et al.*, 1996; Carlsson *et al.*, 1996; Thiede *et al.*, 1996; Webb and Hurskainen, 1996; Perry-O'Keefe *et al.*, 1996).

High-affinity binding provides clear advantages for molecular
20 recognition and the development of new applications for PNAs. For example, 11-13 nucleotide PNAs inhibit the activity of telomerase, a ribonucleo-protein that extends telomere ends using an essential RNA template, while the analogous DNA oligomers do not (Norton *et al.*, 1996).

Neutral PNAs are more hydrophobic than analogous DNA oligomers,
25 and this can lead to difficulty solubilizing them at neutral pH, especially if the PNAs have a high purine content or if they have the potential to form secondary structures. Their solubility can be enhanced by attaching one or more positive charges to the PNA termini (Nielsen *et al.*, 1991).

Findings by Allfrey and colleagues suggest that strand invasion will
30 occur spontaneously at sequences within chromosomal DNA (Boffa *et al.*, 1995; Boffa *et al.*, 1996). These studies targeted PNAs to triplet repeats of the nucleotides CAG and used this recognition to purify transcriptionally active DNA (Boffa *et al.*, 1995) and to

inhibit transcription (Boffa *et al.*, 1996). This result suggests that if PNAs can be delivered within cells then they will have the potential to be general sequence-specific regulators of gene expression. Studies and reviews concerning the use of PNAs as antisense and anti-gene agents include Nielsen *et al.* (1993b), Hanvey *et al.* (1992), and
5 Good and Nielsen (1997). Koppelhus *et al.* (1997) have used PNAs to inhibit HIV-1 inverse transcription, showing that PNAs may be used for antiviral therapies.

Methods of characterizing the antisense binding properties of PNAs are discussed in Rose (1993) and Jensen *et al.* (1997). Rose uses capillary gel electrophoresis to determine binding of PNAs to their complementary oligonucleotide,
10 measuring the relative binding kinetics and stoichiometry. Similar types of measurements were made by Jensen *et al.* using BIAcore™ technology.

Other applications of PNAs include use in DNA strand invasion (Nielsen *et al.*, 1991), antisense inhibition (Hanvey *et al.*, 1992), mutational analysis (Orum *et al.*, 1993), enhancers of transcription (Mollegaard *et al.*, 1994), nucleic acid purification
15 (Orum *et al.*, 1995), isolation of transcriptionally active genes (Boffa *et al.*, 1995), blocking of transcription factor binding (Vickers *et al.*, 1995), genome cleavage (Veselkov *et al.*, 1996), biosensors (Wang *et al.*, 1996), *in situ* hybridization (Thisted *et al.*, 1996), and in a alternative to Southern blotting (Perry-O'Keefe, 1996).

POLYPEPTIDE COMPOSITIONS

20 The present invention, in other aspects, provides polypeptide compositions. Generally, a polypeptide of the invention will be an isolated polypeptide (or an epitope, variant, or active fragment thereof) derived from a mammalian species. Preferably, the polypeptide is encoded by a polynucleotide sequence disclosed herein or a sequence which hybridizes under moderately stringent conditions to a polynucleotide
25 sequence disclosed herein. Alternatively, the polypeptide may be defined as a polypeptide which comprises a contiguous amino acid sequence from an amino acid sequence disclosed herein, or which polypeptide comprises an entire amino acid sequence disclosed herein.

In the present invention, a polypeptide composition is also understood to
30 comprise one or more polypeptides that are immunologically reactive with antibodies generated against a polypeptide of the invention, particularly a polypeptide having the

amino acid sequence encoded by SEQ ID NOs:1-222, or to active fragments, or to variants or biological functional equivalents thereof.

Likewise, a polypeptide composition of the present invention is understood to comprise one or more polypeptides that are capable of eliciting antibodies
5 that are immunologically reactive with one or more polypeptides encoded by one or more contiguous nucleic acid sequences contained in SEQ ID NOs:1-222, or to active fragments, or to variants thereof, or to one or more nucleic acid sequences which hybridize to one or more of these sequences under conditions of moderate to high stringency.

10 As used herein, an active fragment of a polypeptide includes a whole or a portion of a polypeptide which is modified by conventional techniques, *e.g.*, mutagenesis, or by addition, deletion, or substitution, but which active fragment exhibits substantially the same structure function, antigenicity, etc., as a polypeptide as described herein.

15 In certain illustrative embodiments, the polypeptides of the invention will comprise at least an immunogenic portion of an ovarian tumor protein or a variant thereof, as described herein. As noted above, an "ovarian tumor protein" is a protein that is expressed by ovarian tumor cells. Proteins that are ovarian tumor proteins also react detectably within an immunoassay (such as an ELISA) with antisera from a
20 patient with ovarian cancer. Polypeptides as described herein may be of any length. Additional sequences derived from the native protein and/or heterologous sequences may be present, and such sequences may (but need not) possess further immunogenic or antigenic properties.

An "immunogenic portion," as used herein is a portion of a protein that
25 is recognized (*i.e.*, specifically bound) by a B-cell and/or T-cell surface antigen receptor. Such immunogenic portions generally comprise at least 5 amino acid residues, more preferably at least 10, and still more preferably at least 20 amino acid residues of an ovarian tumor protein or a variant thereof. Certain preferred immunogenic portions include peptides in which an N-terminal leader sequence and/or
30 transmembrane domain have been deleted. Other preferred immunogenic portions may contain a small N- and/or C-terminal deletion (*e.g.*, 1-30 amino acids, preferably 5-15 amino acids), relative to the mature protein.

Immunogenic portions may generally be identified using well known techniques, such as those summarized in Paul, *Fundamental Immunology*, 3rd ed., 243-247 (Raven Press, 1993) and references cited therein. Such techniques include screening polypeptides for the ability to react with antigen-specific antibodies, antisera and/or T-cell lines or clones. As used herein, antisera and antibodies are "antigen-specific" if they specifically bind to an antigen (*i.e.*, they react with the protein in an ELISA or other immunoassay, and do not react detectably with unrelated proteins). Such antisera and antibodies may be prepared as described herein, and using well known techniques. An immunogenic portion of a native ovarian tumor protein is a portion that reacts with such antisera and/or T-cells at a level that is not substantially less than the reactivity of the full length polypeptide (*e.g.*, in an ELISA and/or T-cell reactivity assay). Such immunogenic portions may react within such assays at a level that is similar to or greater than the reactivity of the full length polypeptide. Such screens may generally be performed using methods well known to those of ordinary skill in the art, such as those described in Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, 1988. For example, a polypeptide may be immobilized on a solid support and contacted with patient sera to allow binding of antibodies within the sera to the immobilized polypeptide. Unbound sera may then be removed and bound antibodies detected using, for example, ¹²⁵I-labeled Protein A.

As noted above, a composition may comprise a variant of a native ovarian tumor protein. A polypeptide "variant," as used herein, is a polypeptide that differs from a native ovarian tumor protein in one or more substitutions, deletions, additions and/or insertions, such that the immunogenicity of the polypeptide is not substantially diminished. In other words, the ability of a variant to react with antigen-specific antisera may be enhanced or unchanged, relative to the native protein, or may be diminished by less than 50%, and preferably less than 20%, relative to the native protein. Such variants may generally be identified by modifying one of the above polypeptide sequences and evaluating the reactivity of the modified polypeptide with antigen-specific antibodies or antisera as described herein. Preferred variants include those in which one or more portions, such as an N-terminal leader sequence or transmembrane domain, have been removed. Other preferred variants include variants

in which a small portion (*e.g.*, 1-30 amino acids, preferably 5-15 amino acids) has been removed from the N- and/or C-terminal of the mature protein.

Polypeptide variants encompassed by the present invention include those exhibiting at least about 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%,
5 97%, 98%, or 99% or more identity (determined as described above) to the polypeptides disclosed herein.

Preferably, a variant contains conservative substitutions. A "conservative substitution" is one in which an amino acid is substituted for another amino acid that has similar properties, such that one skilled in the art of peptide
10 chemistry would expect the secondary structure and hydropathic nature of the polypeptide to be substantially unchanged. Amino acid substitutions may generally be made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity and/or the amphipathic nature of the residues. For example, negatively charged amino acids include aspartic acid and glutamic acid; positively charged amino
15 acids include lysine and arginine; and amino acids with uncharged polar head groups having similar hydrophilicity values include leucine, isoleucine and valine; glycine and alanine; asparagine and glutamine; and serine, threonine, phenylalanine and tyrosine. Other groups of amino acids that may represent conservative changes include: (1) ala, pro, gly, glu, asp, gln, asn, ser, thr; (2) cys, ser, tyr, thr; (3) val, ile, leu, met, ala, phe;
20 (4) lys, arg, his; and (5) phe, tyr, trp, his. A variant may also, or alternatively, contain nonconservative changes. In a preferred embodiment, variant polypeptides differ from a native sequence by substitution, deletion or addition of five amino acids or fewer. Variants may also (or alternatively) be modified by, for example, the deletion or addition of amino acids that have minimal influence on the immunogenicity, secondary
25 structure and hydropathic nature of the polypeptide.

As noted above, polypeptides may comprise a signal (or leader) sequence at the N-terminal end of the protein, which co-translationally or post-translationally directs transfer of the protein. The polypeptide may also be conjugated to a linker or other sequence for ease of synthesis, purification or identification of the
30 polypeptide (*e.g.*, poly-His), or to enhance binding of the polypeptide to a solid support. For example, a polypeptide may be conjugated to an immunoglobulin Fc region.

Polypeptides may be prepared using any of a variety of well known techniques. Recombinant polypeptides encoded by DNA sequences as described above may be readily prepared from the DNA sequences using any of a variety of expression vectors known to those of ordinary skill in the art. Expression may be achieved in any appropriate host cell that has been transformed or transfected with an expression vector containing a DNA molecule that encodes a recombinant polypeptide. Suitable host cells include prokaryotes, yeast, and higher eukaryotic cells, such as mammalian cells and plant cells. Preferably, the host cells employed are *E. coli*, yeast or a mammalian cell line such as COS or CHO. Supernatants from suitable host/vector systems which secrete recombinant protein or polypeptide into culture media may be first concentrated using a commercially available filter. Following concentration, the concentrate may be applied to a suitable purification matrix such as an affinity matrix or an ion exchange resin. Finally, one or more reverse phase HPLC steps can be employed to further purify a recombinant polypeptide.

Portions and other variants having less than about 100 amino acids, and generally less than about 50 amino acids, may also be generated by synthetic means, using techniques well known to those of ordinary skill in the art. For example, such polypeptides may be synthesized using any of the commercially available solid-phase techniques, such as the Merrifield solid-phase synthesis method, where amino acids are sequentially added to a growing amino acid chain. See Merrifield, *J. Am. Chem. Soc.* 85:2149-2146, 1963. Equipment for automated synthesis of polypeptides is commercially available from suppliers such as Perkin Elmer/Applied BioSystems Division (Foster City, CA), and may be operated according to the manufacturer's instructions.

Within certain specific embodiments, a polypeptide may be a fusion protein that comprises multiple polypeptides as described herein, or that comprises at least one polypeptide as described herein and an unrelated sequence, such as a known tumor protein. A fusion partner may, for example, assist in providing T helper epitopes (an immunological fusion partner), preferably T helper epitopes recognized by humans, or may assist in expressing the protein (an expression enhancer) at higher yields than the native recombinant protein. Certain preferred fusion partners are both immunological and expression enhancing fusion partners. Other fusion partners may be

selected so as to increase the solubility of the protein or to enable the protein to be targeted to desired intracellular compartments. Still further fusion partners include affinity tags, which facilitate purification of the protein.

Fusion proteins may generally be prepared using standard techniques, including chemical conjugation. Preferably, a fusion protein is expressed as a recombinant protein, allowing the production of increased levels, relative to a non-fused protein, in an expression system. Briefly, DNA sequences encoding the polypeptide components may be assembled separately, and ligated into an appropriate expression vector. The 3' end of the DNA sequence encoding one polypeptide component is ligated, with or without a peptide linker, to the 5' end of a DNA sequence encoding the second polypeptide component so that the reading frames of the sequences are in phase. This permits translation into a single fusion protein that retains the biological activity of both component polypeptides.

A peptide linker sequence may be employed to separate the first and second polypeptide components by a distance sufficient to ensure that each polypeptide folds into its secondary and tertiary structures. Such a peptide linker sequence is incorporated into the fusion protein using standard techniques well known in the art. Suitable peptide linker sequences may be chosen based on the following factors: (1) their ability to adopt a flexible extended conformation; (2) their inability to adopt a secondary structure that could interact with functional epitopes on the first and second polypeptides; and (3) the lack of hydrophobic or charged residues that might react with the polypeptide functional epitopes. Preferred peptide linker sequences contain Gly, Asn and Ser residues. Other near neutral amino acids, such as Thr and Ala may also be used in the linker sequence. Amino acid sequences which may be usefully employed as linkers include those disclosed in Maratea *et al.*, *Gene* 40:39-46, 1985; Murphy *et al.*, *Proc. Natl. Acad. Sci. USA* 83:8258-8262, 1986; U.S. Patent No. 4,935,233 and U.S. Patent No. 4,751,180. The linker sequence may generally be from 1 to about 50 amino acids in length. Linker sequences are not required when the first and second polypeptides have non-essential N-terminal amino acid regions that can be used to separate the functional domains and prevent steric interference.

The ligated DNA sequences are operably linked to suitable transcriptional or translational regulatory elements. The regulatory elements

responsible for expression of DNA are located only 5' to the DNA sequence encoding the first polypeptides. Similarly, stop codons required to end translation and transcription termination signals are only present 3' to the DNA sequence encoding the second polypeptide.

5 Fusion proteins are also provided. Such proteins comprise a polypeptide as described herein together with an unrelated immunogenic protein. Preferably the immunogenic protein is capable of eliciting a recall response. Examples of such proteins include tetanus, tuberculosis and hepatitis proteins (*see, for example, Stoute et al. New Engl. J. Med.*, 336:86-91, 1997).

10 Within preferred embodiments, an immunological fusion partner is derived from protein D, a surface protein of the gram-negative bacterium *Haemophilus influenza B* (WO 91/18926). Preferably, a protein D derivative comprises approximately the first third of the protein (*e.g.*, the first N-terminal 100-110 amino acids), and a protein D derivative may be lipidated. Within certain preferred
15 embodiments, the first 109 residues of a Lipoprotein D fusion partner is included on the N-terminus to provide the polypeptide with additional exogenous T-cell epitopes and to increase the expression level in *E. coli* (thus functioning as an expression enhancer). The lipid tail ensures optimal presentation of the antigen to antigen presenting cells. Other fusion partners include the non-structural protein from influenzae virus, NS1
20 (hemagglutinin). Typically, the N-terminal 81 amino acids are used, although different fragments that include T-helper epitopes may be used.

 In another embodiment, the immunological fusion partner is the protein known as LYTA, or a portion thereof (preferably a C-terminal portion). LYTA is derived from *Streptococcus pneumoniae*, which synthesizes an N-acetyl-L-alanine
25 amidase known as amidase LYTA (encoded by the *LytA* gene; *Gene* 43:265-292, 1986). LYTA is an autolysin that specifically degrades certain bonds in the peptidoglycan backbone. The C-terminal domain of the LYTA protein is responsible for the affinity to the choline or to some choline analogues such as DEAE. This property has been exploited for the development of *E. coli* C-LYTA expressing
30 plasmids useful for expression of fusion proteins. Purification of hybrid proteins containing the C-LYTA fragment at the amino terminus has been described (*see Biotechnology* 10:795-798, 1992). Within a preferred embodiment, a repeat portion of

LYTA may be incorporated into a fusion protein. A repeat portion is found in the C-terminal region starting at residue 178. A particularly preferred repeat portion incorporates residues 188-305.

In general, polypeptides (including fusion proteins) and polynucleotides as described herein are isolated. An "isolated" polypeptide or polynucleotide is one that is removed from its original environment. For example, a naturally-occurring protein is isolated if it is separated from some or all of the coexisting materials in the natural system. Preferably, such polypeptides are at least about 90% pure, more preferably at least about 95% pure and most preferably at least about 99% pure. A polynucleotide is considered to be isolated if, for example, it is cloned into a vector that is not a part of the natural environment.

BINDING AGENTS

The present invention further provides agents, such as antibodies and antigen-binding fragments thereof, that specifically bind to an ovarian tumor protein. As used herein, an antibody, or antigen-binding fragment thereof, is said to "specifically bind" to an ovarian tumor protein if it reacts at a detectable level (within, for example, an ELISA) with an ovarian tumor protein, and does not react detectably with unrelated proteins under similar conditions. As used herein, "binding" refers to a noncovalent association between two separate molecules such that a complex is formed. The ability to bind may be evaluated by, for example, determining a binding constant for the formation of the complex. The binding constant is the value obtained when the concentration of the complex is divided by the product of the component concentrations. In general, two compounds are said to "bind," in the context of the present invention, when the binding constant for complex formation exceeds about 10^3 L/mol. The binding constant may be determined using methods well known in the art.

Binding agents may be further capable of differentiating between patients with and without a cancer, such as ovarian cancer, using the representative assays provided herein. In other words, antibodies or other binding agents that bind to an ovarian tumor protein will generate a signal indicating the presence of a cancer in at least about 20% of patients with the disease, and will generate a negative signal indicating the absence of the disease in at least about 90% of individuals without the

cancer. To determine whether a binding agent satisfies this requirement, biological samples (*e.g.*, blood, sera, sputum, urine and/or tumor biopsies) from patients with and without a cancer (as determined using standard clinical tests) may be assayed as described herein for the presence of polypeptides that bind to the binding agent. It will
5 be apparent that a statistically significant number of samples with and without the disease should be assayed. Each binding agent should satisfy the above criteria; however, those of ordinary skill in the art will recognize that binding agents may be used in combination to improve sensitivity.

Any agent that satisfies the above requirements may be a binding agent.
10 For example, a binding agent may be a ribosome, with or without a peptide component, an RNA molecule or a polypeptide. In a preferred embodiment, a binding agent is an antibody or an antigen-binding fragment thereof. Antibodies may be prepared by any of a variety of techniques known to those of ordinary skill in the art. *See, e.g.*, Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, 1988. In
15 general, antibodies can be produced by cell culture techniques, including the generation of monoclonal antibodies as described herein, or via transfection of antibody genes into suitable bacterial or mammalian cell hosts, in order to allow for the production of recombinant antibodies. In one technique, an immunogen comprising the polypeptide is initially injected into any of a wide variety of mammals (*e.g.*, mice, rats, rabbits,
20 sheep or goats). In this step, the polypeptides of this invention may serve as the immunogen without modification. Alternatively, particularly for relatively short polypeptides, a superior immune response may be elicited if the polypeptide is joined to a carrier protein, such as bovine serum albumin or keyhole limpet hemocyanin. The immunogen is injected into the animal host, preferably according to a predetermined
25 schedule incorporating one or more booster immunizations, and the animals are bled periodically. Polyclonal antibodies specific for the polypeptide may then be purified from such antisera by, for example, affinity chromatography using the polypeptide coupled to a suitable solid support.

Monoclonal antibodies specific for an antigenic polypeptide of interest
30 may be prepared, for example, using the technique of Kohler and Milstein, *Eur. J. Immunol.* 6:511-519, 1976, and improvements thereto. Briefly, these methods involve the preparation of immortal cell lines capable of producing antibodies having the

desired specificity (*i.e.*, reactivity with the polypeptide of interest). Such cell lines may be produced, for example, from spleen cells obtained from an animal immunized as described above. The spleen cells are then immortalized by, for example, fusion with a myeloma cell fusion partner, preferably one that is syngeneic with the immunized animal. A variety of fusion techniques may be employed. For example, the spleen cells and myeloma cells may be combined with a nonionic detergent for a few minutes and then plated at low density on a selective medium that supports the growth of hybrid cells, but not myeloma cells. A preferred selection technique uses HAT (hypoxanthine, aminopterin, thymidine) selection. After a sufficient time, usually about 1 to 2 weeks, colonies of hybrids are observed. Single colonies are selected and their culture supernatants tested for binding activity against the polypeptide. Hybridomas having high reactivity and specificity are preferred.

Monoclonal antibodies may be isolated from the supernatants of growing hybridoma colonies. In addition, various techniques may be employed to enhance the yield, such as injection of the hybridoma cell line into the peritoneal cavity of a suitable vertebrate host, such as a mouse. Monoclonal antibodies may then be harvested from the ascites fluid or the blood. Contaminants may be removed from the antibodies by conventional techniques, such as chromatography, gel filtration, precipitation, and extraction. The polypeptides of this invention may be used in the purification process in, for example, an affinity chromatography step.

Within certain embodiments, the use of antigen-binding fragments of antibodies may be preferred. Such fragments include Fab fragments, which may be prepared using standard techniques. Briefly, immunoglobulins may be purified from rabbit serum by affinity chromatography on Protein A bead columns (Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, 1988) and digested by papain to yield Fab and Fc fragments. The Fab and Fc fragments may be separated by affinity chromatography on protein A bead columns.

Monoclonal antibodies of the present invention may be coupled to one or more therapeutic agents. Suitable agents in this regard include radionuclides, differentiation inducers, drugs, toxins, and derivatives thereof. Preferred radionuclides include ^{90}Y , ^{123}I , ^{125}I , ^{131}I , ^{186}Re , ^{188}Re , ^{211}At , and ^{212}Bi . Preferred drugs include methotrexate, and pyrimidine and purine analogs. Preferred differentiation inducers

include phorbol esters and butyric acid. Preferred toxins include ricin, abrin, diphtheria toxin, cholera toxin, gelonin, *Pseudomonas* exotoxin, *Shigella* toxin, and pokeweed antiviral protein.

A therapeutic agent may be coupled (*e.g.*, covalently bonded) to a
5 suitable monoclonal antibody either directly or indirectly (*e.g.*, via a linker group). A direct reaction between an agent and an antibody is possible when each possesses a substituent capable of reacting with the other. For example, a nucleophilic group, such as an amino or sulfhydryl group, on one may be capable of reacting with a carbonyl-containing group, such as an anhydride or an acid halide, or with an alkyl group
10 containing a good leaving group (*e.g.*, a halide) on the other.

Alternatively, it may be desirable to couple a therapeutic agent and an antibody via a linker group. A linker group can function as a spacer to distance an antibody from an agent in order to avoid interference with binding capabilities. A linker group can also serve to increase the chemical reactivity of a substituent on an
15 agent or an antibody, and thus increase the coupling efficiency. An increase in chemical reactivity may also facilitate the use of agents, or functional groups on agents, which otherwise would not be possible.

It will be evident to those skilled in the art that a variety of bifunctional or polyfunctional reagents, both homo- and hetero-functional (such as those described
20 in the catalog of the Pierce Chemical Co., Rockford, IL), may be employed as the linker group. Coupling may be effected, for example, through amino groups, carboxyl groups, sulfhydryl groups or oxidized carbohydrate residues. There are numerous references describing such methodology, *e.g.*, U.S. Patent No. 4,671,958, to Rodwell *et al.*

Where a therapeutic agent is more potent when free from the antibody
25 portion of the immunoconjugates of the present invention, it may be desirable to use a linker group which is cleavable during or upon internalization into a cell. A number of different cleavable linker groups have been described. The mechanisms for the intracellular release of an agent from these linker groups include cleavage by reduction of a disulfide bond (*e.g.*, U.S. Patent No. 4,489,710, to Spitler), by irradiation of a
30 photolabile bond (*e.g.*, U.S. Patent No. 4,625,014, to Senter *et al.*), by hydrolysis of derivatized amino acid side chains (*e.g.*, U.S. Patent No. 4,638,045, to Kohn *et al.*), by

serum complement-mediated hydrolysis (*e.g.*, U.S. Patent No. 4,671,958, to Rodwell *et al.*), and acid-catalyzed hydrolysis (*e.g.*, U.S. Patent No. 4,569,789, to Blattler *et al.*).

It may be desirable to couple more than one agent to an antibody. In one embodiment, multiple molecules of an agent are coupled to one antibody molecule. In another embodiment, more than one type of agent may be coupled to one antibody. Regardless of the particular embodiment, immunoconjugates with more than one agent may be prepared in a variety of ways. For example, more than one agent may be coupled directly to an antibody molecule, or linkers that provide multiple sites for attachment can be used. Alternatively, a carrier can be used.

A carrier may bear the agents in a variety of ways, including covalent bonding either directly or via a linker group. Suitable carriers include proteins such as albumins (*e.g.*, U.S. Patent No. 4,507,234, to Kato *et al.*), peptides and polysaccharides such as aminodextran (*e.g.*, U.S. Patent No. 4,699,784, to Shih *et al.*). A carrier may also bear an agent by noncovalent bonding or by encapsulation, such as within a liposome vesicle (*e.g.*, U.S. Patent Nos. 4,429,008 and 4,873,088). Carriers specific for radionuclide agents include radiohalogenated small molecules and chelating compounds. For example, U.S. Patent No. 4,735,792 discloses representative radiohalogenated small molecules and their synthesis. A radionuclide chelate may be formed from chelating compounds that include those containing nitrogen and sulfur atoms as the donor atoms for binding the metal, or metal oxide, radionuclide. For example, U.S. Patent No. 4,673,562, to Davison *et al.* discloses representative chelating compounds and their synthesis.

A variety of routes of administration for the antibodies and immunoconjugates may be used. Typically, administration will be intravenous, intramuscular, subcutaneous or in the bed of a resected tumor. It will be evident that the precise dose of the antibody/immunoconjugate will vary depending upon the antibody used, the antigen density on the tumor, and the rate of clearance of the antibody.

T CELLS

Immunotherapeutic compositions may also, or alternatively, comprise T cells specific for an ovarian tumor protein. Such cells may generally be prepared *in*

vitro or *ex vivo*, using standard procedures. For example, T cells may be isolated from bone marrow, peripheral blood, or a fraction of bone marrow or peripheral blood of a patient, using a commercially available cell separation system, such as the Isolex™ System, available from Nexell Therapeutics, Inc. (Irvine, CA; see also U.S. Patent No. 5,240,856; U.S. Patent No. 5,215,926; WO 89/06280; WO 91/16116 and WO 92/07243). Alternatively, T cells may be derived from related or unrelated humans, non-human mammals, cell lines or cultures.

T cells may be stimulated with an ovarian tumor polypeptide, polynucleotide encoding an ovarian tumor polypeptide and/or an antigen presenting cell (APC) that expresses such a polypeptide. Such stimulation is performed under conditions and for a time sufficient to permit the generation of T cells that are specific for the polypeptide. Preferably, an ovarian tumor polypeptide or polynucleotide is present within a delivery vehicle, such as a microsphere, to facilitate the generation of specific T cells.

T cells are considered to be specific for an ovarian tumor polypeptide if the T cells specifically proliferate, secrete cytokines or kill target cells coated with the polypeptide or expressing a gene encoding the polypeptide. T cell specificity may be evaluated using any of a variety of standard techniques. For example, within a chromium release assay or proliferation assay, a stimulation index of more than two fold increase in lysis and/or proliferation, compared to negative controls, indicates T cell specificity. Such assays may be performed, for example, as described in Chen *et al.*, *Cancer Res.* 54:1065-1070, 1994. Alternatively, detection of the proliferation of T cells may be accomplished by a variety of known techniques. For example, T cell proliferation can be detected by measuring an increased rate of DNA synthesis (e.g., by pulse-labeling cultures of T cells with tritiated thymidine and measuring the amount of tritiated thymidine incorporated into DNA). Contact with an ovarian tumor polypeptide (100 ng/ml - 100 µg/ml, preferably 200 ng/ml - 25 µg/ml) for 3 - 7 days should result in at least a two fold increase in proliferation of the T cells. Contact as described above for 2-3 hours should result in activation of the T cells, as measured using standard cytokine assays in which a two fold increase in the level of cytokine release (e.g., TNF or IFN-γ) is indicative of T cell activation (see Coligan *et al.*, *Current Protocols in Immunology*, vol. 1, Wiley Interscience (Greene 1998)). T cells that have been

activated in response to an ovarian tumor polypeptide, polynucleotide or polypeptide-expressing APC may be CD4⁺ and/or CD8⁺. Ovarian tumor protein-specific T cells may be expanded using standard techniques. Within preferred embodiments, the T cells are derived from a patient, a related donor or an unrelated donor, and are administered
5 to the patient following stimulation and expansion.

For therapeutic purposes, CD4⁺ or CD8⁺ T cells that proliferate in response to an ovarian tumor polypeptide, polynucleotide or APC can be expanded in number either *in vitro* or *in vivo*. Proliferation of such T cells *in vitro* may be accomplished in a variety of ways. For example, the T cells can be re-exposed to an
10 ovarian tumor polypeptide, or a short peptide corresponding to an immunogenic portion of such a polypeptide, with or without the addition of T cell growth factors, such as interleukin-2, and/or stimulator cells that synthesize an ovarian tumor polypeptide. Alternatively, one or more T cells that proliferate in the presence of an ovarian tumor protein can be expanded in number by cloning. Methods for cloning cells are well
15 known in the art, and include limiting dilution.

PHARMACEUTICAL COMPOSITIONS

In additional embodiments, the present invention concerns formulation of one or more of the polynucleotide, polypeptide, T-cell and/or antibody compositions disclosed herein in pharmaceutically-acceptable solutions for administration to a cell or
20 an animal, either alone, or in combination with one or more other modalities of therapy.

It will also be understood that, if desired, the nucleic acid segment, RNA, DNA or PNA compositions that express a polypeptide as disclosed herein may be administered in combination with other agents as well, such as, *e.g.*, other proteins or
polypeptides or various pharmaceutically-active agents. In fact, there is virtually no
25 limit to other components that may also be included, given that the additional agents do not cause a significant adverse effect upon contact with the target cells or host tissues. The compositions may thus be delivered along with various other agents as required in the particular instance. Such compositions may be purified from host cells or other biological sources, or alternatively may be chemically synthesized as described herein.
30 Likewise, such compositions may further comprise substituted or derivatized RNA or DNA compositions.

Formulation of pharmaceutically-acceptable excipients and carrier solutions is well-known to those of skill in the art, as is the development of suitable dosing and treatment regimens for using the particular compositions described herein in a variety of treatment regimens, including *e.g.*, oral, parenteral, intravenous, intranasal, and intramuscular administration and formulation.

1. ORAL DELIVERY

In certain applications, the pharmaceutical compositions disclosed herein may be delivered *via* oral administration to an animal. As such, these compositions may be formulated with an inert diluent or with an assimilable edible carrier, or they may be enclosed in hard- or soft-shell gelatin capsule, or they may be compressed into tablets, or they may be incorporated directly with the food of the diet.

The active compounds may even be incorporated with excipients and used in the form of ingestible tablets, buccal tables, troches, capsules, elixirs, suspensions, syrups, wafers, and the like (Mathiowitz *et al.*, 1997; Hwang *et al.*, 1998; U. S. Patent 5,641,515; U. S. Patent 5,580,579 and U. S. Patent 5,792,451, each specifically incorporated herein by reference in its entirety). The tablets, troches, pills, capsules and the like may also contain the following: a binder, as gum tragacanth, acacia, cornstarch, or gelatin; excipients, such as dicalcium phosphate; a disintegrating agent, such as corn starch, potato starch, alginic acid and the like; a lubricant, such as magnesium stearate; and a sweetening agent, such as sucrose, lactose or saccharin may be added or a flavoring agent, such as peppermint, oil of wintergreen, or cherry flavoring. When the dosage unit form is a capsule, it may contain, in addition to materials of the above type, a liquid carrier. Various other materials may be present as coatings or to otherwise modify the physical form of the dosage unit. For instance, tablets, pills, or capsules may be coated with shellac, sugar, or both. A syrup or elixir may contain the active compound sucrose as a sweetening agent methyl and propylparabens as preservatives, a dye and flavoring, such as cherry or orange flavor. Of course, any material used in preparing any dosage unit form should be pharmaceutically pure and substantially non-toxic in the amounts employed. In addition, the active compounds may be incorporated into sustained-release preparation and formulations.

Typically, these formulations may contain at least about 0.1% of the active compound or more, although the percentage of the active ingredient(s) may, of course, be varied and may conveniently be between about 1 or 2% and about 60% or 70% or more of the weight or volume of the total formulation. Naturally, the amount of active compound(s) in each therapeutically useful composition may be prepared in such a way that a suitable dosage will be obtained in any given unit dose of the compound. Factors such as solubility, bioavailability, biological half-life, route of administration, product shelf life, as well as other pharmacological considerations will be contemplated by one skilled in the art of preparing such pharmaceutical formulations, and as such, a variety of dosages and treatment regimens may be desirable.

For oral administration the compositions of the present invention may alternatively be incorporated with one or more excipients in the form of a mouthwash, dentifrice, buccal tablet, oral spray, or sublingual orally-administered formulation. For example, a mouthwash may be prepared incorporating the active ingredient in the required amount in an appropriate solvent, such as a sodium borate solution (Dobell's Solution). Alternatively, the active ingredient may be incorporated into an oral solution such as one containing sodium borate, glycerin and potassium bicarbonate, or dispersed in a dentifrice, or added in a therapeutically-effective amount to a composition that may include water, binders, abrasives, flavoring agents, foaming agents, and humectants. Alternatively the compositions may be fashioned into a tablet or solution form that may be placed under the tongue or otherwise dissolved in the mouth.

2. INJECTABLE DELIVERY

In certain circumstances it will be desirable to deliver the pharmaceutical compositions disclosed herein parenterally, intravenously, intramuscularly, or even intraperitoneally as described in U. S. Patent 5,543,158; U. S. Patent 5,641,515 and U. S. Patent 5,399,363 (each specifically incorporated herein by reference in its entirety). Solutions of the active compounds as free base or pharmacologically acceptable salts may be prepared in water suitably mixed with a surfactant, such as hydroxypropylcellulose. Dispersions may also be prepared in glycerol, liquid polyethylene glycols, and mixtures thereof and in oils. Under ordinary conditions of

storage and use, these preparations contain a preservative to prevent the growth of microorganisms.

The pharmaceutical forms suitable for injectable use include sterile aqueous solutions or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersions (U. S. Patent 5,466,468, specifically incorporated herein by reference in its entirety). In all cases the form must be sterile and must be fluid to the extent that easy syringability exists. It must be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms, such as bacteria and fungi. The carrier can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (*e.g.*, glycerol, propylene glycol, and liquid polyethylene glycol, and the like), suitable mixtures thereof, and/or vegetable oils. Proper fluidity may be maintained, for example, by the use of a coating, such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of surfactants. The prevention of the action of microorganisms can be facilitated by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, sorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars or sodium chloride. Prolonged absorption of the injectable compositions can be brought about by the use in the compositions of agents delaying absorption, for example, aluminum monostearate and gelatin.

For parenteral administration in an aqueous solution, for example, the solution should be suitably buffered if necessary and the liquid diluent first rendered isotonic with sufficient saline or glucose. These particular aqueous solutions are especially suitable for intravenous, intramuscular, subcutaneous and intraperitoneal administration. In this connection, a sterile aqueous medium that can be employed will be known to those of skill in the art in light of the present disclosure. For example, one dosage may be dissolved in 1 ml of isotonic NaCl solution and either added to 1000 ml of hypodermoclysis fluid or injected at the proposed site of infusion, (see for example, "Remington's Pharmaceutical Sciences" 15th Edition, pages 1035-1038 and 1570-1580). Some variation in dosage will necessarily occur depending on the condition of the subject being treated. The person responsible for administration will, in any event, determine the appropriate dose for the individual subject. Moreover, for human

administration, preparations should meet sterility, pyrogenicity, and the general safety and purity standards as required by FDA Office of Biologics standards.

Sterile injectable solutions are prepared by incorporating the active compounds in the required amount in the appropriate solvent with various of the other ingredients enumerated above, as required, followed by filtered sterilization. Generally, 5 dispersions are prepared by incorporating the various sterilized active ingredients into a sterile vehicle which contains the basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation of sterile injectable solutions, the preferred methods of preparation are 10 vacuum-drying and freeze-drying techniques which yield a powder of the active ingredient plus any additional desired ingredient from a previously sterile-filtered solution thereof.

The compositions disclosed herein may be formulated in a neutral or salt form. Pharmaceutically-acceptable salts, include the acid addition salts (formed with 15 the free amino groups of the protein) and which are formed with inorganic acids such as, for example, hydrochloric or phosphoric acids, or such organic acids as acetic, oxalic, tartaric, mandelic, and the like. Salts formed with the free carboxyl groups can also be derived from inorganic bases such as, for example, sodium, potassium, ammonium, calcium, or ferric hydroxides, and such organic bases as isopropylamine, 20 trimethylamine, histidine, procaine and the like. Upon formulation, solutions will be administered in a manner compatible with the dosage formulation and in such amount as is therapeutically effective. The formulations are easily administered in a variety of dosage forms such as injectable solutions, drug-release capsules, and the like.

As used herein, "carrier" includes any and all solvents, dispersion media, 25 vehicles, coatings, diluents, antibacterial and antifungal agents, isotonic and absorption delaying agents, buffers, carrier solutions, suspensions, colloids, and the like. The use of such media and agents for pharmaceutical active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active ingredient, its use in the therapeutic compositions is contemplated. Supplementary 30 active ingredients can also be incorporated into the compositions.

The phrase "pharmaceutically-acceptable" refers to molecular entities and compositions that do not produce an allergic or similar untoward reaction when

administered to a human. The preparation of an aqueous composition that contains a protein as an active ingredient is well understood in the art. Typically, such compositions are prepared as injectables, either as liquid solutions or suspensions; solid forms suitable for solution in, or suspension in, liquid prior to injection can also be prepared. The preparation can also be emulsified.

3. NASAL DELIVERY

In certain embodiments, the pharmaceutical compositions may be delivered by intranasal sprays, inhalation, and/or other aerosol delivery vehicles. Methods for delivering genes, nucleic acids, and peptide compositions directly to the lungs *via* nasal aerosol sprays has been described *e.g.*, in U. S. Patent 5,756,353 and U. S. Patent 5,804,212 (each specifically incorporated herein by reference in its entirety). Likewise, the delivery of drugs using intranasal microparticle resins (Takenaga *et al.*, 1998) and lysophosphatidyl-glycerol compounds (U. S. Patent 5,725,871, specifically incorporated herein by reference in its entirety) are also well-known in the pharmaceutical arts. Likewise, transmucosal drug delivery in the form of a polytetrafluoroethylene support matrix is described in U. S. Patent 5,780,045 (specifically incorporated herein by reference in its entirety).

4. LIPOSOME-, NANOCAPSULE-, AND MICROPARTICLE-MEDIATED DELIVERY

In certain embodiments, the inventors contemplate the use of liposomes, nanocapsules, microparticles, microspheres, lipid particles, vesicles, and the like, for the introduction of the compositions of the present invention into suitable host cells. In particular, the compositions of the present invention may be formulated for delivery either encapsulated in a lipid particle, a liposome, a vesicle, a nanosphere, or a nanoparticle or the like.

Such formulations may be preferred for the introduction of pharmaceutically-acceptable formulations of the nucleic acids or constructs disclosed herein. The formation and use of liposomes is generally known to those of skill in the art (see for example, Couvreur *et al.*, 1977; Couvreur, 1988; Lasic, 1998; which describes the use of liposomes and nanocapsules in the targeted antibiotic therapy for intracellular bacterial infections and diseases). Recently, liposomes were developed

with improved serum stability and circulation half-times (Gabizon and Papahadjopoulos, 1988; Allen and Choun, 1987; U. S. Patent 5,741,516, specifically incorporated herein by reference in its entirety). Further, various methods of liposome and liposome like preparations as potential drug carriers have been reviewed (Takakura,
5 1998; Chandran *et al.*, 1997; Margalit, 1995; U. S. Patent 5,567,434; U. S. Patent 5,552,157; U. S. Patent 5,565,213; U. S. Patent 5,738,868 and U. S. Patent 5,795,587, each specifically incorporated herein by reference in its entirety).

Liposomes have been used successfully with a number of cell types that are normally resistant to transfection by other procedures including T cell suspensions,
10 primary hepatocyte cultures and PC 12 cells (Renneisen *et al.*, 1990; Muller *et al.*, 1990). In addition, liposomes are free of the DNA length constraints that are typical of viral-based delivery systems. Liposomes have been used effectively to introduce genes, drugs (Heath and Martin, 1986; Heath *et al.*, 1986; Balazsovits *et al.*, 1989; Fresta and Puglisi, 1996), radiotherapeutic agents (Pikul *et al.*, 1987), enzymes (Imaizumi *et al.*,
15 1990a; Imaizumi *et al.*, 1990b), viruses (Faller and Baltimore, 1984), transcription factors and allosteric effectors (Nicolau and Gersonde, 1979) into a variety of cultured cell lines and animals. In addition, several successful clinical trials examining the effectiveness of liposome-mediated drug delivery have been completed (Lopez-Berestein *et al.*, 1985a; 1985b; Coune, 1988; Sculier *et al.*, 1988). Furthermore, several
20 studies suggest that the use of liposomes is not associated with autoimmune responses, toxicity or gonadal localization after systemic delivery (Mori and Fukatsu, 1992).

Liposomes are formed from phospholipids that are dispersed in an aqueous medium and spontaneously form multilamellar concentric bilayer vesicles (also termed multilamellar vesicles (MLVs). MLVs generally have diameters of from
25 25 nm to 4 μ m. Sonication of MLVs results in the formation of small unilamellar vesicles (SUVs) with diameters in the range of 200 to 500 Å, containing an aqueous solution in the core.

Liposomes bear resemblance to cellular membranes and are contemplated for use in connection with the present invention as carriers for the peptide
30 compositions. They are widely suitable as both water- and lipid-soluble substances can be entrapped, *i.e.* in the aqueous spaces and within the bilayer itself, respectively. It is

possible that the drug-bearing liposomes may even be employed for site-specific delivery of active agents by selectively modifying the liposomal formulation.

In addition to the teachings of Couvreur *et al.* (1977; 1988), the following information may be utilized in generating liposomal formulations.

5 Phospholipids can form a variety of structures other than liposomes when dispersed in water, depending on the molar ratio of lipid to water. At low ratios the liposome is the preferred structure. The physical characteristics of liposomes depend on pH, ionic strength and the presence of divalent cations. Liposomes can show low permeability to ionic and polar substances, but at elevated temperatures undergo a phase transition
10 which markedly alters their permeability. The phase transition involves a change from a closely packed, ordered structure, known as the gel state, to a loosely packed, less-ordered structure, known as the fluid state. This occurs at a characteristic phase-transition temperature and results in an increase in permeability to ions, sugars and drugs.

15 In addition to temperature, exposure to proteins can alter the permeability of liposomes. Certain soluble proteins, such as cytochrome c, bind, deform and penetrate the bilayer, thereby causing changes in permeability. Cholesterol inhibits this penetration of proteins, apparently by packing the phospholipids more tightly. It is contemplated that the most useful liposome formations for antibiotic and
20 inhibitor delivery will contain cholesterol.

 The ability to trap solutes varies between different types of liposomes. For example, MLVs are moderately efficient at trapping solutes, but SUVs are extremely inefficient. SUVs offer the advantage of homogeneity and reproducibility in size distribution, however, and a compromise between size and trapping efficiency is
25 offered by large unilamellar vesicles (LUVs). These are prepared by ether evaporation and are three to four times more efficient at solute entrapment than MLVs.

 In addition to liposome characteristics, an important determinant in entrapping compounds is the physicochemical properties of the compound itself. Polar compounds are trapped in the aqueous spaces and nonpolar compounds bind to the lipid
30 bilayer of the vesicle. Polar compounds are released through permeation or when the bilayer is broken, but nonpolar compounds remain affiliated with the bilayer unless it is

disrupted by temperature or exposure to lipoproteins. Both types show maximum efflux rates at the phase transition temperature.

Liposomes interact with cells *via* four different mechanisms: endocytosis by phagocytic cells of the reticuloendothelial system such as macrophages and neutrophils; adsorption to the cell surface, either by nonspecific weak hydrophobic or electrostatic forces, or by specific interactions with cell-surface components; fusion with the plasma cell membrane by insertion of the lipid bilayer of the liposome into the plasma membrane, with simultaneous release of liposomal contents into the cytoplasm; and by transfer of liposomal lipids to cellular or subcellular membranes, or vice versa, without any association of the liposome contents. It often is difficult to determine which mechanism is operative and more than one may operate at the same time.

The fate and disposition of intravenously injected liposomes depend on their physical properties, such as size, fluidity, and surface charge. They may persist in tissues for h or days, depending on their composition, and half lives in the blood range from min to several h. Larger liposomes, such as MLVs and LUVs, are taken up rapidly by phagocytic cells of the reticuloendothelial system, but physiology of the circulatory system restrains the exit of such large species at most sites. They can exit only in places where large openings or pores exist in the capillary endothelium, such as the sinusoids of the liver or spleen. Thus, these organs are the predominate site of uptake. On the other hand, SUVs show a broader tissue distribution but still are sequestered highly in the liver and spleen. In general, this *in vivo* behavior limits the potential targeting of liposomes to only those organs and tissues accessible to their large size. These include the blood, liver, spleen, bone marrow, and lymphoid organs.

Targeting is generally not a limitation in terms of the present invention. However, should specific targeting be desired, methods are available for this to be accomplished. Antibodies may be used to bind to the liposome surface and to direct the antibody and its drug contents to specific antigenic receptors located on a particular cell-type surface. Carbohydrate determinants (glycoprotein or glycolipid cell-surface components that play a role in cell-cell recognition, interaction and adhesion) may also be used as recognition sites as they have potential in directing liposomes to particular cell types. Mostly, it is contemplated that intravenous injection of liposomal preparations would be used, but other routes of administration are also conceivable.

Alternatively, the invention provides for pharmaceutically-acceptable nanocapsule formulations of the compositions of the present invention. Nanocapsules can generally entrap compounds in a stable and reproducible way (Henry-Michelland *et al.*, 1987; Quintanar-Guerrero *et al.*, 1998; Douglas *et al.*, 1987). To avoid side effects due to intracellular polymeric overloading, such ultrafine particles (sized around 0.1 μm) should be designed using polymers able to be degraded *in vivo*. Biodegradable polyalkyl-cyanoacrylate nanoparticles that meet these requirements are contemplated for use in the present invention. Such particles may be easily made, as described (Couvreur *et al.*, 1980; 1988; zur Muhlen *et al.*, 1998; Zambaux *et al.* 1998; Pinto-Alphandry *et al.*, 1995 and U. S. Patent 5,145,684, specifically incorporated herein by reference in its entirety).

VACCINES

In certain preferred embodiments of the present invention, vaccines are provided. The vaccines will generally comprise one or more pharmaceutical compositions, such as those discussed above, in combination with an immunostimulant. An immunostimulant may be any substance that enhances or potentiates an immune response (antibody and/or cell-mediated) to an exogenous antigen. Examples of immunostimulants include adjuvants, biodegradable microspheres (*e.g.*, polylactic galactide) and liposomes (into which the compound is incorporated; *see e.g.*, Fullerton, U.S. Patent No. 4,235,877). Vaccine preparation is generally described in, for example, M.F. Powell and M.J. Newman, eds., "Vaccine Design (the subunit and adjuvant approach)," Plenum Press (NY, 1995). Pharmaceutical compositions and vaccines within the scope of the present invention may also contain other compounds, which may be biologically active or inactive. For example, one or more immunogenic portions of other tumor antigens may be present, either incorporated into a fusion polypeptide or as a separate compound, within the composition or vaccine.

Illustrative vaccines may contain DNA encoding one or more of the polypeptides as described above, such that the polypeptide is generated *in situ*. As noted above, the DNA may be present within any of a variety of delivery systems known to those of ordinary skill in the art, including nucleic acid expression systems, bacteria and viral expression systems. Numerous gene delivery techniques are well

known in the art, such as those described by Rolland, *Crit. Rev. Therap. Drug Carrier Systems* 15:143-198, 1998, and references cited therein. Appropriate nucleic acid expression systems contain the necessary DNA sequences for expression in the patient (such as a suitable promoter and terminating signal). Bacterial delivery systems involve the administration of a bacterium (such as *Bacillus-Calmette-Guerrin*) that expresses an immunogenic portion of the polypeptide on its cell surface or secretes such an epitope. In a preferred embodiment, the DNA may be introduced using a viral expression system (e.g., vaccinia or other pox virus, retrovirus, or adenovirus), which may involve the use of a non-pathogenic (defective), replication competent virus. Suitable systems are disclosed, for example, in Fisher-Hoch *et al.*, *Proc. Natl. Acad. Sci. USA* 86:317-321, 1989; Flexner *et al.*, *Ann. N.Y. Acad. Sci.* 569:86-103, 1989; Flexner *et al.*, *Vaccine* 8:17-21, 1990; U.S. Patent Nos. 4,603,112, 4,769,330, and 5,017,487; WO 89/01973; U.S. Patent No. 4,777,127; GB 2,200,651; EP 0,345,242; WO 91/02805; Berkner, *Biotechniques* 6:616-627, 1988; Rosenfeld *et al.*, *Science* 252:431-434, 1991; Kolls *et al.*, *Proc. Natl. Acad. Sci. USA* 91:215-219, 1994; Kass-Eisler *et al.*, *Proc. Natl. Acad. Sci. USA* 90:11498-11502, 1993; Guzman *et al.*, *Circulation* 88:2838-2848, 1993; and Guzman *et al.*, *Cir. Res.* 73:1202-1207, 1993. Techniques for incorporating DNA into such expression systems are well known to those of ordinary skill in the art. The DNA may also be "naked," as described, for example, in Ulmer *et al.*, *Science* 259:1745-1749, 1993 and reviewed by Cohen, *Science* 259:1691-1692, 1993. The uptake of naked DNA may be increased by coating the DNA onto biodegradable beads, which are efficiently transported into the cells. It will be apparent that a vaccine may comprise both a polynucleotide and a polypeptide component. Such vaccines may provide for an enhanced immune response.

It will be apparent that a vaccine may contain pharmaceutically acceptable salts of the polynucleotides and polypeptides provided herein. Such salts may be prepared from pharmaceutically acceptable non-toxic bases, including organic bases (e.g., salts of primary, secondary and tertiary amines and basic amino acids) and inorganic bases (e.g., sodium, potassium, lithium, ammonium, calcium and magnesium salts).

While any suitable carrier known to those of ordinary skill in the art may be employed in the vaccine compositions of this invention, the type of carrier will vary

depending on the mode of administration. Compositions of the present invention may be formulated for any appropriate manner of administration, including for example, topical, oral, nasal, intravenous, intracranial, intraperitoneal, subcutaneous or intramuscular administration. For parenteral administration, such as subcutaneous
5 injection, the carrier preferably comprises water, saline, alcohol, a fat, a wax or a buffer. For oral administration, any of the above carriers or a solid carrier, such as mannitol, lactose, starch, magnesium stearate, sodium saccharine, talcum, cellulose, glucose, sucrose, and magnesium carbonate, may be employed. Biodegradable microspheres (e.g., polylactate polyglycolate) may also be employed as carriers for the
10 pharmaceutical compositions of this invention. Suitable biodegradable microspheres are disclosed, for example, in U.S. Patent Nos. 4,897,268; 5,075,109; 5,928,647; 5,811,128; 5,820,883; 5,853,763; 5,814,344 and 5,942,252. One may also employ a carrier comprising the particulate-protein complexes described in U.S. Patent No. 5,928,647, which are capable of inducing a class I-restricted cytotoxic T lymphocyte
15 responses in a host.

Such compositions may also comprise buffers (e.g., neutral buffered saline or phosphate buffered saline), carbohydrates (e.g., glucose, mannose, sucrose or dextrans), mannitol, proteins, polypeptides or amino acids such as glycine, antioxidants, bacteriostats, chelating agents such as EDTA or glutathione, adjuvants (e.g., aluminum
20 hydroxide), solutes that render the formulation isotonic, hypotonic or weakly hypertonic with the blood of a recipient, suspending agents, thickening agents and/or preservatives. Alternatively, compositions of the present invention may be formulated as a lyophilizate. Compounds may also be encapsulated within liposomes using well known technology.

25 Any of a variety of immunostimulants may be employed in the vaccines of this invention. For example, an adjuvant may be included. Most adjuvants contain a substance designed to protect the antigen from rapid catabolism, such as aluminum hydroxide or mineral oil, and a stimulator of immune responses, such as lipid A, *Bordetella pertussis* or *Mycobacterium tuberculosis* derived proteins. Suitable
30 adjuvants are commercially available as, for example, Freund's Incomplete Adjuvant and Complete Adjuvant (Difco Laboratories, Detroit, MI); Merck Adjuvant 65 (Merck and Company, Inc., Rahway, NJ); AS-2 (SmithKline Beecham, Philadelphia, PA);

aluminum salts such as aluminum hydroxide gel (alum) or aluminum phosphate; salts of calcium, iron or zinc; an insoluble suspension of acylated tyrosine; acylated sugars; cationically or anionically derivatized polysaccharides; polyphosphazenes; biodegradable microspheres; monophosphoryl lipid A and quil A. Cytokines, such as
5 GM-CSF or interleukin-2, -7, or -12, may also be used as adjuvants.

Within the vaccines provided herein, the adjuvant composition is preferably designed to induce an immune response predominantly of the Th1 type. High levels of Th1-type cytokines (e.g., IFN- γ , TNF α , IL-2 and IL-12) tend to favor the induction of cell mediated immune responses to an administered antigen. In contrast,
10 high levels of Th2-type cytokines (e.g., IL-4, IL-5, IL-6 and IL-10) tend to favor the induction of humoral immune responses. Following application of a vaccine as provided herein, a patient will support an immune response that includes Th1- and Th2-type responses. Within a preferred embodiment, in which a response is predominantly Th1-type, the level of Th1-type cytokines will increase to a greater extent than the level
15 of Th2-type cytokines. The levels of these cytokines may be readily assessed using standard assays. For a review of the families of cytokines, see Mosmann and Coffman, *Ann. Rev. Immunol.* 7:145-173, 1989.

Preferred adjuvants for use in eliciting a predominantly Th1-type response include, for example, a combination of monophosphoryl lipid A, preferably 3-
20 de-O-acylated monophosphoryl lipid A (3D-MPL), together with an aluminum salt. MPL adjuvants are available from Corixa Corporation (Seattle, WA; see US Patent Nos. 4,436,727; 4,877,611; 4,866,034 and 4,912,094). CpG-containing oligonucleotides (in which the CpG dinucleotide is unmethylated) also induce a predominantly Th1 response. Such oligonucleotides are well known and are described,
25 for example, in WO 96/02555, WO 99/33488 and U.S. Patent Nos. 6,008,200 and 5,856,462. Immunostimulatory DNA sequences are also described, for example, by Sato *et al.*, *Science* 273:352, 1996. Another preferred adjuvant is a saponin, preferably QS21 (Aquila Biopharmaceuticals Inc., Framingham, MA), which may be used alone or in combination with other adjuvants. For example, an enhanced system involves the
30 combination of a monophosphoryl lipid A and saponin derivative, such as the combination of QS21 and 3D-MPL as described in WO 94/00153, or a less reactogenic composition where the QS21 is quenched with cholesterol, as described in WO

96/33739. Other preferred formulations comprise an oil-in-water emulsion and tocopherol. A particularly potent adjuvant formulation involving QS21, 3D-MPL and tocopherol in an oil-in-water emulsion is described in WO 95/17210.

Other preferred adjuvants include Montanide ISA 720 (Seppic, France),
5 SAF (Chiron, California, United States), ISCOMS (CSL), MF-59 (Chiron), the SBAS series of adjuvants (*e.g.*, SBAS-2 or SBAS-4, available from SmithKline Beecham, Rixensart, Belgium), Detox (Corixa, Hamilton, MT), RC-529 (Corixa, Hamilton, MT) and other aminoalkyl glucosaminide 4-phosphates (AGPs), such as those described in pending U.S. Patent Application Serial Nos. 08/853,826 and 09/074,720, the disclosures
10 of which are incorporated herein by reference in their entireties.

Any vaccine provided herein may be prepared using well known methods that result in a combination of antigen, immune response enhancer and a suitable carrier or excipient. The compositions described herein may be administered as part of a sustained release formulation (*i.e.*, a formulation such as a capsule, sponge
15 or gel (composed of polysaccharides, for example) that effects a slow release of compound following administration). Such formulations may generally be prepared using well known technology (*see, e.g.*, Coombes *et al.*, *Vaccine* 14:1429-1438, 1996) and administered by, for example, oral, rectal or subcutaneous implantation, or by implantation at the desired target site. Sustained-release formulations may contain a
20 polypeptide, polynucleotide or antibody dispersed in a carrier matrix and/or contained within a reservoir surrounded by a rate controlling membrane.

Carriers for use within such formulations are biocompatible, and may also be biodegradable; preferably the formulation provides a relatively constant level of active component release. Such carriers include microparticles of poly(lactide-co-
25 glycolide), polyacrylate, latex, starch, cellulose, dextran and the like. Other delayed-release carriers include supramolecular biovectors, which comprise a non-liquid hydrophilic core (*e.g.*, a cross-linked polysaccharide or oligosaccharide) and, optionally, an external layer comprising an amphiphilic compound, such as a phospholipid (*see e.g.*, U.S. Patent No. 5,151,254 and PCT applications WO 94/20078,
30 WO/94/23701 and WO 96/06638). The amount of active compound contained within a sustained release formulation depends upon the site of implantation, the rate and expected duration of release and the nature of the condition to be treated or prevented.

Any of a variety of delivery vehicles may be employed within pharmaceutical compositions and vaccines to facilitate production of an antigen-specific immune response that targets tumor cells. Delivery vehicles include antigen presenting cells (APCs), such as dendritic cells, macrophages, B cells, monocytes and
5 other cells that may be engineered to be efficient APCs. Such cells may, but need not, be genetically modified to increase the capacity for presenting the antigen, to improve activation and/or maintenance of the T cell response, to have anti-tumor effects *per se* and/or to be immunologically compatible with the receiver (*i.e.*, matched HLA haplotype). APCs may generally be isolated from any of a variety of biological fluids
10 and organs, including tumor and peritumoral tissues, and may be autologous, allogeneic, syngeneic or xenogeneic cells.

Certain preferred embodiments of the present invention use dendritic cells or progenitors thereof as antigen-presenting cells. Dendritic cells are highly potent APCs (Banchereau and Steinman, *Nature* 392:245-251, 1998) and have been shown to
15 be effective as a physiological adjuvant for eliciting prophylactic or therapeutic antitumor immunity (*see* Timmerman and Levy, *Ann. Rev. Med.* 50:507-529, 1999). In general, dendritic cells may be identified based on their typical shape (stellate *in situ*, with marked cytoplasmic processes (dendrites) visible *in vitro*), their ability to take up, process and present antigens with high efficiency and their ability to activate naïve T
20 cell responses. Dendritic cells may, of course, be engineered to express specific cell-surface receptors or ligands that are not commonly found on dendritic cells *in vivo* or *ex vivo*, and such modified dendritic cells are contemplated by the present invention. As an alternative to dendritic cells, secreted vesicles antigen-loaded dendritic cells (called exosomes) may be used within a vaccine (*see* Zitvogel *et al.*, *Nature Med.* 4:594-600,
25 1998).

Dendritic cells and progenitors may be obtained from peripheral blood, bone marrow, tumor-infiltrating cells, peritumoral tissues-infiltrating cells, lymph nodes, spleen, skin, umbilical cord blood or any other suitable tissue or fluid. For example, dendritic cells may be differentiated *ex vivo* by adding a combination of
30 cytokines such as GM-CSF, IL-4, IL-13 and/or TNF α to cultures of monocytes harvested from peripheral blood. Alternatively, CD34 positive cells harvested from peripheral blood, umbilical cord blood or bone marrow may be differentiated into

dendritic cells by adding to the culture medium combinations of GM-CSF, IL-3, TNF α , CD40 ligand, LPS, flt3 ligand and/or other compound(s) that induce differentiation, maturation and proliferation of dendritic cells.

Dendritic cells are conveniently categorized as "immature" and "mature" cells, which allows a simple way to discriminate between two well characterized phenotypes. However, this nomenclature should not be construed to exclude all possible intermediate stages of differentiation. Immature dendritic cells are characterized as APC with a high capacity for antigen uptake and processing, which correlates with the high expression of Fc γ receptor and mannose receptor. The mature phenotype is typically characterized by a lower expression of these markers, but a high expression of cell surface molecules responsible for T cell activation such as class I and class II MHC, adhesion molecules (*e.g.*, CD54 and CD11) and costimulatory molecules (*e.g.*, CD40, CD80, CD86 and 4-1BB).

APCs may generally be transfected with a polynucleotide encoding an ovarian tumor protein (or portion or other variant thereof) such that the ovarian tumor polypeptide, or an immunogenic portion thereof, is expressed on the cell surface. Such transfection may take place *ex vivo*, and a composition or vaccine comprising such transfected cells may then be used for therapeutic purposes, as described herein. Alternatively, a gene delivery vehicle that targets a dendritic or other antigen presenting cell may be administered to a patient, resulting in transfection that occurs *in vivo*. *In vivo* and *ex vivo* transfection of dendritic cells, for example, may generally be performed using any methods known in the art, such as those described in WO 97/24447, or the gene gun approach described by Mahvi *et al.*, *Immunology and cell Biology* 75:456-460, 1997. Antigen loading of dendritic cells may be achieved by incubating dendritic cells or progenitor cells with the ovarian tumor polypeptide, DNA (naked or within a plasmid vector) or RNA; or with antigen-expressing recombinant bacterium or viruses (*e.g.*, vaccinia, fowlpox, adenovirus or lentivirus vectors). Prior to loading, the polypeptide may be covalently conjugated to an immunological partner that provides T cell help (*e.g.*, a carrier molecule). Alternatively, a dendritic cell may be pulsed with a non-conjugated immunological partner, separately or in the presence of the polypeptide.

Vaccines and pharmaceutical compositions may be presented in unit-dose or multi-dose containers, such as sealed ampoules or vials. Such containers are preferably hermetically sealed to preserve sterility of the formulation until use. In general, formulations may be stored as suspensions, solutions or emulsions in oily or aqueous vehicles. Alternatively, a vaccine or pharmaceutical composition may be stored in a freeze-dried condition requiring only the addition of a sterile liquid carrier immediately prior to use.

CANCER THERAPY

In further aspects of the present invention, the compositions described herein may be used for immunotherapy of cancer, such as ovarian cancer. Within such methods, pharmaceutical compositions and vaccines are typically administered to a patient. As used herein, a "patient" refers to any warm-blooded animal, preferably a human. A patient may or may not be afflicted with cancer. Accordingly, the above pharmaceutical compositions and vaccines may be used to prevent the development of a cancer or to treat a patient afflicted with a cancer. A cancer may be diagnosed using criteria generally accepted in the art, including the presence of a malignant tumor. Pharmaceutical compositions and vaccines may be administered either prior to or following surgical removal of primary tumors and/or treatment such as administration of radiotherapy or conventional chemotherapeutic drugs. Administration may be by any suitable method, including administration by intravenous, intraperitoneal, intramuscular, subcutaneous, intranasal, intradermal, anal, vaginal, topical and oral routes.

Within certain embodiments, immunotherapy may be active immunotherapy, in which treatment relies on the *in vivo* stimulation of the endogenous host immune system to react against tumors with the administration of immune response-modifying agents (such as polypeptides and polynucleotides as provided herein).

Within other embodiments, immunotherapy may be passive immunotherapy, in which treatment involves the delivery of agents with established tumor-immune reactivity (such as effector cells or antibodies) that can directly or indirectly mediate antitumor effects and does not necessarily depend on an intact host

immune system. Examples of effector cells include T cells as discussed above, T lymphocytes (such as CD8⁺ cytotoxic T lymphocytes and CD4⁺ T-helper tumor-infiltrating lymphocytes), killer cells (such as Natural Killer cells and lymphokine-activated killer cells), B cells and antigen-presenting cells (such as dendritic cells and
5 macrophages) expressing a polypeptide provided herein. T cell receptors and antibody receptors specific for the polypeptides recited herein may be cloned, expressed and transferred into other vectors or effector cells for adoptive immunotherapy. The polypeptides provided herein may also be used to generate antibodies or anti-idiotypic antibodies (as described above and in U.S. Patent No. 4,918,164) for passive
10 immunotherapy.

Effector cells may generally be obtained in sufficient quantities for adoptive immunotherapy by growth *in vitro*, as described herein. Culture conditions for expanding single antigen-specific effector cells to several billion in number with retention of antigen recognition *in vivo* are well known in the art. Such *in vitro* culture
15 conditions typically use intermittent stimulation with antigen, often in the presence of cytokines (such as IL-2) and non-dividing feeder cells. As noted above, immunoreactive polypeptides as provided herein may be used to rapidly expand antigen-specific T cell cultures in order to generate a sufficient number of cells for immunotherapy. In particular, antigen-presenting cells, such as dendritic, macrophage,
20 monocyte, fibroblast and/or B cells, may be pulsed with immunoreactive polypeptides or transfected with one or more polynucleotides using standard techniques well known in the art. For example, antigen-presenting cells can be transfected with a polynucleotide having a promoter appropriate for increasing expression in a recombinant virus or other expression system. Cultured effector cells for use in therapy
25 must be able to grow and distribute widely, and to survive long term *in vivo*. Studies have shown that cultured effector cells can be induced to grow *in vivo* and to survive long term in substantial numbers by repeated stimulation with antigen supplemented with IL-2 (*see, for example, Cheever et al., Immunological Reviews 157:177, 1997*).

Alternatively, a vector expressing a polypeptide recited herein may be
30 introduced into antigen presenting cells taken from a patient and clonally propagated *ex vivo* for transplant back into the same patient. Transfected cells may be reintroduced

into the patient using any means known in the art, preferably in sterile form by intravenous, intracavitary, intraperitoneal or intratumor administration.

Routes and frequency of administration of the therapeutic compositions described herein, as well as dosage, will vary from individual to individual, and may be readily established using standard techniques. In general, the pharmaceutical compositions and vaccines may be administered by injection (*e.g.*, intracutaneous, intramuscular, intravenous or subcutaneous), intranasally (*e.g.*, by aspiration) or orally. Preferably, between 1 and 10 doses may be administered over a 52 week period. Preferably, 6 doses are administered, at intervals of 1 month, and booster vaccinations may be given periodically thereafter. Alternate protocols may be appropriate for individual patients. A suitable dose is an amount of a compound that, when administered as described above, is capable of promoting an anti-tumor immune response, and is at least 10-50% above the basal (*i.e.*, untreated) level. Such response can be monitored by measuring the anti-tumor antibodies in a patient or by vaccine-dependent generation of cytolytic effector cells capable of killing the patient's tumor cells *in vitro*. Such vaccines should also be capable of causing an immune response that leads to an improved clinical outcome (*e.g.*, more frequent remissions, complete or partial or longer disease-free survival) in vaccinated patients as compared to non-vaccinated patients. In general, for pharmaceutical compositions and vaccines comprising one or more polypeptides, the amount of each polypeptide present in a dose ranges from about 25 μ g to 5 mg per kg of host. Suitable dose sizes will vary with the size of the patient, but will typically range from about 0.1 mL to about 5 mL.

In general, an appropriate dosage and treatment regimen provides the active compound(s) in an amount sufficient to provide therapeutic and/or prophylactic benefit. Such a response can be monitored by establishing an improved clinical outcome (*e.g.*, more frequent remissions, complete or partial, or longer disease-free survival) in treated patients as compared to non-treated patients. Increases in preexisting immune responses to an ovarian tumor protein generally correlate with an improved clinical outcome. Such immune responses may generally be evaluated using standard proliferation, cytotoxicity or cytokine assays, which may be performed using samples obtained from a patient before and after treatment.

CANCER DETECTION AND DIAGNOSIS

In general, a cancer may be detected in a patient based on the presence of one or more ovarian tumor proteins and/or polynucleotides encoding such proteins in a biological sample (for example, blood, sera, sputum urine and/or tumor biopsies) obtained from the patient. In other words, such proteins may be used as markers to indicate the presence or absence of a cancer such as ovarian cancer. In addition, such proteins may be useful for the detection of other cancers. The binding agents provided herein generally permit detection of the level of antigen that binds to the agent in the biological sample. Polynucleotide primers and probes may be used to detect the level of mRNA encoding a tumor protein, which is also indicative of the presence or absence of a cancer. In general, an ovarian tumor sequence should be present at a level that is at least three fold higher in tumor tissue than in normal tissue

There are a variety of assay formats known to those of ordinary skill in the art for using a binding agent to detect polypeptide markers in a sample. *See, e.g.,* Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, 1988. In general, the presence or absence of a cancer in a patient may be determined by (a) contacting a biological sample obtained from a patient with a binding agent; (b) detecting in the sample a level of polypeptide that binds to the binding agent; and (c) comparing the level of polypeptide with a predetermined cut-off value.

In a preferred embodiment, the assay involves the use of binding agent immobilized on a solid support to bind to and remove the polypeptide from the remainder of the sample. The bound polypeptide may then be detected using a detection reagent that contains a reporter group and specifically binds to the binding agent/polypeptide complex. Such detection reagents may comprise, for example, a binding agent that specifically binds to the polypeptide or an antibody or other agent that specifically binds to the binding agent, such as an anti-immunoglobulin, protein G, protein A or a lectin. Alternatively, a competitive assay may be utilized, in which a polypeptide is labeled with a reporter group and allowed to bind to the immobilized binding agent after incubation of the binding agent with the sample. The extent to which components of the sample inhibit the binding of the labeled polypeptide to the binding agent is indicative of the reactivity of the sample with the immobilized binding agent. Suitable polypeptides for use within such assays include full length ovarian

tumor proteins and portions thereof to which the binding agent binds, as described above.

The solid support may be any material known to those of ordinary skill in the art to which the tumor protein may be attached. For example, the solid support
5 may be a test well in a microtiter plate or a nitrocellulose or other suitable membrane. Alternatively, the support may be a bead or disc, such as glass, fiberglass, latex or a plastic material such as polystyrene or polyvinylchloride. The support may also be a magnetic particle or a fiber optic sensor, such as those disclosed, for example, in U.S. Patent No. 5,359,681. The binding agent may be immobilized on the solid support
10 using a variety of techniques known to those of skill in the art, which are amply described in the patent and scientific literature. In the context of the present invention, the term "immobilization" refers to both noncovalent association, such as adsorption, and covalent attachment (which may be a direct linkage between the agent and functional groups on the support or may be a linkage by way of a cross-linking agent).
15 Immobilization by adsorption to a well in a microtiter plate or to a membrane is preferred. In such cases, adsorption may be achieved by contacting the binding agent, in a suitable buffer, with the solid support for a suitable amount of time. The contact time varies with temperature, but is typically between about 1 hour and about 1 day. In general, contacting a well of a plastic microtiter plate (such as polystyrene or
20 polyvinylchloride) with an amount of binding agent ranging from about 10 ng to about 10 µg, and preferably about 100 ng to about 1 µg, is sufficient to immobilize an adequate amount of binding agent.

Covalent attachment of binding agent to a solid support may generally be achieved by first reacting the support with a bifunctional reagent that will react with
25 both the support and a functional group, such as a hydroxyl or amino group, on the binding agent. For example, the binding agent may be covalently attached to supports having an appropriate polymer coating using benzoquinone or by condensation of an aldehyde group on the support with an amine and an active hydrogen on the binding partner (*see, e.g.,* Pierce Immunotechnology Catalog and Handbook, 1991, at
30 A12-A13).

In certain embodiments, the assay is a two-antibody sandwich assay. This assay may be performed by first contacting an antibody that has been immobilized

on a solid support, commonly the well of a microtiter plate, with the sample, such that polypeptides within the sample are allowed to bind to the immobilized antibody. Unbound sample is then removed from the immobilized polypeptide-antibody complexes and a detection reagent (preferably a second antibody capable of binding to a different site on the polypeptide) containing a reporter group is added. The amount of detection reagent that remains bound to the solid support is then determined using a method appropriate for the specific reporter group.

More specifically, once the antibody is immobilized on the support as described above, the remaining protein binding sites on the support are typically blocked. Any suitable blocking agent known to those of ordinary skill in the art, such as bovine serum albumin or Tween 20™ (Sigma Chemical Co., St. Louis, MO). The immobilized antibody is then incubated with the sample, and polypeptide is allowed to bind to the antibody. The sample may be diluted with a suitable diluent, such as phosphate-buffered saline (PBS) prior to incubation. In general, an appropriate contact time (*i.e.*, incubation time) is a period of time that is sufficient to detect the presence of polypeptide within a sample obtained from an individual with ovarian cancer. Preferably, the contact time is sufficient to achieve a level of binding that is at least about 95% of that achieved at equilibrium between bound and unbound polypeptide. Those of ordinary skill in the art will recognize that the time necessary to achieve equilibrium may be readily determined by assaying the level of binding that occurs over a period of time. At room temperature, an incubation time of about 30 minutes is generally sufficient.

Unbound sample may then be removed by washing the solid support with an appropriate buffer, such as PBS containing 0.1% Tween 20™. The second antibody, which contains a reporter group, may then be added to the solid support. Preferred reporter groups include those groups recited above.

The detection reagent is then incubated with the immobilized antibody-polypeptide complex for an amount of time sufficient to detect the bound polypeptide. An appropriate amount of time may generally be determined by assaying the level of binding that occurs over a period of time. Unbound detection reagent is then removed and bound detection reagent is detected using the reporter group. The method employed for detecting the reporter group depends upon the nature of the reporter

group. For radioactive groups, scintillation counting or autoradiographic methods are generally appropriate. Spectroscopic methods may be used to detect dyes, luminescent groups and fluorescent groups. Biotin may be detected using avidin, coupled to a different reporter group (commonly a radioactive or fluorescent group or an enzyme).

- 5 Enzyme reporter groups may generally be detected by the addition of substrate (generally for a specific period of time), followed by spectroscopic or other analysis of the reaction products.

To determine the presence or absence of a cancer, such as ovarian cancer, the signal detected from the reporter group that remains bound to the solid support is generally compared to a signal that corresponds to a predetermined cut-off value. In one preferred embodiment, the cut-off value for the detection of a cancer is the average mean signal obtained when the immobilized antibody is incubated with samples from patients without the cancer. In general, a sample generating a signal that is three standard deviations above the predetermined cut-off value is considered positive for the cancer. In an alternate preferred embodiment, the cut-off value is determined using a Receiver Operator Curve, according to the method of Sackett *et al.*, *Clinical Epidemiology: A Basic Science for Clinical Medicine*, Little Brown and Co., 1985, p. 106-7. Briefly, in this embodiment, the cut-off value may be determined from a plot of pairs of true positive rates (*i.e.*, sensitivity) and false positive rates (100%- specificity) that correspond to each possible cut-off value for the diagnostic test result. The cut-off value on the plot that is the closest to the upper left-hand corner (*i.e.*, the value that encloses the largest area) is the most accurate cut-off value, and a sample generating a signal that is higher than the cut-off value determined by this method may be considered positive. Alternatively, the cut-off value may be shifted to the left along the plot, to minimize the false positive rate, or to the right, to minimize the false negative rate. In general, a sample generating a signal that is higher than the cut-off value determined by this method is considered positive for a cancer.

In a related embodiment, the assay is performed in a flow-through or strip test format, wherein the binding agent is immobilized on a membrane, such as nitrocellulose. In the flow-through test, polypeptides within the sample bind to the immobilized binding agent as the sample passes through the membrane. A second, labeled binding agent then binds to the binding agent-polypeptide complex as a solution

containing the second binding agent flows through the membrane. The detection of bound second binding agent may then be performed as described above. In the strip test format, one end of the membrane to which binding agent is bound is immersed in a solution containing the sample. The sample migrates along the membrane through a region containing second binding agent and to the area of immobilized binding agent. Concentration of second binding agent at the area of immobilized antibody indicates the presence of a cancer. Typically, the concentration of second binding agent at that site generates a pattern, such as a line, that can be read visually. The absence of such a pattern indicates a negative result. In general, the amount of binding agent immobilized on the membrane is selected to generate a visually discernible pattern when the biological sample contains a level of polypeptide that would be sufficient to generate a positive signal in the two-antibody sandwich assay, in the format discussed above. Preferred binding agents for use in such assays are antibodies and antigen-binding fragments thereof. Preferably, the amount of antibody immobilized on the membrane ranges from about 25 ng to about 1 μ g, and more preferably from about 50 ng to about 500 ng. Such tests can typically be performed with a very small amount of biological sample.

Of course, numerous other assay protocols exist that are suitable for use with the tumor proteins or binding agents of the present invention. The above descriptions are intended to be exemplary only. For example, it will be apparent to those of ordinary skill in the art that the above protocols may be readily modified to use ovarian tumor polypeptides to detect antibodies that bind to such polypeptides in a biological sample. The detection of such ovarian tumor protein specific antibodies may correlate with the presence of a cancer.

A cancer may also, or alternatively, be detected based on the presence of T cells that specifically react with an ovarian tumor protein in a biological sample. Within certain methods, a biological sample comprising CD4⁺ and/or CD8⁺ T cells isolated from a patient is incubated with an ovarian tumor polypeptide, a polynucleotide encoding such a polypeptide and/or an APC that expresses at least an immunogenic portion of such a polypeptide, and the presence or absence of specific activation of the T cells is detected. Suitable biological samples include, but are not limited to, isolated T cells. For example, T cells may be isolated from a patient by routine techniques (such

as by Ficoll/Hypaque density gradient centrifugation of peripheral blood lymphocytes). T cells may be incubated *in vitro* for 2-9 days (typically 4 days) at 37°C with polypeptide (e.g., 5 - 25 µg/ml). It may be desirable to incubate another aliquot of a T cell sample in the absence of ovarian tumor polypeptide to serve as a control. For CD4⁺ T cells, activation is preferably detected by evaluating proliferation of the T cells. For CD8⁺ T cells, activation is preferably detected by evaluating cytolytic activity. A level of proliferation that is at least two fold greater and/or a level of cytolytic activity that is at least 20% greater than in disease-free patients indicates the presence of a cancer in the patient.

As noted above, a cancer may also, or alternatively, be detected based on the level of mRNA encoding an ovarian tumor protein in a biological sample. For example, at least two oligonucleotide primers may be employed in a polymerase chain reaction (PCR) based assay to amplify a portion of an ovarian tumor cDNA derived from a biological sample, wherein at least one of the oligonucleotide primers is specific for (*i.e.*, hybridizes to) a polynucleotide encoding the ovarian tumor protein. The amplified cDNA is then separated and detected using techniques well known in the art, such as gel electrophoresis. Similarly, oligonucleotide probes that specifically hybridize to a polynucleotide encoding an ovarian tumor protein may be used in a hybridization assay to detect the presence of polynucleotide encoding the tumor protein in a biological sample.

To permit hybridization under assay conditions, oligonucleotide primers and probes should comprise an oligonucleotide sequence that has at least about 60%, preferably at least about 75% and more preferably at least about 90%, identity to a portion of a polynucleotide encoding an ovarian tumor protein that is at least 10 nucleotides, and preferably at least 20 nucleotides, in length. Preferably, oligonucleotide primers and/or probes hybridize to a polynucleotide encoding a polypeptide described herein under moderately stringent conditions, as defined above. Oligonucleotide primers and/or probes which may be usefully employed in the diagnostic methods described herein preferably are at least 10-40 nucleotides in length. In a preferred embodiment, the oligonucleotide primers comprise at least 10 contiguous nucleotides, more preferably at least 15 contiguous nucleotides, of a DNA molecule having a sequence recited in SEQ ID NOs:1-222. Techniques for both PCR based

assays and hybridization assays are well known in the art (*see, for example, Mullis et al., Cold Spring Harbor Symp. Quant. Biol., 51:263, 1987; Erlich ed., PCR Technology, Stockton Press, NY, 1989*).

One preferred assay employs RT-PCR, in which PCR is applied in
5 conjunction with reverse transcription. Typically, RNA is extracted from a biological sample, such as biopsy tissue, and is reverse transcribed to produce cDNA molecules. PCR amplification using at least one specific primer generates a cDNA molecule, which may be separated and visualized using, for example, gel electrophoresis. Amplification may be performed on biological samples taken from a test patient and
10 from an individual who is not afflicted with a cancer. The amplification reaction may be performed on several dilutions of cDNA spanning two orders of magnitude. A two-fold or greater increase in expression in several dilutions of the test patient sample as compared to the same dilutions of the non-cancerous sample is typically considered positive.

15 In another embodiment, the compositions described herein may be used as markers for the progression of cancer. In this embodiment, assays as described above for the diagnosis of a cancer may be performed over time, and the change in the level of reactive polypeptide(s) or polynucleotide(s) evaluated. For example, the assays may be performed every 24-72 hours for a period of 6 months to 1 year, and thereafter
20 performed as needed. In general, a cancer is progressing in those patients in whom the level of polypeptide or polynucleotide detected increases over time. In contrast, the cancer is not progressing when the level of reactive polypeptide or polynucleotide either remains constant or decreases with time.

Certain *in vivo* diagnostic assays may be performed directly on a tumor.
25 One such assay involves contacting tumor cells with a binding agent. The bound binding agent may then be detected directly or indirectly via a reporter group. Such binding agents may also be used in histological applications. Alternatively, polynucleotide probes may be used within such applications.

As noted above, to improve sensitivity, multiple ovarian tumor protein
30 markers may be assayed within a given sample. It will be apparent that binding agents specific for different proteins provided herein may be combined within a single assay. Further, multiple primers or probes may be used concurrently. The selection of tumor

protein markers may be based on routine experiments to determine combinations that results in optimal sensitivity. In addition, or alternatively, assays for tumor proteins provided herein may be combined with assays for other known tumor antigens.

DIAGNOSTIC KITS

5 The present invention further provides kits for use within any of the above diagnostic methods. Such kits typically comprise two or more components necessary for performing a diagnostic assay. Components may be compounds, reagents, containers and/or equipment. For example, one container within a kit may contain a monoclonal antibody or fragment thereof that specifically binds to an ovarian
10 tumor protein. Such antibodies or fragments may be provided attached to a support material, as described above. One or more additional containers may enclose elements, such as reagents or buffers, to be used in the assay. Such kits may also, or alternatively, contain a detection reagent as described above that contains a reporter group suitable for direct or indirect detection of antibody binding.

15 Alternatively, a kit may be designed to detect the level of mRNA encoding an ovarian tumor protein in a biological sample. Such kits generally comprise at least one oligonucleotide probe or primer, as described above, that hybridizes to a polynucleotide encoding an ovarian tumor protein. Such an oligonucleotide may be used, for example, within a PCR or hybridization assay. Additional components that
20 may be present within such kits include a second oligonucleotide and/or a diagnostic reagent or container to facilitate the detection of a polynucleotide encoding an ovarian tumor protein.

The following Examples are offered by way of illustration and not by way of limitation.

EXAMPLE 1

IDENTIFICATION OF CDNAS ENCODING OVARIAN AND ENDOMETRIAL TUMOR PROTEINS

An ovarian/endometrial tumor cell line subtracted library was
 5 constructed. A library was prepared from endometrial and ovarian tumor cell lines:
 EndoTL 391-73 (100% undifferentiated endometrial carcinoma), OTL 298-95 (100%
 moderately differentiated papillary serous ovarian adenocarcinoma) and OTL 522-24
 (30% mesothelial cells/70% poorly differentiated metastatic ovarian adenocarcinoma).
 This library was subtracted with liver, pancreas, skin, bone marrow, resting PBMC,
 10 stomach, and brain cDNA and spiked with eukaryotic elongation factor 1 α . Resulting
 cDNA was cloned into the pcDNA3.1(+) (Invitrogen) vector to generate the ovarian
 tumor cell line subtraction 4 library (OTCLS4). The OTCLS4 library contained
 117,200 clones (background 58,400), with a 1333 bp average insert size (inserts ranged
 from 200 to 5650 bp).

15 Thirty clones were sequenced. Of these 12 were full length. The clones
 may be grouped as follows (SEQ ID NOs are provided in Table 2):

- 7 Novel
- 4 Homo sapiens aldehyde dehydrogenase 6 (ALDH6) mRNA
- 3 Human ferritin heavy chain mRNA, complete cds
- 20 2 Human lysyl oxidase gene, partial cds
- 2 Human mitochondrion, complete genome
- 1 Homo sapiens aldehyde reductase 1 (low Km aldose reductase) ALDR1)
mRNA
- 1 Homo sapiens chromosome 11q12.2 PAC clone pDJ519o13
- 25 1 Homo sapiens chromosome-associated polypeptide C (CAP-C) mRNA
- 1 Homo sapiens clone 24452 mRNA sequence
- 1 Homo sapiens dipeptidylpeptidase IV (CD26, adenosine deaminase
complexing protein 2) (DPP4 mRNA)
- 1 Homo sapiens guanine nucleotide binding protein (G protein), beta
polypeptide 2-like 1 (GNB2L1), mRNA
- 30 1 Homo sapiens heat shock 27kD protein 1 (HSPB1) mRNA
- 1 Homo sapiens homeo box B2 (HOXB2) mRNA

- 1 Homo sapiens mRNA for KIAA0865 protein, partial cds
- 1 Homo sapiens mRNA; cDNA DKFZp564A2416 (from clone DKFZp564A2416)
- 1 Homo sapiens NADH-ubiquinone oxidoreductase 39kDA subunit mRNA,
5 nuclear gene encoding mitochondrial protein, complete cds
- 1 Homo sapiens Sk/Dkk-1 protein precursor, mRNA, complete cds
- 1 Homo sapiens sodium channel, nonvoltage-gated 1 alpha (SCNN1A) mRNA
- 1 Homo sapiens SRP1 mRNA, partial sequence
- 1 Homo sapiens zinc finger protein SLUG (SLUG) gene, complete cds
- 10 1 Human 28S ribosomal RNA gene
- 1 Human cofilin mRNA, partial cds
- 1 Human DNA sequence from clone 967N21 on chromosome 20p12.3-13
- 1 Human fibroblast collagenase inhibitor mRNA, complete cds
- 1 Human fibroblast mRNA for aldolase A
- 15 1 Human HepG2 3' region MboI cDNA, clone hmd6a06m3
- 1 Human MAP kinase kinase MEK5c mRNA, complete cds
- 1 Human mRNA for coupling protein G(s) alpha-subunit (alpha-S1)
- 1 Human mRNA for KIAA0026 gene, completecds|gi|4808630|gb|AF100620.1|
AF100620 Homo sapiens transcription factor-like protein MRGX (MRGX)
20 mRNA, complete cds
- 1 Human mRNA for KIAA0064 gene, complete cds
- 1 Human mRNA for KIAA0204 gene, complete cds
- 1 Human plasminogen activator inhibitor-1 (PAI-1) mRNA, complete cds
- 1 Human protocadherin 43 mRNA, 3' end of cds for alternative splicing PC43-
25 12
- 1 Human putative RNA binding protein Koc1 mRNA, complete cds
- 1 Human TCB gene encoding cytosolic thyroid hormone-binding protein,
complete cds
- 1 Human ubiquitin-homology domain protein PIC1 mRNA, complete cds
- 30

Table 2
Ovarian/Endometrial Carcinoma Associated cDNA Sequences

Sequence	SEQ ID NO	Comments
32609	36	<i>Homo sapiens</i> aldehyde dehydrogenase 6 (ALDH6) mRNA
32515	4	<i>Homo sapiens</i> aldehyde reductase 1 (low Km aldose reductase) (ALDR1) mRNA
32562	29	<i>Homo sapiens</i> Chromosome 11q12.2 PAC clone pDJ519o13
32523	9	<i>Homo sapiens</i> chromosome-associated polypeptide C (CAP-C) mRNA
32551	24	<i>Homo sapiens</i> clone 24452 mRNA sequence
32518	6	<i>Homo sapiens</i> dipeptidylpeptidase IV (CD26, adenosine deaminase complexing protein 2) (DPP4) mRNA
32534	13	<i>Homo sapiens</i> guanine nucleotide binding protein (G protein), beta polypeptide 2-like 1 (GNB2L1), mRNA
32507	2	<i>Homo sapiens</i> heat shock 27kD protein 1 (HSPB1) mRNA
32533	12	<i>Homo sapiens</i> homeo box B2 (HOXB2) mRNA
32565	20	<i>Homo sapiens</i> mRNA for KIAA0865 protein, partial cds
32553	19	<i>Homo sapiens</i> mRNA; cDNA DKFZp564A2416 (from clone DKFZp564A2416)
32561	28	<i>Homo sapiens</i> NADH-ubiquinone oxidoreductase 39kDa subunit mRNA, nuclear gene encoding mitochondrial protein, complete cds
32510	3	<i>Homo sapiens</i> Sk/Dkk-1 protein precursor, mRNA, complete cds
32546	16	<i>Homo sapiens</i> sodium channel, nonvoltage-gated 1 alpha (SCNN1A) mRNA
32559	27	<i>Homo sapiens</i> SRP1 mRNA, partial sequence
32506	1	<i>Homo sapiens</i> zinc finger protein SLUG gene, complete cds
32519	7	Human 28S ribosomal RNA gene
32602	22	Human cofilin mRNA, partial cds
32569	31	Human DNA sequence from clone 967N21 on chromosome 20p12.3-13
32525	10	Human ferritin heavy chain mRNA, complete cds
32557	26	Human fibroblast collagenase inhibitor mRNA, complete cds
32517	5	Human fibroblast mRNA for aldolase A
32568	30	Human HepG2 3' region MboI cDNA, clone hmd6a06m3
32548	17	Human lysyl oxidase gene, partial cds
32520	8	Human mitochondrion, complete genome
32617	23	Human mRNA for coupling protein G(s) alpha-subunit (alpha-S1)

32572	32	Human mRNA for KIAA0026 gene, complete cds gi 4808630 gb AF100620.1 AF100620 <i>Homo sapiens</i> transcription factor-like protein MRGX (MRGX) mRNA, complete cds
32600	21	Human mRNA for KIAA0064 gene, complete cds
32537	14	Human mRNA for KIAA0204 gene, complete cds
32552	25	Human plasminogen activator inhibitor-1 (PAI-1) mRNA, complete cds
32615	39	Human protocadherin 43 mRNA, 3' end of cds for alternative splicing PC43-12
32613	38	Human putative RNA binding protein Koc1 mRNA, complete cds
32610	37	Human TCB gene encoding cytosolic thyroid hormone-binding protein, complete cds
32539	15	Human ubiquitin-homology domain protein PIC1 mRNA, complete cds
32619	40	Novel
32576	33	Novel
32608	35	Novel
32607	34	Novel
32620	41	Novel
32550	18	Novel
32529	11	Novel

Using the methods outlined above, an additional 162 clones were isolated and sequenced. The cDNA sequences are shown in SEQ ID NO:42-203.

SEQ ID NO:204-209 represent additional clones from the OTCL S4 library. SEQ ID NO:206 (clone 57881), 208 (clone 57884), 107 (clone R0199:A07) and 80 (clone R0198:F02) represent novel sequences. The remaining sequences are shown in Table 3, which includes additional results from homology searches.

Table 3

Sequence	SEQ ID NO	Comments
57877	204	H. Sapiens novel gene from PAC 117P20, chromosome 1
57879	205	Urokinase plasminogen activator surface receptor (uPAR)
57882	207	Lysophospholipase 1 (LYPA1)
57888	209	IGF-II mRNA binding protein 3 (IMP-3) mRNA
R0198:H03	99	<i>Homo sapiens</i> laminin
R0199:B03	111	Human cyclin protein gene, complete cds
R0200:A12	158	<i>Homo sapiens</i> monocarboxylate transporter (MCT3) mRNA
R0199:C12	125	Unigene: Hs93379
R0200:A10	157	Human mRNA for KIAA0101 gene, complete cds
R0198:D01	61	Unigene: Hs42116
R0200:C02	164	Human proliferating cell nuclear antigen (PCNA) gene
R0200:G02	193	<i>Homo sapiens</i> Xq28 BAC RP5-1014016

EXAMPLE 2

5 ANALYSIS OF CDNA EXPRESSION USING MICROARRAY TECHNOLOGY

In additional studies, sequences disclosed herein were found to be overexpressed in specific tumor tissues as determined by microarray analysis. Using this approach, cDNA sequences are PCR amplified and their mRNA expression profiles in tumor and normal tissues are examined using cDNA microarray technology essentially as described (Shena *et al.*, 1995). In brief, the clones are arrayed onto glass slides as multiple replicas, with each location corresponding to a unique cDNA clone (as many as 5500 clones can be arrayed on a single slide, or chip). Each chip is hybridized with a pair of cDNA probes that are fluorescence-labeled with Cy3 and Cy5, respectively. Typically, 1 μ g of polyA⁺ RNA is used to generate each cDNA probe.

15 After hybridization, the chips are scanned and the fluorescence intensity recorded for both Cy3 and Cy5 channels. There are multiple built-in quality control steps. First, the probe quality is monitored using a panel of ubiquitously expressed genes. Secondly, the control plate also can include yeast DNA fragments of which complementary RNA may be spiked into the probe synthesis for measuring the quality of the probe and the

20 sensitivity of the analysis. Currently, the technology offers a sensitivity of 1 in 100,000

copies of mRNA. Finally, the reproducibility of this technology can be ensured by including duplicated control cDNA elements at different locations.

A total of 428 clones from the OCTLS4 library were analyzed on Ovarian Chip-3. The following table, Table 4, provides a list of probes used to
5 interrogate these clones. A total of 16 clones were identified which showed at least 2-fold overexpression in ovarian tumors when compared to non-ovarian essential normal tissues and had a mean non-ovarian essential normal tissue expression of less than 0.2. These clones are represented by SEQ ID NO:204-209 and by SEQ ID NO:61, 99, 111, 125, 157, 158, 164 and 193.

10

Table 4

Tissue	Clone ID	Microarray ID	Tumor information
Ovarian tumor Adrenal gland normal	261A SPACT37	391cy3 391cy5	Stage IIIC
Ovary tumor Skin normal	264A 396A	392cy3 392cy5	Stage IIIC
Ovary tumor Thymus normal	265A SPACT56	393cy3 393cy5	Stage IIIC
Ovary tumor Bronchus normal	288A 600C	394cy3 394cy5	Stage IIIC
Ovary tumor	854A 785B	395cy3 395cy5	
Ovary tumor Bone normal	855A 407B	396cy3 396cy5	Grade III, Stage IA
Ovary tumor Peritoneum epithelium normal	856A 484A	397cy3 397cy5	Serous papillary
Ovary tumor Pituitary gland	603A SPACT52	398cy3 398cy5	Metastatic adenocarcinoma, Grade III, Stage III
Ovary tumor Skeletal muscle normal	857A SPACT40	399cy3 399cy5	Papillary serous cystadenocarcinoma Grade III, Stage IB
Ovary tumor Stomach normal	385A SPACT55	400cy3 400cy5	Papillary serous adenocarcinoma
Ovary tumor Spleen normal	392A SPACT54	401cy3 401cy5	Papillary serous neoplasm, Stage IC
Ovary tumor Pancreas normal	858A 862A	402cy3 402cy5	Papillary serous cystadenocarcinoma Grade II-III, Stage IA

Ovary tumor Ovary normal	859A S27	403cy3 403cy5	Papillary serous adenocarcinoma Grade II-III, Stage IIB
Ovary tumor Spinal cord normal	605A SPACT45	404cy3 404cy5	Serous borderline tumor, stage IIIC
Ovary tumor Heart normal	495A SPAAM1	405cy3 405cy5	Papillary serous carcinoma, Grade II, Stage III
Ovary tumor Ovary normal	381C S7	414cy3 414cy5	Mucinous adenocarcinoma, Grade I, Stage IB
Ovary tumor Ovary normal	382A S449A	416cy3 416cy5	Mucinous adenocarcinoma
Ovary tumor metastases Small intestine normal	428B SPACT53	417cy3 417cy5	Mucinous adenocarcinoma
Ovary tumor Esophagus normal	491A 502B	418cy3 418cy5	Endometriod adenocarcinoma
Ovary tumor Colon normal	335A 199A	419cy3 419cy5	Endometriod adenocarcinoma Grade II, Stage II
Ovary tumor Thyroid gland normal	494A SPACT46	421cy3 421cy5	Adenocarcinoma Grade III, Stage II- III
Ovary tumor PBM (resting)	860A 783A	42cy3 422cy5	Endometriod adenocarcinoma Grade II-III, Stage IIIC
Ovary tumor Aorta normal	604A 415A	423cy3 423cy5	Clear cell carcinoma
Ovary tumor Trachea normal	607A 776A	424cy3 424cy5	Clear cell, Stage IA
Ovary tumor Trachea normal	S25 CT25	425cy3 425cy5	Granulosa cell tumor, Stage IA
Ovary tumor Pancreas normal pool	S22 PAN2000	426cy3 426cy5	Granulosa cell tumor, Stage IA
Ovary tumor Breast (HMEC) normal	386A S92	427cy3 427cy5	Germ cell tumor, Stage I
Ovary tumor Bladder normal	602A 328B/C	429cy3 429cy5	Papillary serous carcinoma, Grade III, Stage IIB
Ovary tumor Bone marrow normal	S23 SPACT49	430cy3 430cy5	Papillary serous adenocarcinoma Grade III, Stage IIIC

Ovary tumor Lung normal	606A SPAAm2	428cy3 428cy5	Papillary serous cystadenocarcinoma Grade II, Stage IIIB
Ovary tumor metastases Kidney normal	383A 302B	431cy3 431cy5	Metastatic papillary adenocarcinoma, Grade III, Stage IIIA
Ovary tumor metastases PBMC (activated)	384A S40.782A	423cy3 423cy5	Papillary serous adenocarcinoma Grade II, Stage IIIB
Ovary tumor metastases Ovary tumor match with CY3	426A 603A	433cy3 433cy5	Papillary serous adenocarcinoma Grade III, Stage IIIB
Ovary tumor metastases Liver normal	429A 270B	434cy3 434cy5	Papillary adenocarcinoma Grade III, Stage III
Ovary tumor Brain normal	427A SPACT50	435cy3 435cy5	Papillary serous adenocarcinoma Grade III, Stage IIIC
Ovary tumor Bone normal	855A 407B	436cy3 436cy5	Grade III, Stage IA
Ovary tumor Spinal cord normal	605A SPACT45	437cy3 437cy5	Serous borderline tumor, Stage IIIC
Ovary tumor Heart normal	495A SPAAm1	438cy3 438cy5	Papillary serous carcinoma, Grade II, Stage III
Ovary tumor Ovary normal	381C S7	439cy3 439cy5	Mucinous adenocarcinoma, Grade I, Stage IB

EXAMPLE 3

SYNTHESIS OF POLYPEPTIDES

5 Polypeptides may be synthesized on a Perkin Elmer/Applied Biosystems Division 430A peptide synthesizer using Fmoc chemistry with HPTU. (O-Benzotriazole-N,N,N',N'-tetramethyluronium hexafluorophosphate) activation. A Gly-Cys-Gly sequence may be attached to the amino terminus of the peptide to provide a method of conjugation, binding to an immobilized surface, or labeling of the peptide.

10 Cleavage of the peptides from the solid support may be carried out using the following cleavage mixture: trifluoroacetic acid:ethanedithiol:thioanisole:water:phenol (40:1:2:2:3). After cleaving for 2 hours, the peptides may be precipitated in cold methyl-t-butyl-ether. The peptide pellets may then be dissolved in water containing 0.1% trifluoroacetic acid (TFA) and lyophilized prior to purification by C18 reverse

15 phase HPLC. A gradient of 0%-60% acetonitrile (containing 0.1% TFA) in water (containing 0.1% TFA) may be used to elute the peptides. Following lyophilization of the pure fractions, the peptides may be characterized using electrospray or other types of mass spectrometry and by amino acid analysis.

From the foregoing it will be appreciated that, although specific

20 embodiments of the invention have been described herein for purposes of illustration, various modifications may be made without deviating from the spirit and scope of the invention. Accordingly, the invention is not limited except as by the appended claims.

CLAIMS

What is claimed:

1. An isolated polynucleotide comprising a sequence selected from the group consisting of:
 - (a) sequences provided in SEQ ID NO: 1-222;
 - (b) complements of the sequences provided in SEQ ID NO: 1-222;
 - (c) sequences consisting of at least 20 contiguous residues of a sequence provided in SEQ ID NO: 1-222;
 - (d) sequences that hybridize to a sequence provided in SEQ ID NO: 1-222, under moderately stringent conditions;
 - (e) sequences having at least 75% identity to a sequence of SEQ ID NO: 1-222;
 - (f) sequences having at least 90% identity to a sequence of SEQ ID NO: 1-222; and
 - (g) degenerate variants of a sequence provided in SEQ ID NO: 1-222.
2. An isolated polypeptide comprising an amino acid sequence selected from the group consisting of:
 - (a) sequences encoded by a polynucleotide of claim 1; and
 - (b) sequences having at least 70% identity to a sequence encoded by a polynucleotide of claim 1; and
 - (c) sequences having at least 90% identity to a sequence encoded by a polynucleotide of claim 1.
3. An expression vector comprising a polynucleotide of claim 1 operably linked to an expression control sequence.
4. A host cell transformed or transfected with an expression vector according to claim 3.

5. An isolated antibody, or antigen-binding fragment thereof, that specifically binds to a polypeptide of claim 2.

6. A method for detecting the presence of a cancer in a patient, comprising the steps of:

- (a) obtaining a biological sample from the patient;
- (b) contacting the biological sample with a binding agent that binds to a polypeptide of claim 2;
- (c) detecting in the sample an amount of polypeptide that binds to the binding agent; and
- (d) comparing the amount of polypeptide to a predetermined cut-off value and therefrom determining the presence of a cancer in the patient.

7. A fusion protein comprising at least one polypeptide according to claim 2.

8. An oligonucleotide that hybridizes to a sequence recited in SEQ ID NO: 1-222 under moderately stringent conditions.

9. A method for stimulating and/or expanding T cells specific for a tumor protein, comprising contacting T cells with at least one component selected from the group consisting of:

- (a) polypeptides according to claim 2;
- (b) polynucleotides according to claim 1; and
- (c) antigen-presenting cells that express a polypeptide according to claim 2,

under conditions and for a time sufficient to permit the stimulation and/or expansion of T cells.

10. An isolated T cell population, comprising T cells prepared according to the method of claim 9.

11. A composition comprising a first component selected from the group consisting of physiologically acceptable carriers and immunostimulants, and a second component selected from the group consisting of:

- (a) polypeptides according to claim 2;
- (b) polynucleotides according to claim 1;
- (c) antibodies according to claim 5;
- (d) fusion proteins according to claim 7;
- (e) T cell populations according to claim 10; and
- (f) antigen presenting cells that express a polypeptide according to claim 2.

12. A method for stimulating an immune response in a patient, comprising administering to the patient a composition of claim 11.

13. A method for the treatment of a cancer in a patient, comprising administering to the patient a composition of claim 11.

14. A method for determining the presence of a cancer in a patient, comprising the steps of:

- (a) obtaining a biological sample from the patient;
- (b) contacting the biological sample with an oligonucleotide according to claim 8;
- (c) detecting in the sample an amount of a polynucleotide that hybridizes to the oligonucleotide; and
- (d) compare the amount of polynucleotide that hybridizes to the oligonucleotide to a predetermined cut-off value, and therefrom determining the presence of the cancer in the patient.

15. A diagnostic kit comprising at least one oligonucleotide according to claim 8.

16. A diagnostic kit comprising at least one antibody according to claim 5 and a detection reagent, wherein the detection reagent comprises a reporter group.

17. A method for inhibiting the development of a cancer in a patient, comprising the steps of:

(a) incubating CD4+ and/or CD8+ T cells isolated from a patient with at least one component selected from the group consisting of: (i) polypeptides according to claim 2; (ii) polynucleotides according to claim 1; and (iii) antigen presenting cells that express a polypeptide of claim 2, such that T cell proliferate;

(b) administering to the patient an effective amount of the proliferated T cells,

and thereby inhibiting the development of a cancer in the patient.

SEQUENCE LISTING

<110> Corixa Corporation
 Xu, Jiangchun
 Pyle, Ruth
 Secrist, Heather

<120> COMPOSITIONS AND METHODS FOR THE THERAPY
 AND DIAGNOSIS OF OVARIAN AND ENDOMETRIAL CANCER

<130> 210121.501PC

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 <222> (1)...(375)
 <223> n = A,T,C or G

tttttttttt	tttttttttt	tttttttgna	ttataaanac	atttatttaa	tctatgaaaa	60
taatgnacaa	taaatacttt	ccccttttcc	tattattaaa	naattttaat	aaataatnta	120
cagtctaaaa	cataaaaaag	aggaaaatag	gncctcttag	ttatttttaa	naaagncccc	180
ctanagttta	attattcctg	anatttcatt	ggaaggagtc	taccaaacgg	aatttttctg	240
ngngaatttt	aaaanataac	cgagtgccca	atattttaga	agaagaagaa	aggaagngga	300
ttaaacgcta	attcagtaat	acctgaattt	tagcaaaaca	cataagtcta	tgcgactgag	360
gnggggagan	gntcg					375

<210> 13
 <211> 658
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(658)
 <223> n = A,T,C or G

ctctctcttt	cactgcaagg	cggcggcagg	agaggttgtg	gtgctagttt	ctctaagcca	60
tccagtgcc	tctctgtcgc	tgcagcgaca	cacgctctcg	ccgccgccat	gactgagcag	120
atgacccttc	gtggcaccct	caagggccac	aacggctggg	taaccagat	cgctactacc	180
ccgcagttcc	cggacatgat	cctctccgcc	tctcgagata	agaccatcat	catgtggaaa	240
ctgaccaggg	atgagaccaa	ctatggaatt	ccacagcgtg	ctctgcgggg	tactcccac	300
tttgtagtg	atgtggttat	ctctcagat	ggccagtttg	ccctctcang	ctcctgggat	360
ggaaccctgc	gcctctggga	tctcacaacg	ggcaccacca	cgaggcgatt	tgtgggccat	420
accaaggatg	tgcttgagtg	tggccttctc	tttgacaacc	cggcagattg	nccttttgat	480
ctcnanaata	aaaccatcaa	ncatttgat	accctggng	tggtgcaaat	ccctgttcca	540
ngaaganaac	cncttcanaa	ngggggtcct	tgtgnncnt	ttttnccca	acncaacaac	600
cctnttattn	nntncctngg	gttggaasaan	ctggcnnggn	tnganccggn	tnactggg	658

<210> 14
 <211> 686
 <212> DNA
 <213> Homo sapien

<220>

<221> misc_feature
 <222> (1)...(686)
 <223> n = A,T,C or G

<400> 14

cctttttttt	ttttttttt	ttttttttt	aacattatac	tgncattttt	atcataacaa	60
tataaacaat	ttttatcatc	atcctgaata	ttactttata	aanatatata	ttttaaaagg	120
ntttcaaaac	atttttcaac	ccagcatttg	agaataaagc	attaagagtt	ttgnatacag	180
taacacattc	atgngataag	ngnatgaatt	tacaaccata	cataatatgg	atatatggat	240
atatatttat	ataaaaaaca	aacttggcca	naagttaagg	ntacctacna	agttgtccaa	300
gtaaattatg	cttggcaaaa	caattataaa	attcaaatac	cacatgcatt	tttaaatacat	360
ctaaatcact	gcaaaccaang	gtcaagcatt	ccaaangttt	taaaatnang	gggggangang	420
ggaancnggc	cctccaannt	taaagggcc	gtttaaaacc	cccttgacc	ccccccaca	480
ggngntttt	aactnccncc	catttntgtt	gtttgnnt	ttcnccgggg	ccttcttttg	540
cccttggang	gggccncccc	cccctgggcc	ttccnaaata	aaagggagga	aaanngnntt	600
cccacgnccc	ccccgnatg	natnctctcc	tntaaaaaaa	ngggngggnc	gngannctaa	660
nnggagnggt	ttggcnaanc	acttct				686

<210> 15
 <211> 725
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(725)
 <223> n = A,T,C or G

<400> 15

cctttttttt	tttttttgat	ttttacaaat	attgnttatt	ttaatgaagc	tggtacagac	60
aatgtccatt	taaaacccat	atcccaggcc	aaaaagtaca	aataaaatca	aaaagagcag	120
tggtctgntg	tattcatctt	tgatgtata	gctttattaa	ttngctaata	aaaattanaa	180
cttttctggg	atcttctgac	aagattttta	aaaaatctta	aaatgccttt	tcttcagtga	240
aggcactttt	ggagttncca	ataaaggggn	ccccccctnc	catcttnact	tnaacctgat	300
attntntttg	tgnggggggg	ggngggngaa	attttaaaaa	tatnttaatt	taaggaaagg	360
ncattttttc	acagtctaag	ttctntgnaa	aacttncatt	ttcccacnga	aagnganagt	420
tnangaannc	ccccnngggc	ncnccccacc	ntgnggggca	anttgnaaan	tnattatnga	480
acncttggtg	ttgnttgaat	tntttntgnt	aacgnnnaat	tgcgtgnaag	aangctatcg	540
ttntctgtaa	aaaaagggga	aacttttntc	atantntccn	ntannttctt	tttanaaacc	600
ccnacccccc	ctaaatgtga	nccnccgatn	ttttncgggg	gntggatntt	nntcngccct	660
tcnncncccg	cccttttttt	anacgccnat	ttatattttt	taantttatn	taantttctca	720
tntct						725

<210> 16
 <211> 196
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(196)
 <223> n = A,T,C or G

<400> 16

cngaaggtn	cctncaccct	ggcatcctcc	cctccttccn	acttntgccc	ccaccccatg	60
tctctgtcct	tgtcccagcc	aggccctgct	ccctctccag	ccttgacagc	ccctccccct	120
gcctatgcca	ccctggggcc	ccgcccctct	ccagggggct	ctgcaggggc	cagttcctcc	180
gcctgtcctc	tggggg					196

nnnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	1380
nnnnncggnnn	cnnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	1440
nnnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	nnc	1493

<210> 19
 <211> 1602
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(1602)
 <223> n = A,T,C or G

<400> 19						
ggaaaaatcaa	gatgtggctg	aagatcagag	gctcagttag	caacctgtgt	tgtagcagtg	60
atgtcagtc	attgattgtc	tttagagagt	taatgttaca	aaaaagaatt	cttaataatc	120
agacaaacat	gatctgctga	ggacacatgc	gctttttag	aatttaacat	ctgggtgttt	180
tctgaaaaaa	tatatataca	tatattgctt	tatttgaaac	aaattaaaat	atgctgcatt	240
tgaaaaaaa	aaaaaaaaaa	aaaaaaaaaa	aaaaaaaaaa	aaaaaaaaaa	aaaaaaaaaa	300
aaaaaaaaa	aaaaaaaaa	anggggggn	ccccccng	gnngnnttt	ttgnaaantc	360
cccccccn	gapntnggn	ncccnacnc	ggcccannt	ttantttaan	cccnccccc	420
cttggggccc	ccctnnngg	ggggnnttna	tttccaaan	cccccaanng	ngggggtnt	480
tnntttcncc	aaaaancnt	tttttttnaa	accnccccc	ggaaccccn	ccccccctt	540
ttcntttaag	ggggggngg	gnntttntcc	ccccttttg	gaaaancccc	ctttttttt	600
tggggggccc	aaaaaaaaacc	ccccctttng	naccnnnnan	ggggggggg	ggggnaancc	660
tttgggaaaa	cccccccnng	gggagngaaa	anccctttt	ttccccccc	ccctttttgt	720
tttcctnngc	cccaaaaacc	ccntcccccn	ntgggggann	tnggcngng	annnnannan	780
cccnnaaaan	gnccccccc	cccnnnngn	gaaaaancc	cccnnaang	ggnttntntc	840
cnnggggana	aaaanccng	gggggggncn	ttttccccc	tttngnccc	naaangggg	900
gggccccct	tgggcnnna	aaaacccct	ttntntcccn	cccccgngg	ggggnnnttt	960
ccccccnaaa	ntccccccc	ctngccccna	angggaaac	ccccnnngn	gggtcccttn	1020
gggnccccc	cnntttttt	cccccnngg	gcgggggng	nggggggga	nccccgng	1080
gggcctttcc	nnnngtttt	cncccnnc	cctntnnng	gggtgaaan	aaccccccn	1140
ngnntntnt	anccccccna	nannngncc	ccnnttttg	tnccccccn	cngaanncn	1200
acccccccc	ccnannttt	tttgnnnng	gncnccccn	gngntntnt	ncccccccc	1260
cccccccc	ccgggggng	ggntttttt	gnnnnnnnn	nccccngg	ggggngcccc	1320
ncccccncc	ggnttttgg	ngnnncccc	ctnntttnt	tnnnccncc	cccccccc	1380
cgcnttttn	gngngngng	nnnnnngcn	ccccctnn	gntcnntnt	cncccccn	1440
nnnnnnnnn	nnnnnnnnn	nnnnnnnnn	nnnnnnnnn	nnnnnnnnn	nnnnnnnnn	1500
ncnncngcn	tcnnnnnnn	nnnnnnnnn	nnnnnnnnn	nnngnnngng	nnnnnnncn	1560
nnnnncnnn	nnnngcnnn	ngnnngcnc	cgcccnenn	cc		1602

<210> 20
 <211> 1633
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(1633)
 <223> n = A,T,C or G

<400> 20						
agcagccag	ccatcagccc	ctgaatccac	ctcaccact	cgccagacct	ttttgtcgaa	60
gttcagtgc	ttccttagcc	ttccaatgaa	gcctctacct	gcctgagatg	tccaaggtaa	120
tccatcagct	gaggctctca	gagaatgaaa	gtgtggccct	gcaggaaact	ttggactgga	180
ggagaaagct	ctgtgaggaa	ggacaagact	ggcagcagat	cctgcaccac	gctgagccca	240

gggtgcctcc	cccaccacct	tgcaagaagc	ccagccttct	gaagaagccg	gaaggggcct	300
cctgcaacag	gctgccgtct	gagctctggg	acaccaccat	ttgatgtggc	ctgaactgca	360
gacttacaaa	atagaactgc	ctactgattc	cgggctgcaa	caacagaagg	ctgccttctg	420
acatgcgctg	gggcttctct	ccacgcattt	agacaaaaaa	agcacaggac	acagacacta	480
aatatatgag	atcccgtgtg	tgtgtgtgtg	tgtttgtgtg	tgtgtgtgtg	ggttctttct	540
tatccatctc	gnngngatac	actctgattt	tcaagctcct	catttacggg	tcttgtgcta	600
cccctaggta	ncaagaaaaa	aggctgggaa	aaagtgtggg	cgtggncnan	agcgananaa	660
gtancggngg	gaaaggagcn	antccatgca	cacttctgta	ccngtngttt	ttntnacngg	720
ntcaaacagg	nntgnntnat	tggncnttnc	caangggggg	ttnttttant	aannaccnng	780
nnntrncngg	ggannaanan	nannnnnnna	nnnnnnnttt	nggnnnnccn	cccttggggg	840
ggnnnnantt	ggggcncnct	cnctccccc	cctcncnccc	ccctccccc	tcacnncgnc	900
ncncntnnn	cnccggcgcn	netccnctc	nnccnccnnn	ntcgncccn	nnnggggggg	960
gcggggnngn	ncnccnctct	netccnccnn	cccccccn	cncnccn	ncnccncccc	1020
cncccnccc	nnnnccccc	cnccncccc	ccccccnn	nnnnnnnn	nnnnnnnnnn	1080
nnnnccccc	cccccnccc	cccccnccn	cnnnnnnn	nnnnnnnn	nnnnnnnnnn	1140
nnnnnnnn	nnnnnnnn	nnnnnnnn	nnnggggcn	ngnnnnnn	nnnnnnnn	1200
nnnnnnnn	nnccccccc	cnnnnnnn	nnnnnnnn	nncccccn	nnngnnccn	1260
nnnnnnngnn	ngnggggggn	gnnnnnnn	nnnnnnnn	nnngnnnn	nnnnnnnn	1320
nnnnnnnnng	ggnnnnnnnn	nnnnnnnn	nnnnnnnnng	nnnnnnngnn	nnnnnnnnnn	1380
ngnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	nnngnnnnnn	nnnnnnnnnn	nnnnnnngnn	1440
nnnnnnnnnn	nccgcccc	cgnnccnnnn	nnnnngnnnn	nnnnnnnnnn	nnnnnnnnnn	1500
nnnnnnnnnn	nnnnnnnnng	gggnngcg	ngnnggggn	nnnnngnnnn	nnnnnnnnnc	1560
cncccccn	nnnnnnnnnn	nnnnnnnnnn	nnnnngnnnn	nnngnnnnng	nnnnnnccn	1620
nnccccccng	nnn					1633

<210> 21

<211> 1462

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(1462)

<223> n = A,T,C or G

<400> 21

gggctcccaa	aatggcgaag	tgaggctg	gggactcgct	gagcagcgga	gggggagcgt	60
gcagagccgc	tgccgccctc	acagtcggga	gcccggccgt	gccgtgccgt	aggggaacatg	120
cacttttcca	ttcccgaac	cgagtcgccg	agcggggaca	gcgccggctc	cgcctacgtg	180
gcctataaca	ttcacgtgaa	tggagtcctg	cactgtcggg	tgcgtacag	ccagctcctg	240
gggctgcacg	agcagcttcg	gaaggagtat	ggggccaatg	tgcttcctgc	attcccccca	300
aagaagcttt	tctctctgac	tctgtctgag	gtagaacaga	ggagagagca	gttagagaag	360
tacatgcaag	ctgttcggca	agacccattg	cttgggagca	gcgagacttt	caacagtttc	420
ctgcgtcggg	cacaacagga	gacacagcag	gtccccacag	aggaagtgtc	cttgggaagtg	480
ctgctcagca	acgggcagaa	agttctggtc	aacgtgctaa	cttcagatca	gactgaggat	540
gtcctggagg	ctgtagctgc	aaagctggat	cttcagatg	acttgattgg	atactttagt	600
ctattcttag	ttcgagaaaa	agaggatgga	gccttttctt	ttgtacngaa	gttgcaanaa	660
tttganctgc	cttatgtgtc	tgtcaccagc	cttcgagtca	anantataan	atgtgctaag	720
gaaganttat	tgggactctc	ctatgatnac	nattnatgga	naacccggtt	ggccttnaac	780
cttctttttg	ctcanacggt	nttaaaatat	ttagnccngg	ggngggatct	ttggtcaccc	840
aaggaaaaan	nacccggnaa	ntttaaaatt	ttttgnnaaa	aaaaaaannn	ttccnaaaaa	900
gggaatttct	ttnaaaantg	gccccaaana	ccttngngnn	cttngngnnn	ntttgnnctt	960
ttanncccna	nggggggngg	ntttncnna	aaaaaaattt	nttttnnngg	gnnnnnccnn	1020
nnccnnnnna	annnnnnnn	nnnnnccn	cnngnnnnnn	nnntnnaaag	nnntttnnng	1080
gnccccnaaa	aatngggggn	ncnntttttt	nttttncn	nnnnnnnnnn	nnnnnnnggg	1140
ggggggggnc	ccnnnnnttt	ttnnnnnnnn	nnnnnnnnnn	nnnnccnncc	ccnnntnnaa	1200
annnnnnnnn	nnnnnnnnnn	aannnnnnnn	nnnnnnnnnn	nnngggggggn	nnnnnnnnnn	1260
nnnnnnnnnc	ccnnnnnnnn	ncnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	1320

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nnnnnnnnnt ntntngnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn gnnnnnnnnn 1380
tnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn 1440
nnnnnnnnnn nnnnnnaaaa an 1462

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<210> 22
<211> 1601
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)...(1601)
<223> n = A,T,C or G

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```

<400> 22
cccgaagcac gacgcagagc ctccggtgtg gctgtctctg atggtgtcat caaggtgttc 60
aacgacatga aggtgcgtaa gtcttcaacg ccagaggagg tgaagaagcg caagaaggcg 120
gtgctcttct gcctgagtga ggacaagaag aacatcatcc tggaggaggg caaggagatc 180
ctggtgggcg atgtgggccg gactgtcgac gaccctacg ccacctttgt caagatgctg 240
ccagataagg actgccgcta tgccctctat gatgcaacct atgagaccaa ggagagcaag 300
aaggaggatc tgggtgttat cttctgggcc ccgaggtctg cgcccttaa gagcaaatg 360
atztatgcca gctccaagga cgccatcaag aagaagctga cagggatcaa gcatgaattg 420
caagcaaat gctacgaaga ggtcaaggac cgtgcaccc tgcagaaan ctggggggca 480
gtgcccgta tctccttgaa ggcaaagcct tttgtgaacc cccttctggc cccctgcctg 540
gaagcatctt ggcaagcccc ccnccctgcc ccttgggggg ttgcnaggct tgccccctt 600
ccttccana accggaaggy gcttgggggg gatccccan caggggggga aggggcnant 660
ccctttnccc ccannnttg ccnaaacng ncccccccc ncccccttg nanttttcc 720
nttnttnccc ttccatncc cntttngng ggtntntng gncctttcc ccnaanntg 780
gggnttttn gnaancntt tttnaaann nccntnttt gggggnctnn nnaaannccn 840
naanccccna ngntntncc cccccccn ngggncccc cccccnnt ntntnnng 900
ggggggggnn aaanccccn nnnnnnnnn nnnnnnnnn nnaaaaaaa aannantcn 960
cccccnntt ttccccccc nccccngg gggnnccnn tccccccn ttttttccc 1020
naannnnnt gggnnncna anntttttt tnnancccn cnnntnnnn nnnnnctcn 1080
nngnnnnnt ttncnntnt nttnnnnnn nnnnnnnnn nnnnnnantn nnaannnnn 1140
nnnngnnaaa acnatncccc ctncctttn cccnngggn ncnnnnncct ttncccccn 1200
nnnnnnnnn ttttnccngn nnnncnnna nggnccttn nntnaannn nccccctcc 1260
nngnnngnn nccccangg nganaantg ggnncccc cccnnngcn nnnnaanttt 1320
nnnttgggg gnnnnnccc ccgcgcgc ctcnctcc ccttcgcgc gccccgcgc 1380
gccgtccgc ccgcccccc nctccnctc ccgcgctc ctnccttnc tctccnccg 1440
gccccgcgc cgcgccgct cgnctncg ncncnnnn cennnnnnn nnncgnnnn 1500
ananaagnn ccnaccnat ccccccgcc nccccccnt nccgnnnnng nnnnnnnng 1560
nncgcccnc cccccncc cccnttcgn cccccctt n 1601

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<210> 23
<211> 1566
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)...(1566)
<223> n = A,T,C or G

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```

<400> 23
ttttttttt tttttgattt ttttaaatgc tgcacaacac aatatttatt tcatttgttt 60
cttttatttc attttatttg tttgtgctg ctgttttatt tatttttact gaaagtgaga 120
gggaactttt gtggcctttt ttctttttc tgtaggccgc ctttaagctt ctaaatttgg 180
aacatctaag caagctgaag ggaaaagggg gtttcgcaaa atcactcggg ggaagggaaa 240

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gggttgctttg	ttaatcatgc	cctatgggtg	gtgattaact	gcttgtacaa	ttaccgtttc	300
actttttaatt	aattgtgctt	aaggctttaa	ttaaattttg	gggttccctt	cttagagcag	360
ctcgtactga	cgaagggtga	tgcgctgaat	gatgtcacgg	cagtcgttga	acacacggcg	420
gatgttctca	gtgtcccagc	gcangtgaaa	tgagggttagc	agtagtgacg	cccatctcca	480
ctggcagtg	tgatcctcag	aaactcatct	cgaatgaagt	acttggcccn	ggtcacgcgt	540
gggttctctt	cnggctcngg	agtancatnc	tcangagtag	ggtagcgagc	aaattctgga	600
aagaagcctc	aatcttcnat	ttcccncaa	ggactttctc	ancganccan	atcttgcttg	660
tttganggaa	ccaggaatcc	cngnnnaatg	gngcncaacc	ccttcttggt	ggttncccaa	720
aangcccntt	gaaaaaagg	ttcaaaaanc	cctccctgcc	anggccgggg	ttngggncct	780
gggnttgncc	cccccccg	naaaaaancn	ctnntttnnn	naaancttgn	nttggnttgg	840
ggnccecccc	ccccnaaaaa	aaaanaaaag	gggnnnnnnn	cnccccnnt	nnttttnaaa	900
aanaccceng	gggnannccc	ccctttttgg	gggggggggn	tnnntttnnn	nnncnnnggg	960
ggcccccccc	cccnnnnnna	aaanaattnt	ggggaaannn	nnnanntttt	ttnncccccc	1020
ccnnngnnaa	aantnnngnn	tnnccnaaaa	tnncccnaaa	nnnnnngccc	ccnnnnnnnn	1080
aaaannnnnn	nntnnnnnnn	nnnnnaanaa	nnnnnccenn	tntannncnn	nnnnntnncn	1140
naaaanngng	gncnnnnann	nnnnnnnnnn	tngnnnnnnn	nnnnnnnnnn	cnnttttttn	1200
ccnaaanntn	nnnnntnnnn	nnnggggggn	aannngncnn	cnccccccna	annnnccnc	1260
nnnnggggn	nccccnnngg	gccccnnnnn	nnnnccnngn	nnnnnnnnnn	nnnnnnnnnn	1320
nnnnnnnnnn	nncccnngnn	nnnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	1380
nnnnccnnnn	nnnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	nnnnngnnncn	1440
nnngnccnc	nnnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	1500
nnnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	1560
nncccc						1566

<210> 24
 <211> 651
 <212> DNA
 <213> Homo sapien

 <220>
 <221> misc_feature
 <222> (1)...(651)
 <223> n = A,T,C or G

<400> 24						
cgctgggttg	cgactcccg	acgtaggtag	tttggtgggc	cggttctga	ggccttgctt	60
ctctttactt	ttccactcta	ggccacgatg	ccgcagtagc	agacctggga	ggagttcagc	120
cgcgctgccc	agaagcttta	cctcgtgac	cctatgaagg	cacgtgtggt	tctcaaata	180
aggcattctg	atgggaactt	gtgtgttaaa	gtaacagatg	atttagtttg	tttgggtgat	240
aaaacagacc	aagctcaaga	tgtaaagaag	attgagaaat	tccacagtca	actaatgcga	300
cttatggtag	ccaaggaagc	ccgcaatgtt	accatggaaa	ctgagtgaat	ggtttgaaat	360
gaagactttg	tcgtgtactt	aggaagtaaa	tatcttttat	tagagaaagt	gttgggacag	420
aaagtacttt	atgtaactaa	gtgggctggt	cagaacttan	aggcattttt	tgtaatttct	480
ttttaattac	tttananaag	tagggatgca	aatgttttca	gttagaaagc	ctttatttac	540
ttttggaaat	tgaacaanaa	atgctttgtc	ttanaactgg	agaatatttg	atggtaggga	600
aacatgtaat	ggttctctgg	caaaattggn	tcannatttg	aatgaaann	n	651

<210> 25
 <211> 676
 <212> DNA
 <213> Homo sapien

 <220>
 <221> misc_feature
 <222> (1)...(676)
 <223> n = A,T,C or G

<400> 25

gggggacaga	gactcagatg	aggacagagt	ggtttccaat	gtgttcaata	gatttaggag	60
cagaaatgca	aggggctgca	tgacctacca	ggacagaact	ttccccaatt	acagggtgac	120
tcacagccgc	attggtgact	cacttcaatg	tgatcatttc	ggctgctgtg	tgtgagcagt	180
tggacacgtg	aggggggggt	gggtgagaga	gacaggcagc	ttgnanntnn	ttgcttngan	240
ntttcncnta	naacccgcna	gcgcttnggt	agggtnnngc	anggatgncn	nncntttntc	300
nnaagncncc	ngttcngngt	canttgcttg	ntcntctaa	ctcnnnnnnc	ccccnttnn	360
gtctcctnng	ngntcnaccc	nnctctgntc	ttngntcnng	nttgnccctg	nnnttnnttc	420
nnngctcngc	ncgtntgggt	nnntgngnat	nannctnanc	gngtttntnn	attntnnctn	480
ncgtngancn	catntgancc	ttntnnngnt	nttcgnetnn	ntcgancgtn	ttcngggncn	540
cncncgnnt	ctnnctnncc	tcnccctttt	ntcntcttgn	ttgtggcntn	acctnnctcn	600
ttctntgtnt	ncnngccttn	nngtgnnncn	gatagtcnnc	cctntttggn	aatatctntn	660
tnntcncccc	cctccc					676

<210> 26

<211> 657

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)... (657)

<223> n = A,T,C or G

<400> 26

tttttttttt	tttttgctgg	gtggtaactc	tttatitcat	tgtccggaag	aaagatggga	60
gtgggaacag	ggtggacact	gtgcaggctt	cagcttccac	tccgggcagg	attcaggcta	120
tctgggaccg	cagggaactgc	caggtgacac	gccctggctc	ccgaggcagg	caggcaaggt	180
gacgggaactg	gaagcccttt	tcnagccctt	ggaggagctg	gtccgtccac	aagcaatgag	240
tgccactctg	cagtttgag	gggatggata	aacagggaaa	caactgtgcat	tcctcacagc	300
caacagtgtg	ggtcttggtg	aagccccggc	gctgagctaa	gctcaggctg	ttccagggag	360
ccacaaaact	gcaggtagtg	atgtgcaaga	ntccatcctg	cagttttcca	gcaatganaa	420
actcctcctg	cggttgtggg	acctggggaa	gtatccgcan	acctctcctg	gcgggggtgt	480
agacnaaccg	gatgtcaccg	gcaccccta	aagnttgga	ccctttatac	atcttgggca	540
tcttgancct	ataacgctgg	tataaggngg	ntnggtngac	ttttggmngt	cccccaant	600
gcccttgana	ccaaggccgn	aattncnaaa	ggcccctgng	gggggggggg	accagn	657

<210> 27

<211> 646

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)... (646)

<223> n = A,T,C or G

<400> 27

ggaangctga	agaattaaca	ntttgactnc	taaatgtgat	actggntngt	anattccctt	60
agagcagaaa	ggagaggggc	acatattaat	ttgtatcgct	tttgcttctc	tttggctttt	120
tgtgtcttag	aatttggaag	tggttcattt	ctgttgctgg	tatgaggatt	tcgaataact	180
agtaatcgaa	aaccatatcc	tgtaatttaa	taaaaaaac	taaggaagaa	aaaaccctcc	240
aattttccca	aatgcaatca	gtgtaactag	gggctgtgtt	tctgcattaa	aataaatggt	300
tcangctttg	tggctctgat	caaggctctc	attaaaaaat	tggagttcac	cctagnctt	360
ttccccctctg	tgactgggct	cntccccac	cnetcttagg	tatcgagtt	attatgggnt	420
ncaaatnaag	naatangntt	nncaaattn	accaaanaaa	gcattttttt	cactgcnttn	480
tnattggggg	gttggcccaa	ccnctcaat	ggntcttanc	atggntggnt	acccgcnacc	540
tttncntnaa	cttggngnaa	ncnnggcnn	tacnnttct	gggggnaaat	ngtntccnnc	600
cantccccnc	ncntncnanc	cgaancnaa	agggnaancn	ngggg		646

<210> 28
 <211> 407
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(407)
 <223> n = A,T,C or G

<400> 28
 caagagtctt tgaaataagc ccatttgagc cctggataac aagggataaa gtggagcggga 60
 tgcacatcac agacatgaaa ttgcctcacc tgcctggcctt agaagacctt ggtattcagg 120
 caacaccact ggaactcaag gccattgagg tgctgcggcg tcatcgcaact taccgctggc 180
 tgtctgctga aattgaggat gtgaagccgg ccaagaccgt caacatttag tgcctcctga 240
 gcagctcttg gttttggcgt cttttgggtc ggcccatgtg gtttgagcac ccagccaggc 300
 ggtctcttta gaggatcctg tacacagttc cactattaaa acatttcagg ttgaaaaana 360
 nnnnnnnnnn nnnnnnnnnn nanannnnnn nnnnnnnnnn nnnnnng 407

<210> 29
 <211> 625
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(625)
 <223> n = A,T,C or G

<400> 29
 tttttttttt tttttttttt tttttttttg gggaccaaatt ttottnnttt gaaggaatgg 60
 nacaaatcaa acgaacttaa gnggatgttt tggnacaaact tattgaaaag gnaaaggaaa 120
 ccccaacatg catgcactgn cttggggacc anggaagtca cccacagggt ntggggaaat 180
 tancccnagg nttanctttc attatcactg nntcccangg ngngcttgna aaanaaanat 240
 tccncccgag cacattnngg cncctccatn ttgcncaggt tggncacgtg gncaccaat 300
 tctttgaagg ctttcaccng ctnattnaag naangggctt caatgaaanc acaccantgg 360
 ggggnatttt tgntnnnngc ccattgggca attcccaana tggctgaatc aaattttttt 420
 nccaaagnca ngcccctcca atggattnaa anccccntnc caatanaaca nnnngntttt 480
 ttatcctcca agaaaaattn ggcccntntn gggntggaag gtttnantat tacaagcncc 540
 ttccttttaa tggggaaaaa nttttgnnaa annttaaaac cncntcgcca agntttnaaa 600
 aggggnaggna ngcngnggggt tacnn 625

<210> 30
 <211> 643
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(643)
 <223> n = A,T,C or G

<400> 30
 cttagaatt ggccagcct cagatcctgt ctttagcaac cagctaatat ttaccagag 60
 gtactgcaat agagtatttc aaaatggaat caggatctgg tgggcctcag aaattgtctc 120
 tttctgagt ttcaatttgg ttctcctgga tgttttgctc tgttttggtta cctgtaatat 180
 agggaaacac aacttttttt gggaaagccc tttagaccca gcttgctagt tgcataataa 240


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taaattttct gttcctaaaa aaaaaaaaaa aaaaaaaaaa aaaaanaaaaa aaaaaaaaaa 300
aaaaaaaaaa aaggnngnaa naaaaaaaaa ananggggcc gntaaaacnn ggggggggcc 360
cntcaanttt aaagggccct ttaaancccc tnnnnaancc nccntgggcc nttttnnttc 420
ccaccttttg gnggnngnc cncccccgg nctttttttg noctgggggg ncccccccc 480
tggtcnttnc ttanaaaan nangaanttg cctcccttnt cngaaaang ntcttttttt 540
ttnggggggg gggggggggg ggaannnggg ggggggtggg ggaaaaattn nggggntttg 600
ggaaccnggg gcccttgccc ttnggaaaag aaccntggg ttt 643

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<210> 31
<211> 645
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)...(645)
<223> n = A,T,C or G

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<400> 31
gtgaaagctg taaacacct tttatggaag aaaagaaata aaatgtagtt gtcaagtcta 60
aaaaatagta gaaacgggaa tcataatgaa tacatgcaat gaatttaaaa tgtaaaaatg 120
aattttaaaa gtaaaaagg ctctgtggtg taatttttct taactacaag agtctaaata 180
cactgctttt ctttaagagt tcattttaat tagtaacgtc aaacaaaatt attctagata 240
atgagcccta caaattacta ctactagcaa ctgtcatttt ttactcgggc atoctctagg 300
tgtcttacat tctcatttta ttcttacaac gaactcatcc tccagaagga cttcacctc 360
cagaaggact catcctccag aangactcat cctccaaagg acttctccag aagggggaaa 420
tggaagaccc gggtaaactg ctcagggctt atcacagaac tatgtttgag cctgacttcg 480
tttgaactct aaagcccaca tgctctttct actgccccat gcttctcaag gnaccagact 540
cttatttntc gcacttttga gaatctnaag atcctgantic attttaaata aatttagttt 600
tttggggagn agccnnaaaa aaaaaaaaaa ggcgcctcc ncnnt 645

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<210> 32
<211> 668
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)...(668)
<223> n = A,T,C or G

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<400> 32
tcccgttctg ttttaaacag aaaataaaag gagtgtgaag tccttttctc atttcaaagt 60
tgctaccagt gtatgcagta attagaacaa agaanaaaca ttcagtagaa cattttattg 120
cctagttgac aacattgott gaatgctggt gggttoctatc cctttgacac tacacaattt 180
tctaatatgn gttaatgcta tgtgacaaaa cgccctgatt cctagtgcc aaggttnaac 240
ttaatgtata tacctgaaaa cccatgcatt tgtgctcttt ttttttttta tggngcttga 300
agtaaaacag cccatnctnt gcaagtccat gtatgcngcn cttaagcntt ctatctttgc 360
tcaaantgnt gaangatggg gaccttggtc catggcttgc gnatttgatc ntaangnncn 420
tttctancta tgntatgagg cacnngccct attggaggnc gccccnggtt tccgaaaag 480
ngcnntnttg tngngaattg cnnctcggan ttcaanaata tncggcnntt gntttgnang 540
ccnngnnnan caatcaggng ngcccctcna antcatgnaa gccccgnntn aanncnctnc 600
nctnttctcg nnntgggnnt tccattgccn gcctcgacgn ggttngcctc tcnccggcnn 660
cncgcncg 668

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<210> 33
<211> 682
<212> DNA

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<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(682)

<223> n = A,T,C or G

<400> 33

ggcttgtccg	agttgatatg	cgtatgcttt	gcctaaaaag	ccttaggaaa	ttagacttga	60
gtcacaacca	tataaaaaag	cttccagcta	caattggaga	cctcatacac	cttcaagaac	120
ttaacctgaa	tgacaatcac	ttggagtcac	ttagtgtagc	cttgtgtcat	tctacactcc	180
agaagtact	tcggagtttg	gacctcagca	agaacaaaat	caaggcactc	cctgtgcagt	240
tttgccagct	ccaggaactt	aagaatttaa	aacttgacga	taatgaattg	attcaatttc	300
cttgcaagat	aggacaacta	ataaaccttc	gctttttgtc	agcagctcga	aataagcttc	360
cattttttgcc	tagtgaattt	agaaatttat	cccttgaata	cttggatctt	tttggaata	420
ctttttgaaca	accaaagtc	cttccagtaa	taaagctgca	agcaccatta	actttattgg	480
aatcttctgc	acgaaccata	ttacataata	aggattccat	atggctcttc	atattcattt	540
ccattccatc	tctgcccagn	atttggggat	acccgcanaa	aatttggggg	ttggggggaa	600
aaatntggnc	tggaactttt	tttanttnaa	gggaaataat	naggggngga	aggggggggt	660
ttntggnctg	cccccccccg	gn				682

<210> 34

<211> 1549

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(1549)

<223> n = A,T,C or G

<400> 34

ttgagagata	cctccctcct	tctgctcagc	tgccctgcag	taattaaact	ctttctctgc	60
tgcaacaccc	ctactgttct	ccgtgtattg	gctttttctg	gcagcaggaa	ggaaaagctg	120
atgcgatgct	ctcagtgccg	cgctgccaaa	tactgtagtg	ctaagtgtca	gaaaaaagct	180
tggccagacc	acaagcggga	atgcaaattg	cttaaaagct	gcaaaccag	atatcctcca	240
gactccgttc	gacttcttgg	cagagttgtc	ttcaaactta	tggttgagc	accttcagaa	300
tcagagaagc	tttactcatt	ttatgatctg	gagtcaaata	ttaacaaact	gactgaagat	360
aagaaaagg	gcctcaggca	actogtaagt	acatttcaac	atttcattg	agaagaaata	420
caggatgcct	ctcagctgcc	acctgccttt	gacctttttg	aagcctttgc	aaaagtgtc	480
tgcaactctt	tcaccatctg	taatgcggag	atgcaggaag	ttggtgttgg	cctatatccc	540
agtatctctt	tgctcaatca	cagctgtgac	cccaactgtt	cgattgtgtt	caatgggccc	600
cacctcttac	tgcgagcagt	ccgagacatc	gaggtgggag	aggagctccc	atctgtcct	660
ggatatgctg	atgaccagt	agggagcgcc	cggaagcagc	tgagggacca	gtactgcttt	720
tgaatgtgac	tggtttcccg	ttgccaaaac	ccaggacaan	ggatgctgga	tatggcttaa	780
cctgggggga	tgaaccaang	tttttgggaa	ngggaaagnt	tnaaanaaaa	tcccctggna	840
aaaaaaantt	tnnaanaaaa	accttgggan	ggggccccc	ttgggaaaaa	ngggggggan	900
nnngggtnt	tnngnccnnt	ttnnccccn	nnnnannnct	ttannnnngn	nnantttttt	960
nnaangggg	ntnnccccn	nttttnnaann	ntntntcccc	nnnnnanggg	ggggttnncn	1020
nnnccccng	ggggnncnnn	ntnaacnccn	nnctntnggn	ggaaancntt	tttttncttc	1080
nnccnnggnc	cccnanantt	tttcccagaa	nccccccng	ggggnngnnn	gaaangnnnn	1140
nnnccctcnn	gggggttncc	ccnnnaaaaa	aaannnggnt	ttttttttna	nganccgggg	1200
acnccccnnn	naaanntttt	tnnaaagegc	cccccnntnt	nnggnnnnnn	nggnannnnn	1260
nnnttngnnn	nttngcccnc	cnttnnnngn	nccnctcenn	nnnnnnnnnn	nnnnnnnnnn	1320
nnnnnnnnnn	nnnnnnnnnn	cntntanntn	ntgnaaaaaa	nggnnnnnng	nnnnnnnnnn	1380
nnnnnnnnng	cccccnngng	nnnnnnnnnn	nnnnnnnnnn	gggggngngn	ggnnngcnnn	1440
nnnnnnnnnn	nnnccccnnn	nnnnnnnnnn	nnnnnnnnnn	ncgnnnnnnn	nnnnnnnnnn	1500
nnnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	nnnncngng	nnggnnanc		1549

<210> 35
 <211> 1440
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(1440)
 <223> n = A,T,C or G

<400> 35
 ctaatctaag cctcaaactc gttattgggg ctataaagaa aacgtttact taccagctg 60
 aaacagggtta agaataattct taatctcatt atagataatt gccccattgg gacttgaaat 120
 acaacacctt gtgctgaaaa cttcagggtg gcaatatttg aaggtttcgt tgtagaagag 180
 ttttaacatta actcctatctt tgacttacaa atcttgtttc tcatcactaa aatgcttttg 240
 aattaataat ccaacccaca tgagctgaga gtttttcttt tgttagaaaa gaaacagaca 300
 tctttctgta tgaaagtata aattgtatgg ttttagatac ataagaattg acaaaagcga 360
 gcgaaatctt tgtacttctg agttcttgct gtatgtatgt tttgttttaa atctgattag 420
 ggacacccag cagctggccg ggattcttgg attgctcctt gggagttaag attgtcaata 480
 ctctgtgtaa gcaagggatt tcagccatag aacaaagatt tattgttgcc acctgaaaag 540
 tttacaagta tttattgtgt atttgataca ttgcttgaaa aagatgaaat ctgttaaaga 600
 ttcttttccg atgtccaggt taagaagaaa cctccttgta ttgagtgaat ttatatgtta 660
 aatgggtatta gagaatgtag gtggnataga aattggattt ttcttgngng tngaacaacc 720
 tcaagttcgg caaagtttaa aatttggatt aaacaagaaa aannggttca nggttgnaaa 780
 angggacttg nttagggang ggacaanggc ctttaaanna ccngcgtccc ttctccnggc 840
 nggcnnngcg ggcccnccc caanctnntc cangcttctg nccnccnccn nccncccttt 900
 cctnntncca cnaanntctt tnncctttt tacngggggn ggggnnnccn ncncgggcn 960
 cngnntnccg cncccanaaa nncnncntt ttccnncnnc ccttttncnn nnncttttnc 1020
 cnnncccccc cccgnnnnnn nnnnnnnnnn nnnnnnnnnn nggnnnnnnn cccnnnnnnn 1080
 nnnnnnnnnn nnnnnnnnnn nnnnnnggnc nngggnnnnn ttntntnnnn gggggnccnn 1140
 nnnnnnnngc nnnnnnnnnn ngnnnnnnnn nnnnncgnnc nnnnnnnnnn nnnnnnnnnn 1200
 nnnnnnnnnn ncccngnna ncnaanncn nnnnnnnnnn nnnnnnnnnn nnnnnnnncn 1260
 cnnnnnnnnn nngnnngnn nngnnncnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnngn 1320
 nnnnnncggn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn 1380
 nnnnnnnccn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn gnnncgaaga nggcnaccg 1440

<210> 36
 <211> 1496
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(1496)
 <223> n = A,T,C or G

<400> 36
 tgcataccgt ggaaggcg cagggtcttt gtggattgca tgttgacatt gaccgtgaga 60
 ttcggcttca aaccaatact gcctttggaa tatgacagaa tcaatagccc agagagctta 120
 gtcaaagacg atatcacggt ctaccttaac caaggcactt tottaagcag aaaatattgt 180
 tgaggttacc tttgctgcta aagatccaat cttctaacgc cacaacagca tagcaaattcc 240
 taggataatt cacctcctca tttgacaaat cagagctgta attcacttta acaaattacg 300
 catttctatc acgttacta acagcttatg ataagtctgt gtagtcttcc ttttctccag 360
 ttctgttacc caatttagat taagtaaagc gtacacaact ggaaagactg ctgtaataac 420
 acagccttgt tatttttaag tcctattttg atattaattt ctgattaagt tagtaaataa 480
 cacctggatt ctatggagga cctcgtctt catccaagtg gcctgagtat ttcactggca 540
 ggttgngaatt ttttcttttc ctctttgggg atccaaatga tgatgtgcaa ttcattgtta 600

acttggggaa	acttgaaagg	ggttcccata	tancttcaaa	acaaaaacca	aatggtgtta	660
tccngacgga	tctttttatg	ggtntctaact	agtactttnc	taattgggga	aaagnaanng	720
cttttnagttt	tgcnnaaatta	agtttggggg	aagggcnata	attaaaaatt	gagggccccc	780
tnacnaaaac	caactggggg	ngtntaacga	aaaaccctgt	tttnaaaagg	gggccttttn	840
ccccctnnnn	ngnatntna	nttccccnt	ttgccntttc	cntttttnnn	naaacttttt	900
nnntttttctc	cccnancnnn	naaangngna	nnnggtntcc	ccccnangtt	nnntttnttc	960
nnnnnannna	ccccccctt	ngnggnccn	nnngggcntt	ttctcntngn	naanngttnt	1020
nnnannccct	tttgcnnnnn	gggnnttgng	nttcggngng	ccnngggggg	nnnnccnnnn	1080
gnnngnnnnn	gannangann	nnnggnggnc	gtntnnnnng	ccgcggggnn	nnngnnnncg	1140
ngnnnnnnng	nnnnngnnn	cnnngnnnnn	nnngnnnnnn	nnnnnangnn	nnnnnnnnnn	1200
nnngngnnng	ngnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	1260
nnnnnnnnnn	aannnnnnnn	nnnnnnnnnn	ngnnnnnnng	nnnnnnngnn	nnnnngnnnn	1320
nnnnnnnnnn	nnngnnnnnn	nnnnnnnnnn	nnngnnngcg	nnnnnnngnn	nnngnnnnnn	1380
nnnnnnngng	gnnnnnnng	gnnnnnnnn	nnccgcnnn	nnngnnnnnn	cnnnnnnnnn	1440
gncnnnnnnn	cnnngnnnn	nnnnnnnnnn	nnngnnntng	nnnnnccggn	gnnttc	1496

<210> 37
 <211> 1604
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(1604)
 <223> n = A,T,C or G

<400> 37						
atgcagtcct	ggatggagcc	gactgcatca	tgctgtctgg	agaaacagcc	aaaggggact	60
atcctctgga	ggctgtgccc	atgcagcacc	tgattgcccg	tgaggcagag	gctgccatct	120
accacttgca	attatttgag	gaactccgcc	gcctggcgcc	cattaccagc	gacccacag	180
aagccacgcg	cgtgggtgcc	gtggaggcct	ccttcaagtg	ctgcagtggg	gccataatcg	240
tcctcaccaa	gtctggcagc	tctgctcacc	aggtggccag	ataccgcca	cgtgcccaca	300
tcattgctgt	gacccggaat	ccccagacag	ctcgtcagc	ccacctgtac	cgtggcatct	360
tcctgtgct	gtgcaaggac	ccagtcacag	aggcctgggc	tgaggacgtg	gacctccggg	420
tgaactttgc	catgaatgtt	ggcaaggccc	gaggcttctt	caagaaggga	gatgtggtca	480
ttgtgctgac	ccgatggcgc	ccctgctccg	gnttcaccaa	caccatgcgt	gttggtccctg	540
tgcgngatg	gaccccanag	ccctccttct	agcncctgtg	ccacccctt	tcccanccaa	600
tccattaagn	cannaangct	tgtanaactt	cactctggnc	tgtaaacntg	gncacntggt	660
nggtngggac	accttgggaa	ggaaaaatca	acncctcant	tgnaaaattg	gggtaangnt	720
tgccantcnt	gtttttaaan	gggacnagnc	gcgaggaagg	gctnantttn	ttanantnnn	780
agggggcccc	cnncccnat	nnanangggg	caaanaacgg	nanggnaaat	ngnttnnnnc	840
cttngnnngc	ccccccnnng	ganncccn	nnccngggnn	nnnnagnngg	gntcancnnc	900
ntncccttnt	nctnnntgng	gtnnnccnnn	nnnccnnnnn	cacgttnaaa	annnaaatnn	960
ngncccnnnn	gnngcctca	cncnnttngn	ggnnngaccn	anccaccnng	cnnatnggng	1020
ntggnagggn	ctctnccnca	aancantnng	gncttcgtna	ngngtgnnnn	nnnnnnnnna	1080
ncnngntnnn	nnccnnngc	nannnngttn	cnnngntccn	cccacttgtn	tnncnannng	1140
ngttnnnngn	tngannntcn	nnngntgnat	cccggnaana	cnannnnccg	ncnccnggcnn	1200
ncnccnncn	gnncnntccc	nnncccnatn	nnngnggnnn	nctgcnanct	nnnnngancn	1260
cnnnnnnnnn	gnncannccg	antngngnng	nnnnnnnnnn	nnnnnnnnnn	nnnnnnnnnn	1320
nnnnnnnnnn	nccgnttntg	ctngcagtag	ntnccngnnt	ntcnnnnnnn	ngnnnnnnnn	1380
ncnnnnnnnn	nctngnacnt	tngnacgcnn	nagtcgacnt	nctnggacnt	nnnnnnccant	1440
cnncccnngt	nnngnnntngn	ngcnnacnnn	nnnacnnngg	cgnnnnnnnn	ncatnnccnnc	1500
nctnanannn	ggtngngngg	nnncccttccn	nnnnagnnnn	natanngnccn	nnanncccn	1560
nnnnnnnnnn	ngnnnnnnnn	nnnccnccga	nanntgncac	nacg		1604

<210> 38
 <211> 280
 <212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(280)

<223> n = A,T,C or G

<400> 38

tttttttttt	tttttaattt	atcagngcgt	aaaaatcttc	aaaatagcgt	agtgaggctc	60
atgacagtgc	tgcccccatg	gaaatgtagc	cttttggtgc	gtttaaacac	tgtcacacca	120
tctatgactg	tcccattggg	ctgaagtgtg	gtggcaaact	aagcatccta	taagacaagc	180
taaagcttgc	tttttgccag	tcagttgaaa	gtcttgcatc	tcttcactga	tgcactttct	240
ttaggtattg	atagtcagaa	gcacaaagca	tttattatgc			280

<210> 39

<211> 378

<212> DNA

<213> Homo sapien

<400> 39

cgagtttata	atcctataat	gaagaatact	ggcacaggca	atgctcactc	gaaaacttca	60
agtaatttct	agttggtttt	ggaatgcttg	ataaaagttcc	tttacagcgt	tattttcctg	120
atttgttttg	gttttagatca	aagttcaaat	taatttttaac	ttagctaata	aactcatcac	180
caggacagtt	ggagggggta	ggccgaggtt	aaatgggtcca	cgtttcaaaa	atgttaatgg	240
ctaataccata	attaaagaag	gtttaactgt	tactgaagtt	tacaagtttt	attgtcatga	300
acatgaaata	caaacacgat	ggcttcgaaa	tgtctttcaa	taaatgtttc	tgcaatttata	360
tggaaaaaaa	aaaaaaaaa					378

<210> 40

<211> 2039

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(2039)

<223> n = A,T,C or G

<400> 40

caacttttgt	agaagtattt	ttttctctgt	aatattttta	ttggctcata	aagatgtttt	60
catactctga	ctcctaataa	agtgaatta	cagtagatta	tattaacaaa	atacttttta	120
ggtagccatg	cttgagactt	tttaaaaata	taactttttc	cttaaaagtt	tcagctatag	180
caaaaggtag	ttatgtatgc	cagaccta	atgagctgcc	accaacaccc	ctagaacttt	240
cagccatggt	gtcttcagaa	ttgtagcgca	tttctgaatc	tagcaaatcc	tccttttacc	300
cgttgaatgt	tttgaatgcc	ctgactctac	cagcgcccat	aaatgatctc	tagaaggact	360
gttagtacca	acctgttttt	caactttgaa	gctaaaaacc	ctgatattgt	aatattatgg	420
tgcatagcag	aggtctcgga	aaaaaaatat	ttctgttcac	tttactttca	ggttaaaaat	480
gtttctaaca	cgcttgcaac	ttcccttatg	gcattaatct	tgttgaggga	gagagacaga	540
atcctggact	ctccaaagta	tttaactgaa	agtagggcct	gctctgacag	ggcccatgtc	600
ccacaaggct	ggcttnggcc	tcaggggggg	gctttggctg	gtgcttggga	tgaaaaattgn	660
tggnancngg	tntttgggga	taaanggacc	aaanggacca	gccaaaagcn	aaaaaatngg	720
gnnttttaaa	ngccttgggg	ggnttacctt	tttcntttta	angnnggttt	naaagnatta	780
gggctaang	ccanttttnc	aaaaaangct	cccnananaa	aatgggtggaa	aagggnccct	840
tttgngcgac	aggncctttg	nggaaaattg	ccccancng	ggcccttttt	tgnccccccc	900
nncccaaaaa	aaagntgggn	ngaagnnttn	ttaaaaccct	nnngngngccc	ntttttttng	960
nnaaancnc	cncncngggg	gncgccccnc	ttntttnttt	ntnttcceng	ggngncennnt	1020
tttttncgg	cngaccncnc	gggntcaan	nnctgnanaa	gnngntatct	ggcnggggnn	1080
gcgcnngaaa	gnnnnnggnn	ncngnggggg	nnnncgcncg	nnannnttnt	ngggggnaaa	1140

aaaaaaganc	cctctnttnc	tctcttntnt	naanntnnnn	ngnnnnnnan	ncnngcnnnn	1200
gnngngnngn	nnnnnnngnc	nnncnnnnnn	ggggggnggg	cnncncnnc	nnnnantnng	1260
gggcgactcn	tnnnnnnccc	cactncgggn	nccnnnnncn	ggngngngcn	mntntngang	1320
tccgnntgt	gtntgnnnng	ncnnncncnc	cncgnnnnnc	tnnnnntntg	mntnngnnng	1380
ggggngnncn	nncccncncg	tgnnnnntnt	nnnnnnnnnn	nnganggna	nnncnnncnn	1440
nnnnnnnnnn	ggggngcnnn	nnccnnncnn	tnnnnnnngg	gnggnggggn	gnnnnnnnnn	1500
nnnggannng	nnnnnnnnnn	nnncncncnn	nnnnnttng	cgnnnnnnnc	nnccnnngnn	1560
nnnnntnnnn	nnnnnnnnnn	nnncnnnnnn	nnnnnnnnnn	nnnnnnnnng	nnnnnnnnnn	1620
nnncnnnnnn	nnnnnnngng	gnnnnancgn	tgngcngng	tnnnnnnnn	nnnnnnnnnn	1680
nnnnnnnnnn	nnnnnnngnn	nnnnnnnnnn	nnangnnnnn	nnnnngnnnn	nnccnnnnnn	1740
gnnnnnnnnn	cnntgcgagc	nnngncnnnn	nnccnnntgn	nnnnnnngnn	tcgcnccnnn	1800
nnnnncgngg	ggcgtntnnn	ncnccecgcn	gntgncnnnn	nngncnnnnn	ncnnnnnnnn	1860
ngnnntnnnn	cnnnnnnnncg	nnnnnnnnnc	nnnagngnnn	ngngnncnnc	nnccnnnatn	1920
gannnnnnnn	ncnncnnnnn	nnnnncgnnn	nngcnnngnn	ngnnnnnnnn	nnnnntcncn	1980
ncnccnnngn	nnngnnnnnn	nnncncncgn	gngnnnnngn	cccgtccgcg	cgngcgcg	2039

<210> 41
 <211> 319
 <212> DNA
 <213> Homo sapien

<400> 41	
tttttttttt	aaaaaaaaag agtttattta gaaagtatca tagtgtaaac aaacaaattg 60
taccactttg	attttcttgg aatacaagac tcgtgatgca aagctgaagt tgtgtgtaca 120
agactcttga	cagttgtgct tctctaggag gttgggtttt tttaaaaaaa gaattatctg 180
tgaaccatac	gtgattaata aagatttcct ttaaggcaga ggctgggtcga gatgctgctg 240
ttatcttctg	cctcagacag acagtataag tggctctgtt tctaagattc ctaccaccag 300
ttactttggg	ccaagtatc 319

<210> 42
 <211> 524
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)... (524)
 <223> n = A,T,C or G

<400> 42	
cctttttttt	tttttttttt ttttctgatt tcaagtcaag atttattgct ttacaaacaa 60
acattatact	tggctcttaat agaaaaatga caccagatac atccaaaata catttcacat 120
tgggatagct	gccagttcag cacaaaacat acattactag gagcagggag gcatgaaaat 180
aaactatata	ttactttttg gtacgtcagg aacacttttg cctgaagtaa gccctttagt 240
actatttttt	attttattta tttttttaat ccacccatct gcacactggn ccttttagtac 300
tctttaagta	taaaacttta cttgtcctgg gctttgaccc ttgtgtttga tctaaatgac 360
atttcaaaca	taaatgtctt ttgactagtg cgcttactgn tatgtacana atttaaaatg 420
tgatcgttng	aatntaaaat ctgggtttgat acatgatata aaagttgtat atttaaaatn 480
caagaaatgt	ttttggggaa tatttctact aaagaatttt aaat 524

<210> 43
 <211> 103
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)... (103)

<223> n = A,T,C or G

<400> 43

```
cctttttttt ttttttttgc nngaaataag gaatctataa atctgaaata aagaaatccc    60
atttttaaatt aaattgttaa agagacacat aagaaaaaac act                        103
```

<210> 44

<211> 425

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(425)

<223> n = A,T,C or G

<400> 44

```
gtgcacaaga taatgtactg acatctctag caatcttttt tgccagtggc tttaaattgc    60
caataagtta aagaatattg ttcttatggg ttaaattttt attcttattt tcacatttaa    120
atattttttt cttaattttt gtggatacat aatattgtgt tatatgtatg ccatatatgg    180
tatattttga tgcaggcata ctctatataa taatcacatt agaggaaatg agatatccat    240
tacctctagc atttattctt tttattacaa gncaattcaa ttgtacactt tttagttatt    300
tttaaattta caatgttatt gattacaggg tcatttttat ggtcataata aaaaatttta    360
tacaaaacgt gtaaaatcta tacatttctg agttctgaat aaatattttt taaaaatttt    420
aaaaa                                           425
```

<210> 45

<211> 492

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(492)

<223> n = A,T,C or G

<400> 45

```
gtcgactgcc cccacogctg ggcggcgctg cggggcaccc aggcctctgca gtcagcgccg    60
cgccgggaat cctgtaccgg ggcgggaata agtaccagac cattgacaac taccagccgt    120
accggtgcgc agaggacgag gagtgcggca ctgatgagta ctgcgctagt cccaccgcgc    180
gaggggacgc aggcgtgcaa atctgtctcg cctgcaggaa gcgccgaaaa cgctgcatgc    240
gtcacgctat gtgctgcccc gggaattact gcaaaaatgg aatatgtgtg tcttctgatc    300
aaaatcattt ccgaggagaa attgaggaaa coactactga aagctttggg aatgatcata    360
gcaccttggg tgggtattcc agaagaacca ccttgtcttc aaaaatgtat cacaccaaag    420
gacaagaagg ttctgtttgt ctccggtcat cagactgtgc ctgangattg tgttgtgcta    480
gacacttctg gt                                           492
```

<210> 46

<211> 499

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(499)

<223> n = A,T,C or G

<400> 46

cctttttttt	ttttttttat	aacattttata	taatgtgcta	acaatgaatc	catccatgat	60
ttattgtttg	taatgaactt	aaaataaccc	tttaciaaatt	aaaatcattt	tttcaaaccat	120
gacttcata	tgaatgggtt	ctgttaaaaa	agtaaaagtt	gaattttcca	gccaatntag	180
catctaggac	ctgaatcttg	ccaatatcct	accactatc	ttcattccta	cctcctaccc	240
cttcaaatca	gctcctccag	actttcctat	ttctgtcacc	ccagttcaaa	atgggtttca	300
ccatgcattt	gatgtaaaat	gtgcaagtgc	gatatgactt	cacaaagtat	caattgtgtg	360
gacaatgata	actactgtga	cactgctagc	accctgtggt	aaaagtaaga	agcaacaaaa	420
ttacacaggg	ttcctttctg	atgaatgcag	nanggattca	agaaatccca	ganctggaaa	480
aagattttca	atagatctg					499

<210> 47
 <211> 537
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(537)
 <223> n = A,T,C or G

<400> 47	
gtcgacattt	ttctgaggaa tagtttgtga ttccaatgca ggtgtcttca ttaccattac 60
ctctacactg	cagaagaagc aaaactcctt tattagaatt actgcacatg tgtatgggga 120
aaatagttct	gaaaggctag aatgatacaa gtgagcaaaa gttggtcagc ttggctatgg 180
agtggtggca	ataatctcta aacattccaa aagaccatga gctgaaccta aactcccttg 240
gaatctgaac	aaaggaatat aaaattgcca ttgaaaaact gaccagctaa tctggacctc 300
agagatagat	cagccagtgg cccaaagcca ttccaagtac agaaattata gagactacag 360
ctaaataaat	ttgaacatta aatataattt taccactttt tgtctttata agcatatttg 420
taaactcaga	actgagcaga agtgacttta ctttctcaag tttgatactg agttgactgn 480
ttcccttata	cctcaccctt tccccttccc tttcctaagg caatagtgca caactta 537

<210> 48
 <211> 556
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(556)
 <223> n = A,T,C or G

<400> 48		
gtcgactttt	tttttttttt ttagnnntat aaaatatttt atttacagta gagctttaca 60	
aaaatagtct	taaattaata caaatccctt ttgcaatata acttatatga ctatcttctc 120	
aaaaacgtga	cattcgatta taacacataa actacattta tagttgttaa gtcaccttgt 180	
agtataaata	tgttttcatc ttttttttgt aataagggtac ataccaataa caatgaacaa 240	
tggacaacaa	atcttatttt gttattcttc caatgtaaaa ttcatctctg gccaaaacaa 300	
aattaaccaa	agaaaagtaa aacaattgtc cctctgttca acaatacagt cctttttaat 360	
tatttgagag	tttatctgac agagacacag cattaaactg aaagcaccat ggcataaagt 420	
ctagtaacat	tatcctcaaa agctttttcc aatgnctttc ctncactgn ttattcagta 480	
tttggccagt	acaaaataaa gattgggtct caactctctc tttcattagt ctcaagngtt 540	
cctattatgc	actgag	556

<210> 49
 <211> 355
 <212> DNA
 <213> Homo sapien


```

<400> 49
gtcgaccgag cctctccac cctcagtcgc atagacttat gtgttttgct aaaattcagg      60
tattactgaa ttagcgttta atccacttcc tttcttcttc ttctaaaata ttgggcactc      120
ggttatcttt taaaattcac acagaaaaat tccgtttggt agactccttc caatgaaatc      180
tcaggaataa ttaaactcta gggggacttt cttaaaaata actagaggga cctattttcc      240
tcttttttat gttttagact gtagattatt tattaataat ctttaataat aggaaaagg      300
gaaagtattt attgtacatt attttcatag attaaataaa tgtctttata atacc          355

```

```

<210> 50
<211> 507
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(507)
<223> n = A,T,C or G

```

```

<400> 50
cctttttttt ttttttttaa aaaaaaaaaa ttctgtttat tgtaataatt aaataagagt      60
aaacatttta aaacatataa aaataacttt aaaatatagt aacactttac aaaatatgta      120
tctaattaaa aatacatata catagcatcc ctcaaaactat acaaatatag aatatatatt      180
catgaaattc tttanaaata taacatctat tctttgaata aagcttaaaa tttgtttata      240
attttcaaac taanaaaaga agtagngaat aatagctcca tccaatttat aattgtctta      300
aagagaatga ttatgtatca tttcttgctt gtcttttcta ataccagtc aatcacctgt      360
acagcattgt tgtttgctgt tttcttcatt tcttcaaata gacccttga aagtttttaa      420
gatccttttag atagaactta gagatttcaa agagacgctg gctgcatgca gtgaaacatt      480
catgagtcctc ggtaatactg ngtttct          507

```

```

<210> 51
<211> 538
<212> DNA
<213> Homo sapien

```

```

<400> 51
gtcgacgcaa aagtttgact aaactttacc tttttatagt ttcacttttt aagttatatt      60
tagaatatat tgatagatta taaattgatt gtgaaacttt tttctgaatt ttttcaacat      120
gttttactca gttacatgag ttaaaggata ttttcagtc tttctgactt aattgcagtc      180
tttaaaaaaa cccaccctat tgttctactt gttatatgct tattcatata gtaaattcat      240
ttcaagggtt atgccagtgg gtattattgg tgctttttga agttgagggt aaccatccag      300
gaagggtctg ttaatgttat gttcatctat aatggcatag gggaaatata tatattttta      360
atattgtaaa catttgact gaataacctt tttttcccc cctccgcaag caaaactggt      420
tgaacagcgg atgaagatat ggaattcaaa gctctaattg acctttttga agagaagttg      480
tggcttatgt ggagtttaca tgggcctctg atggaagaaa gctaactctg ttagtatt      538

```

```

<210> 52
<211> 504
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(504)
<223> n = A,T,C or G

```

```

<400> 52
cctttttttt ttttttttta aagtacaaat tcagtttatt catctgttta tgacacagta      60
cacaggaggc aaagtgtttc acatcataga cttcacttcc aactccttgg aatgttcatt      120

```

tctttggcgtt	acaggagaga	ctagacagga	aggccaggca	atgcttaggc	aactaaaatg	180
aggttggggg	taatgctaac	gtcacctca	cagggatggc	cacggggact	gttattcgca	240
agctggtttt	ctagacctgt	tagctggaag	catggtgagc	accatttctg	gacgctcagg	300
cogntcggg	cttcagtcac	ntccaccaca	caggtacagc	agcgctttct	ggtagtcgcc	360
cttagtgtct	tgctggatat	aatagtacag	ggacttgccg	tactttctct	tgaattcaga	420
cctaattttc	aacatgtcca	cttcactgng	ggagaccatg	attctgatca	ggacccttat	480
ctcgcgcccc	cttgccttc	atgg				504

<210> 53
 <211> 489
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(489)
 <223> n = A,T,C or G

<400> 53	
gtcgacttta	gatgtacagg
ccagagggaac	aagcatgtct
cccagccttag	agttcttctt
tcagctcaac	tcacagcttc
aaatgagggc	tcacattgc
aatgaagtga	ttctannatt
atgcaaagt	tttgaaatga
ctgctttcac	ttagtgtct
tgatatttt	
	60
ctctgccaag	atccatctaa
gccctttgct	ctggagggaag
accctgggag	tttcctgagg
tcgaagtatt	caataccgct
agcatatgca	gccaaaccaag
ggaaagtcac	ccaacacactt
actgtaggaa	caagcatgat
cttgntactg	
	489

<210> 54
 <211> 577
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(577)
 <223> n = A,T,C or G

<400> 54	
cctttttttt	tttttttttt
aaggaaataa	tcacctacaa
gcagactgaa	aacatttccc
taatgcataa	tgtttactcc
gtgccanaaa	ctattctaac
tattgtcatt	ggtattacat
tatagaatta	tagataatga
agaatggggg	aaggataatt
aaaaggattt	aagataggta
cttacactag	gaaagacttt
	60
tacatggcgtt	ttaattattg
tctcaagttg	gcttacaaat
aatggttggg	ttcctttggg
catgaattga	acatttcttt
gaacaaatag	aaaaaaatcc
gaataaaaagt	aagtaaaaata
aaagctgggt	aaagggatag
tgtagtagta	tctctgttct
aacctaagta	ttacaaataa
gcattat	
	577

<210> 55
 <211> 483
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature

<222> (1)...(483)

<223> n = A,T,C or G

<400> 55

cctttttttt	tttttttcac	caataattat	tttattcagg	gagtaaagt	tattaattgc	60
caaaatacga	attttaaatt	tgagaagtac	agatttgtaa	gtatatattt	gtttgaaatg	120
tatcanattg	gccttttatt	ggcttattgg	tatttagngc	cagcacttac	aatgtgaact	180
cagcaacaga	agataattct	tatgaaatca	acattcaact	tacatgaaat	aacttaaaaa	240
cttaccaaca	atagtctaata	gattatatac	ctttaccaa	caatgtctaa	tgaaagtcca	300
aatgtaaaaa	tttaaaaaatt	aaaattatag	aatataattt	ttacacatca	attgttttgt	360
agcaccatct	cgcaaagnaa	atatcatgtt	tattctgtag	ctaaaatttc	tcccacaaag	420
cagaaattgt	ttggaatata	caaaaagaca	acccattaac	aagtaacttt	aagtaatgta	480
ggt						483

<210> 56

<211> 521

<212> DNA

<213> Homo sapien

<400> 56

gtcgaccaga	cttaagcatc	gagtttttac	catottccac	tttaagctaa	gttatgatac	60
ctattccatt	cacaattggg	gttcttttta	aggtttgcaa	atttcagcca	atttttagc	120
taagattggt	ctgatcagct	caaaaagatt	tggcttagtg	ttttcattgc	aaattataat	180
tgctgtagag	ccacacacaa	cttttgaact	tttaattata	agtgttatgg	ctaaagttaa	240
ttactgaaaa	tttcagtaaa	atgtgtgaat	gtttctttat	gtattaacct	catagcagta	300
aatgacttgc	tggtgtttta	tttttctaag	gcactttaat	agacttctgt	tgaaaacttc	360
agtgttaaca	tttttatagt	ttgtactaaa	tttaaccgtg	atataaaaaat	gaattttatg	420
catagatcag	gaatttttaa	ttaaagggtt	tttcttttaa	aaaaaaaaaa	aaaaaggcgc	480
gccgctcgag	tctagagggc	ccgtttaaac	ccgctgatca	g		521

<210> 57

<211> 542

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(542)

<223> n = A,T,C or G

<400> 57

cctttttttt	tttttttaca	acttcacatt	ctttaatgtt	cattcagaat	attaaatgcc	60
attaattgac	catcattatt	ataaaattta	ctatttagat	aagtgaagtt	tagtacagtg	120
ctattttaaag	tatggaactg	ttactgggtg	gtgatcagta	cagaaattga	gactaagcat	180
ttagaaacct	agagcaattt	gacgtagcaa	tcttctgtct	gttgaatcta	ataacaaaaa	240
aaattttttt	aattttgcat	atctttttta	aatttaattt	gtcaaggaat	tcatttttag	300
catatttttac	aaaaacatca	ttctcctatg	gagactattt	ggaaatacaa	ataagaaaac	360
tggttctttac	cacagatagt	tttttagaac	ctgttttagn	gtaaagccat	catttagtat	420
aaagncatct	attattactg	ttactctgaa	gtgggtactg	agcattacaa	cagtnggtng	480
gattataagt	ttgttttacta	aanatgctag	gattttattaa	ctcatgtata	tattttattga	540
ga						542

<210> 58

<211> 261

<212> DNA

<213> Homo sapien

<400> 58

```

gtcgacagag aaggctctatg tcaacagagt tgttatctca tagagccagt tttcaaagct 60
ccttctgcat tgtcactcac tgatcagggtg atgaattctt cctagatagt cgccccactcc 120
acctcctact taacctgaga ctcatatttt agctatttct gcttttgtaa aaataattca 180
gatattaaac tccaatttta atctatcatc caagggtaga tgtagttgct tagtagcatt 240
ttggaaaaaa aaaaaaaaaa g 261

```

```

<210> 59
<211> 480
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(480)
<223> n = A,T,C or G

```

```

<400> 59
cctttttttt tttttttaaa atatagaagt tctgagttag acctgttttag ctcanaatag 60
tgggctaaac taccataaaa ttctctgtat atcttaaagt gtaatgggtc aaaaactcca 120
gaaaatcatc agttgataac acacctacag ataagtgcac gggtaggagg ggatagccaa 180
gtgcccatga taatttgacc tcagtaaatt aaactgggca atacacatat ttgctattct 240
gatactgcat tagacttata aaattccatc taataagcat tcataaaaact ggacctctct 300
gtatatatct agcttagaca gggataggga aaagaataac tgaagaaact agcttacaat 360
agctagggtt cgtcagggtt attctatcca gccagaaacc accaccagag agaagctgag 420
ccattcagct gnetgtctcc tctccctctg ttgtaatagt catgcctagg ccttgcctgca 480

```

```

<210> 60
<211> 493
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(493)
<223> n = A,T,C or G

```

```

<400> 60
cctttttttt tttttttggt ccttctgttt atttcatttt ggatactcag tgaatgttaa 60
ttaaccagga aacttaaaag ttatttcaat tatgaacctc ttcaatcctt catcaattat 120
tttgagtatt ctggtcttaa aaacatctct ttcttctaca aacttctgaa agagatgaac 180
acctccacct acaccaaatt aatgtgcttt gctggccaaa agtacacgtc catttttact 240
taacagtcta aggaaagtct ggtgcaaatt actataataa tctgggttgt aaatgggttc 300
tgagggtgaga atgagatcat attttacaaa aagtttttca ctacttagta caagcttaca 360
aaactcagac cactcaccag aaaaaaatcg gcatttatat agttgngtta cttttgggtt 420
cctgcatctt ttcacatctg gctcatttac atcattttct tcatcttcca aagtggagtt 480
agctactaca tta 493

```

```

<210> 61
<211> 532
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(532)
<223> n = A,T,C or G

```

```

<400> 61

```

```

tttttttttt tttttttgaa aaatataaaa ttttaataaa ggctacatct cttaattaca 60
ataattattg taccaagtaa ttttccttaa atgaactott tataatgcat aatttacagt 120
ataagtagaa caaaatgtca tgacaaaagt cattgagtag aagacttgta ataaaaaggc 180
ataaaatata ttatatacata aacccttttc aaaaaacaag ggaaagcttg agccctcaat 240
atagggcgac acacggagcg ggtgaccgtg caggtagagg tactgtactg atttaaagtc 300
aagcactaga gatagnggat taatactctt ttgccgtaca ctatatacag atgtatagta 360
caagtaacaa tggcaaacag aatgtacaga ttaacttaac acaaaaaccc gaacatcaaa 420
atgaagggtg gtggaggaaa ggtgctgctg ggtctcccta caactgttca tttctttgng 480
gggcaggggg tagttcctga atggctgngg tccaatgact aatgtaaaac aa 532

```

```

<210> 62
<211> 567
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(567)
<223> n = A,T,C or G

```

```

<400> 62
gtcgactttt tttttttttt taagtatttt aggcataattt aataaataac ttcagtaaatt 60
agcactgtaa aaagtgaact gttaaaacta aaggcactta aaacaagaat gtgactagtgt 120
tgaaacaaga tgggcaactc aaatggtgag aagtaaacad acagtgggtct gttatggcac 180
taactcaaag taagactcgc gtaggtgaga gctgttgcat agccacagta taacttcaca 240
tgttcattaa aaaggcaaat tgaccgctaa aacttcaaag aaaaagtact cataaaaaaaa 300
gtcttacctc aaaattgcaa acaaatacat taaaagatta gaagagggtga tagaaagcac 360
cagacattaa acaaaataaa aataataaaa taaattcaac tcaaaagggtc cccattcagc 420
aaatactttg taaaagtatg gcctgtatgt aaatagttgc taaatcaagg acttttttagc 480
agaaaattgc tcggtttctt tatctaaggc ttgaatttgt aaagngaagg cataaaagtt 540
nccaaacatt aagtaactct taaaatg 567

```

```

<210> 63
<211> 247
<212> DNA
<213> Homo sapien

```

```

<400> 63
gtcgacaaac aaacttggct tgataatcat ttgggcagct tgggtaagta cgcaacttac 60
ttttccacca agaactgtc agcagctgcc tgcttttctg tgatgtatgt atcctgttga 120
cttttccaga aattttttta gagtttgagt tactattgaa ttaatcaga ctttctgatt 180
aaagggtttt ctttcttttt taataaaaca catctgtctg gtgtggtatg aaaaaaaaaa 240
aaaaaag 247

```

```

<210> 64
<211> 330
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(330)
<223> n = A,T,C or G

```

```

<400> 64
cctttttttt tttttttttt tttttgacat ggagtcttac tctgtcaccg aggttgaggt 60
gcagtagtgc aagctcggct cactgcaacc tcaggcagga ctatttttta ttatttttaa 120
tacctgcaaa aggggaatct cacatgcaca tccgtgtttc tacanaaatc tgcgatcgat 180

```

ggcagatctg	tttgcccttg	ngtgtccaca	tgaaccattt	ggcaaaggca	tccaatgcta	240
acggggccca	ccaactacaa	cggaggcaac	aactctgnng	attnnttttc	acagaaagag	300
taaaatttca	ttcaaccgtt	ccatgtcgac				330

<210> 65
 <211> 486
 <212> DNA
 <213> Homo sapien

<400> 65						
cctttttttt	tttttttact	aggcaaagaa	ctttattaat	ctttgtttca	aacttgattc	60
ccaggcttct	toggcttaat	tagctgcaaa	gaatgaattg	tgtataagca	aaaactgaaa	120
agagctgcag	tgtccaaggg	gcttgggctt	aaaaatatta	gagatctaga	ttttatcaga	180
tccataaaca	aaaatttctt	aaaaagcagt	cataatataa	aatagcagct	cccagtaact	240
tcttcaggtt	ttatcttcag	aagttgactc	aattcagttt	gcctcattct	tggaaacctc	300
atcaaaattc	tccacaagat	ctggaacttc	atcatcatca	tcctctccag	tagcaagtgg	360
tgcttttcca	tccacagatt	gtttgggcag	agcttcggcc	agtctcctta	aactagtcag	420
actatccgca	ccaagctggg	ttaagatgct	gggtagcatt	tctgtcagct	gctttgtctc	480
agcatg						486

<210> 66
 <211> 503
 <212> DNA
 <213> Homo sapien

<400> 66						
gtcgaccgtc	agacagcaac	tcagagaata	accagagaac	aaccagattg	aaacaatgga	60
ggatctttgt	gtggcaaaaca	cactctttgc	cctcaattta	ttcaagcatc	tggcaaaagc	120
aagccccacc	cagaacctct	tcctctcccc	atggagcatc	tcgtccacca	tggccatggg	180
ctacatgggc	tccaggggca	gcaccgaaga	ccagatggcc	aagggtgctt	agttaaatga	240
agtgggagcc	aatgcagtta	cccccatgac	tccagagaac	tttaccagct	gtgggttcac	300
gcagcagatc	cagaagggtg	gttatcctga	tgcgattttg	caggcacaag	ctgcagataa	360
aatccattca	tccttcgcgt	ctctcagctc	tgcaatcaat	gcattccacag	ggaattatatt	420
actggaaagt	gtcaataaag	tgtttgggtg	gaagtctgcg	agcttcgggg	aagaatatat	480
tcgactctgt	cagaaatatt	act				503

<210> 67
 <211> 519
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(519)
 <223> n = A,T,C or G

<400> 67						
cctttttttt	tttttttgaa	ttaaattttt	ttttattttt	acaccataat	ccaattctag	60
ttatcttaat	tgaatttgaa	aactttttca	attgcattaa	atttacaata	aagttctccc	120
acattacact	aaagcattcc	tcatgtttca	cttcagctac	tcagatactg	aatgagtaaa	180
atcattttat	tggctctctt	ttaatctaat	ccttcaaatg	cacattgttt	aaaaactgac	240
taggtcaaaa	atagttacnc	ctgcagggtg	acctattcag	actttgccaa	actcctccaa	300
gttcaatata	aattgacgtt	ttcagagctc	aaagtcaatt	ttacggaaac	gctgttcctc	360
cttttccatg	gagccaatct	gggttaattt	ttcatttaaa	ttcttcttct	gcctgtttgc	420
tgcggaactc	tttgagctgc	tgtagccgct	cgatagtttc	anaaatgggtg	cgttccccgt	480
ggaccttatt	gtcctcttgt	goggatatna	acagtgccca			519

<210> 68

<211> 495
 <212> DNA
 <213> Homo sapien

<400> 68
 gtcgactaaa gctgaagaga taaaagaggt tgtggggcta tgtcttaaga caaaagaaca 60
 tttagaaaac ctcaggaaat gatcagagtg ggatagatgt tactagaaga aacaaagaaa 120
 ttgaattcaa ttaggagtta gaatcattta caaagcaatg gggaaagtaa gccctaaaa 180
 actattgtag catatagtaa ccagagccaa actctcataa tatattcccc aaggcaaaag 240
 aaaaatattt acaagatttg cgttgtttta tatgtttgca aacttattta ataagtctgg 300
 cttttagatg ttcatatctg agtctgcatt caatcaaaat gtcttggcta aacttcatga 360
 aaaaacccca gcctcataaa ttagtagttg gaaaaggag gcatatttag agctttttca 420
 gataattgta tttctttgat acattagact ggacacacag tagtttgttt aaggtttaatt 480
 gcaatattgc aatga 495

<210> 69
 <211> 525
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(525)
 <223> n = A,T,C or G

<400> 69
 gtcgacgcca ccatgttcga ggcgcgcctg gtccagggct ccatcctcaa gaagggtgtt 60
 gaggcactca aggacctcat caacgaggcc tgctgggata ttagctccag cgggtgtaaac 120
 ctgcagagca tggactcgtc ccacgtctct ttggtgcagc tcacctgcg gtctgagggc 180
 ttcgacacct accgctgcga ccgcaacctg gccatgggag tgaacctcac cagtatgtcc 240
 aaaatactaa aatgcgccgg caatgaagat atcattacac taagggccga agataacgcg 300
 gataccttgg cgctagtatt tgaagcacca aaccaggaga aagtttcaga ctatgaaatg 360
 aagttgatgg atttagatgt tgaacaactt ggaattccag aacaggagta cagctgtgta 420
 gtaaagatgc cttctggtga atttgcacgt atatgccgag atctcagcca tattggagat 480
 gctgntgtaa tttcctgtgc aaaagacgga gtgaaatttt ctgca 525

<210> 70
 <211> 511
 <212> DNA
 <213> Homo sapien

<400> 70
 gtcgacattt tatatataat actactaatg gcatagatta acaaaatatt ttacatgtag 60
 gaaaggacat aagattactt ttaaagaata gtatgaaata cacaatattc aaatgtgttt 120
 gcaatgccta ccaaatttca aatgtgcctg gatcatgtat aaattaagga aagaaaaaag 180
 gatcatgtat aaattaagga aagaaaaaat gtaagtatac aacctacacg gtaaaaaaaa 240
 aaaccaaaca cctggttaaa aatatctatt taagctcgag tgtataacct taaacaattt 300
 gtgtatcact agaaaaatgg atttatttagt aaaatttagg gcagagattt tattttggac 360
 accactgcct ttgtagaaaa atccaaagtg gcataaaaag aaaaataaaa tattaaaaga 420
 aaaaatatat attatcattc ccatgttccc atcctgttac tagcattgct gttctggtgc 480
 atcaatcctg agtactctaa cttttgattt a 511

<210> 71
 <211> 464
 <212> DNA
 <213> Homo sapien

<400> 71

```

cctttttttt tttttttgga agagcttctt gcactgttat aagaaagaac atgtgggaga      60
ttgcaaacia agcaacataa agagtataca gcctgtagga gtctgactaa agtaaaaaaa      120
actcatgtct ttgttttagtg agtatctgta tactaagtta atgcaatgcc aattagattc      180
aaattaaatc aagtacaagc aaatgtactg aaagtattag gaatgcatca tctactttgc      240
taaataaatt gcactccgca ttctgcaatt acatgagcat gccattggtg taatattggt      300
tatataacat ttaacatgtt agtttttaaa agaattgtaga tacattcata gagatcagta      360
tttttacaga tgtttttact ataaaaggaa ccatgtataa cattgatttt taccttcagt      420
tttgataata ggctgaagac tgccttcaat cactttaatt ttg                               464

```

```

<210> 72
<211> 234
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(234)
<223> n = A,T,C or G

```

```

<400> 72
aataaaannt gaacaaaagg aaaagggtgga tataaagtgg aacctgtggg aaagaggcaa      60
gggctgcagg acagaagaga ctgggaactg caggggccct gggactcagg aggagatgct      120
gattcagctc ataggtgacc cagtcctggc cccggctgtt cccaagagaa ggctgtaagt      180
accaggggag gtggtgaagca ggatggagga aaaatcagag gactgggggt cgac          234

```

```

<210> 73
<211> 143
<212> DNA
<213> Homo sapien

```

```

<400> 73
gtcgactaaa taagtcaatt cctggaattt gaaagagcaa ataaagacct gagaaccttc      60
cagaaaaaaaa aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa      120
aaaaaaaaaa aaaaaaaaaa ggg                               143

```

```

<210> 74
<211> 533
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(533)
<223> n = A,T,C or G

```

```

<400> 74
gtcgacataa tctagggcatg aagagcaaaa atatcccttc cggagtcttt gaagctgaaa      60
atataaaaca aataaaaaat aaaaaaataa aaaccacaaa aaatgttgaa ccaaacctcc      120
ctgctaattc ccatgccac gttctttccc accctgttcc cagtcttctg acaaactgtg      180
tacatagcgg actcctcctt tctcctccga ggtggtttta aaggcttttt ggtgtataga      240
agtttgtcca ttgtaaaaac tccggattgc gttcctcccc gccttcggcc ccttccttc      300
cctaaagtga tgggctttct cttttctctt tttagtttac ccggtttctt ttttaagtaat      360
gtggaagaaa atggtttatt ttgtattgng gtattgaata ttgngttcct ttttatgagg      420
caaacctgat tgtaaacctt atgtaactat agactggaaa aaaatgagcc gngccaaaag      480
tctncccttc tgtttcttca gcacattgac ccatnncaca cacatacaca cca          533

```

```

<210> 75
<211> 485

```


<212> DNA
<213> Homo sapien

<400> 75

gtcgaccttc	cctaggctgt	ttctgctggg	cgctccgcga	agatgcagct	caagccgatg	60
gagatcaacc	ccgagatgct	gaacaaagt	ctgtcccggc	tgggggtcgc	cggccagtgg	120
cgcttcgtgg	acgtgctggg	gctggaagag	gagtctctgg	gctcggtgcc	agcgcctgcc	180
tgcgcgctgc	tgctgctggt	tcccctcacg	gcccagcatg	agaacttcag	gaaaaagcag	240
attgaagagc	tgaagggaca	agaagttagt	cctaaagtgt	acttcatgaa	gcagaccatt	300
gggaattcct	gtggcacaaat	cggacttatt	cacgcagtgg	ccaataatca	agacaaactg	360
ggatttgagg	atggatcagt	tctgaaacag	tttctttctg	aaacagagaa	aatgtcccct	420
gaagacagag	caaaatgctt	tgaaaagaat	gaggccatac	aggcagccca	tgatgccgtg	480
gcaca						485

<210> 76
<211> 417
<212> DNA
<213> Homo sapien

<400> 76

cacgctgggt	ttgcatcttc	aggagacgct	cgtagccctc	gcgcttctcc	tcggccaatt	60
cgcggaagaa	gtggctcacg	ccttccagag	ccacatcadc	gcggtcgaaa	tagaagccca	120
gagagaggta	ggtgtaggag	gcctgcaggt	acaaattgac	caggctgttg	acggctgcct	180
ccacgtcggg	ggaataatc	tgacgaatct	gggagctcat	ggttggttgg	caagaaggag	240
ctaaccacaa	aaacggtgct	ggcaggtccc	agaagcagga	gatggccgag	aagatggtcc	300
cggagggttc	aagcggagag	gaaatcggag	ggcggtcgga	ggctggaaga	gagtccccgg	360
atctgttccg	tccaaacact	gttgaagcaa	gagacagacc	cgcgggaccg	cgtcgac	417

<210> 77
<211> 547
<212> DNA
<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(547)

<223> n = A,T,C or G

<400> 77

gtcgaccttt	tattaagaat	atattttatc	aggcattttg	ataacaaact	gttactctaa	60
gtatagggtg	tttaccaggt	gtattttaaa	aagtaaatga	atcccactgt	agtttttctt	120
gaaggaaaaa	tcattttctc	agttgctgag	gggtactaaa	agcttcatac	acattagcag	180
caaagtcttt	cacttgctcc	attgtcaaca	gacacctgaac	aaaatgacta	ggtgtttcac	240
tgcaaaactg	atggatctgt	ccgtttacta	ttggaattat	cttagctaaa	ggcaggctga	300
cactggaaag	actattcata	gagttacat	gttgcaggtc	ctgttcagta	ggtcgaaaga	360
actcagccat	attgtctaga	agtctactaa	aacctcgggt	taaacaggta	ttcaaaactg	420
tactaaaatc	tgggctttcc	aacatgtctc	tagtttcatt	gagaagttta	atagtggtaa	480
tgtctcgagg	agaangtcca	caggcctgca	ctgctaattg	agttttctca	tctggcatca	540
tataatg						547

<210> 78
<211> 499
<212> DNA
<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(499)

<223> n = A,T,C or G

<400> 78

cctttttttt	tttttttttt	tttnnaaaaa	aaatcttttt	ttatttcaaa	gattgcttct	60
tatattgaag	ctcatattaa	agcaacagta	caatgttcat	aaaatataag	tgtgatgccg	120
taacattttc	ttacatgtca	gaatactgat	atttatatgt	atactaaaat	aagaacttta	180
aaattgtaca	aatagatata	ttaaaaatga	catagaaata	gggcgtctnt	cactgaaaca	240
agacagttat	atctggcacg	tattagttta	agatgaaagt	agaagcaaaa	agatttacia	300
gaatcagcag	taacaagatt	gatgctcaag	agacataatt	gtacattgna	ttgtacatac	360
attgtatggg	tttaagctgg	ctgaatntta	tatatittca	gtttaaaaat	gcactacata	420
tagagtgtcc	agagtttaag	gcgaaattac	agctcanaac	tgntgncctt	tctaattttg	480
gggaagcttn	tttgacaac					499

<210> 79

<211> 370

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(370)

<223> n = A,T,C or G

<400> 79

cctttttttt	tttttttttt	ttttaaggag	caatgacatt	tcctagaagt	tactttaaga	60
atttccctag	agggtcgggt	atcatctcan	ccagatcttt	ctcatccttc	aaggccctgt	120
ttggtacagc	ttgctaggaa	gctgttccag	actgcagcag	ccctctctgg	ggtctctcta	180
ccacttccca	ggcactcana	acttgtgcct	cannanactg	ttttgtggca	ctgncccatt	240
ctctgattct	ccatgtgagc	tggtttttatc	ccatccagca	tggtctgtgaa	atcctaaagg	300
ttcaaaccac	agccactctt	cacctatatt	tcccccaaat	ggctagcacg	ggaaagggcc	360
caaaggtagg						370

<210> 80

<211> 428

<212> DNA

<213> Homo sapien

<400> 80

gtcgacaaaa	agggaaggaa	ggagagacag	ataactctca	gtcattttaa	aaactacaat	60
aaaatattat	gaattatcaa	ttagatcaaa	gttcctcaca	gctatattta	tataggtaaa	120
aaaaaattaa	ataggctaaa	tgcccaaaaa	tttaagactg	gcaaaaata	cttggctaaa	180
tactgtgcgt	ctctattaaa	taccatgttt	cagaagaatt	attaatgaca	tgagaatatg	240
ctcaaaatac	atattgatat	gtgcaaatac	atattgcaaa	gtaagattat	agaatgatcc	300
tagttcaaaa	atgtcacata	tatatgtatt	taaaaaaaaa	ggcagttaag	atttacaaca	360
aaatgttagt	ggtgggacct	tctggtagga	atacagattt	ttttttattc	agaagttttt	420
tgatgtcg						428

<210> 81

<211> 533

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(533)

<223> n = A,T,C or G

<400> 81

cctttttttt	tttttttatt	tttaaaattt	ttttattttg	aaataattat	aaattatcag	60
aaagtgtcaa	acaaagccca	gtcagggtccc	atgtaccagt	ttcactgcca	ccatctttaa	120
aggaggatta	gacgaatctg	actgctaaaa	gtggcccagg	gattctggag	aaaatccaac	180
aggtttgcta	tcaggaaagc	aatttcactt	acaattcagg	tttgactgca	agtgaagtg	240
gttgaaacaa	gtgagaagnt	gattgcttcc	tcatataata	gtctaaatgt	aggtgtccaa	300
gcctggaata	gaggtcctgg	tcctctaagt	tctcaggaac	acaggcttct	tttagccact	360
ccacatctct	aggggtgttg	cctcatggtc	caaaatggng	actggaattc	cagccatcac	420
atntgctttc	caggcagcaa	aatggaagaa	ggggcacana	agaacagaga	tgacaatagg	480
tataaacaag	ctctcttttt	aaaggagatt	cccaggagct	gctacatgac	act	533

<210> 82
 <211> 493
 <212> DNA
 <213> Homo sapien

<400> 82	
gtcgacccgc	gaagatgcag
tgctgtcccc	gctgggggtc
aggagtctct	gggctcggtg
cggcccagca	tgagaacttc
gtcctaaagt	gtacttcatg
ttcacgcagt	ggccaataat
agtttctttc	tgaaacagag
atgaggccat	acaggcagcc
gacaaggtga	att

<210> 83
 <211> 501
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(501)
 <223> n = A,T,C or G

<400> 83	
cctttttttt	tttttttgta
cagaatgcag	aaaacacatc
ggtgaacact	tgaatgtgag
cccagcgctc	aggggagcag
ctgccccatc	gcctccagtt
atggcagtg	ccaatggtgg
agccctcacc	tggaacattg
tcaccaggtc	agatcccact
ggaggcatga	gcccagtcac

<210> 84
 <211> 454
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(454)
 <223> n = A,T,C or G

<400> 84

```

cctttttttt tttttttttt ttttatgcta ataaaacatc ataatttaag gactacactg      60
catttttttaa ttccataaat tataatcctt taacatatat gaaagtttca tattcttaaa      120
gngcttttaa atatatttaa tttttttaac aagtggaaaa gaatgtttct taaaagacat      180
ttaatttttt agtggaattt aatattacca aaaacattct gtgcataaca atttgaataa      240
caattttttt atcttcaaga aatgggattt ttatataaaa tacacatgta gcaactgaatg      300
ccaaagtgat ggggtatccat ggtcanaatt caaaattaga ttcgctatta aacctgtctg      360
gtttgtgtcc tgagtgaana atgatctcga gctggggagg gaggtgcatt gggtaatcag      420
tgcttttgaa ggtgaatttc cttgctgnga aata                                     454

```

```

<210> 85
<211> 509
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(509)
<223> n = A,T,C or G

```

```

<400> 85
gtcgaccgct ctcagctctc ggcgcacggc ccagcttcct tcaaaatgtc tactgttcac      60
gaaatcctgt gcaagctcag cttggagggt gatcactcta ccccccaag tgcataatggg      120
tctgtcaaaag cctatactaa ctttgatgct gagcgggatg ctttgaacat tgaacagacc      180
atcaagacca aaggtgtgga tgaggtcacc attgtcaaca ttttgaccaa ccgcagcaat      240
gcacagagac aggatattgc cttgcgctac cagagaagga ccaaaaagga acttgcataca      300
gcaactgaagt cagccttatc tggccacctg gagacggtga ttttgggcct attgaagaca      360
cctgtcagat atgacgcttc tgagctaaaa gcttccatga aggggctggg aaccgacgag      420
gactctctca ttgagatcat ctgctccaga accaaccagg agctgcagga aattaacaga      480
gtctacaang aaatgtacaa gactgatct                                     509

```

```

<210> 86
<211> 520
<212> DNA
<213> Homo sapien

```

```

<400> 86
gtcgacgggc gccagggtct ttgtggattg catgttgaca ttgaccgtga gattcggcctt      60
caaaccaata ctgcctttgg aatatgacag aatcaatagc ccagagagct tagtcaaaga      120
cgatatcacg gtctacctta accaaggcac tttcttaagc agaaaatatt gttgaggtta      180
cctttgctgc taaagatcca atcttctaac gccacaacag catagcaaatt cctaggataa      240
ttcacctcct catttgacaa atcagagctg taattcactt taacaaatta cgcatttcta      300
tcacgttcac taacagctta tgataagtct gtgtagtctt ccttttctcc agttctgtta      360
cccaatttag attagtaaag cgtacacaac tggaaagact gctgtaataa cacagccttg      420
ttatttttaa gtcctatttt gatattaatt tctgattagt tagtaaataa cacctggatt      480
ctatggagga cctcggctct catocaaagt gcctgagtat                                     520

```

```

<210> 87
<211> 171
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(171)
<223> n = A,T,C or G

```

```

<400> 87
gtcgacgagt acagtatcag ctgagctgac cttactctga ggactaactc ttttgctgga      60

```

35

agcgggtttct gatttacagc tcttggtttc tcccagacat gttggtggga gagattttgg 120
 tttttaaggg gttgtagat ggagtaaann ttctttaagn nttattttt t 171

<210> 88
 <211> 508
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(508)
 <223> n = A,T,C or G

<400> 88
 cctttttttt tttttttttt tttttgnagt aaaaaatctt tatttccaaa atgatttggt 60
 agccaaaaga actataaacc acctaacaag acttttggtta gaaagagact tgatgcttct 120
 tataaattcc ccattgcaaa caaaaaataa caatocaaca agagtcattgt taccattct 180
 tagccattaa cctgggtttta agtctccaaa atcaggattt taaaatgtac ccaactggga 240
 ccaaatacaa acatgagaca ctaggngggc ttgtccttga ttaggaatca ccagcttaag 300
 gaactttatc atgggctgag agttagatag atagcttana acaacattgc aaaagnnggt 360
 gcttctacat gaggactttt ttccccccaa gtagaaaaat aattaaatct tngtcttctt 420
 tatattgngc tttttttggg agaaagcaat tcatttaagg atttaaaaca tgttggtatc 480
 aaaggtagtt canagatgta ataatggt 508

<210> 89
 <211> 508
 <212> DNA
 <213> Homo sapien

<400> 89
 gtcgacggga taaatagaaa gcagaatgaa ttaatggaaa agaactcggc tgttaggcca 60
 ttctctaaat tctagttag ccaaaagttt atgtgtggtt tggggcttca tttatttct 120
 tcatgagtaa aatggaataa tacctaacag gcaggctctg gaagttggaa atcacatata 180
 cacacacaca cacacagaca cacacacaca cgatcaatca ttagctcat attagatgtt 240
 caataaataa cagctactac agatgcctat cagttgagta agtagttcat taaattgagc 300
 tcccaaaggt ctcttctctt cacatccata tccgtttctg cagcaatcaa atagatacat 360
 gattgttttt ctgtaagaaa ttactgcaaa gagaatcttt ttctctact aactgttcct 420
 tctacctggt ataggagata aatgtacgtt tcttaattag ctgacttttt agtatgtcat 480
 ttctgaagga aaaataaatt aaccttaa 508

<210> 90
 <211> 531
 <212> DNA
 <213> Homo sapien

<400> 90
 gtcgacagca gtcccgcgtt ctctccttga atccactcgc cagcccgcgc cctctgcgc 60
 ccgcaccctg cacaccgcgc cctctcctgt gccaggaaact tgctactacc agcaccatgc 120
 cctaccaata tccagcactg accccggagc agaagaagga gctgtctgac atcgctcacc 180
 gcatcgtggc acctggcaag ggcattcctg ctgcagatga gtccactggg agcattgcca 240
 agcggctgca gtccattggc accgagaaca ccgaggagaa ccggcgcttc taccgccagc 300
 tgctgctgac agctgacgac cgcgtgaacc cctgcattgg ggggtgtcat ctcttccatg 360
 agacactcta ccagaaggcg gatgatgggc gtcccttccc ccaagttatc aaatccaagg 420
 gcggtgttgt gggcatcaag gtagacaagg gcgtggtccc cctggcaggg acaaattggcg 480
 agactaccac ccaaggggtg gatgggctgt ctgagcgctg tgcccagtac a 531

<210> 91
 <211> 426

<212> DNA

<213> Homo sapien

<400> 91

gtcgacaatt	gaggcctaca	agagagggga	gcctaggagc	ttggattgac	cttctagtca	60
accacctgac	ttcagcacac	cattacaatc	gggagactaa	accaacaacc	agaggatcta	120
aaatgtcaca	ttcagatttt	caggaagaaa	atcttcatta	cagtggagca	caaagtgtcc	180
atacaagaca	tcattgagga	gccatgctgt	ccccttctaa	cctgaaacac	attctttccc	240
atcctggttg	ggcttctgta	cctccttatt	aatttatgaa	cctgaagttg	cttgaagtgt	300
tttgggctta	ataaatgggg	tgaaagtata	ggtagcagta	acacctacat	gaaacaatac	360
accttggtac	ttttaatcta	aattactttt	cttttttaag	totactttta	aaataaatac	420
ttctgt						426

<210> 92

<211> 223

<212> DNA

<213> Homo sapien

<400> 92

gtcgactttt	aaagcaattg	actaggagaa	actatttgta	gcttatataa	caaggactat	60
atataataaa	aaaactattt	ctatgaaaat	cttaaaatta	cacacagtcc	gatgaaata	120
atcatatatt	aaaaaggcaa	accagaaaaa	taaatacaga	tgaccaaaat	ccatgtgaca	180
tatttggoct	aattagtaat	tagaaaaaaa	aaaaaaaaaa	aaa		223

<210> 93

<211> 486

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(486)

<223> n = A,T,C or G

<400> 93

cctttttttt	tttttttttt	tttttttttt	tctcaaatat	ccaattttat	tttatcattc	60
tgcattggg	ggatgcgatc	tgagctagg	atcggaattc	ccaggcctat	anatttttaa	120
accacaccac	aggggtaaac	cttaaaagaa	gngaaacct	acactatata	tatttccatt	180
tctaaatata	gtatattaca	naagttttaa	tatnccacct	ntgngtactt	acaactntaa	240
aaagatncaa	tanctctacc	aattataaat	aatgtancat	ttcatattaa	agacattatc	300
gtncaatgga	anaataggaa	ccctntaacg	tatcactatc	aagggttagng	tctatatcta	360
cttganataa	aatactgaaa	attcagngta	tgaagccaaa	tcctgattta	acaagttatt	420
ggtagtataa	gtgataagtg	ttanctgatg	aagggaaggc	aaatgtggta	atttatatct	480
ctgaca						486

<210> 94

<211> 214

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(214)

<223> n = A,T,C or G

<400> 94

cctttttttt	tttttttttt	tttttttttt	tttttngcaa	cacaagtcaa	tctttattga	60
aaactgcagt	attaatacat	aacaattctt	gttacaataa	acgtgctttt	ganattttta	120

```

aatctgagct catctcatca gattgcataa aaaattaaaa tagtntcaat tgacacctaa    180
ctgaactggc tcaggatgga aattccattc cttg                                214

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<210> 95
<211> 463
<212> DNA
<213> Homo sapien

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```

<400> 95
gtcgaccaga attcagagcg aatggtcaca gttggtcgct gggcaaaggg aatgagtgcg    60
gactatgaag aaatttttgga tgtacctaaa cgcacaaaac ccaaaacaaa aatacctaaa    120
gttgtaatt tttgataaca gctagcacta tcatgagtta ctacctcatt gttactttct    180
aaaccaggcc cgcttcacga gttagagttg agctcccctg tagccaggac tatgctgtag    240
atatcagtat gatctgggtg tggccaaaaa caattttctt tattctgtct atcaaatagt    300
acttctacca ctggttgag aaaattgaag aaaagaataa gatgattaaa tgaattctct    360
aaaagaacat attttaagag acagaactta gacataacca agtagttgta tacctgattg    420
taacaatcat cttttataaa agcaaaatta tgcataaatg taa                    463

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```

<210> 96
<211> 606
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(606)
<223> n = A,T,C or G

```

```

<400> 96
gtcgacttta aaagtgcctc ggcattcctgt attacatgtc atagaattgt aaagtcaaca    60
tcaattacta gtaatcattc tgcactcact ggggtgcatag catgggttaga ggggctagag    120
atggacagtc atcaactggc ggatatagcg gtacatatga tccttagcca ccagggcaca    180
agcttaccag tagacaatac agacagagct tttgttgagc tgtaactgag ctatggaata    240
gcttctttga tgtacctctt tgccttaaat tgctttttag ttctaagatt gtagaatgat    300
cctttcaaat tgtaatcttt tctaacagag atattttaat atacttgctt tcttaaaaaa    360
caaaaaaact actgtcagta ttaatactga gccagactgg catctacaga ttccagatct    420
atcattttat tgattcttaa gcttgtatta aaaactaggc aatatcatca tggatacata    480
ggagaagaca cattacaat cattcattgg gccttttatc tgtctatcca tccatcatca    540
tttgaggcct aatatatgcc aagtactcac atggtatgca ttgngacata aaaaagactg    600
tctata                                606

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```

<210> 97
<211> 530
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(530)
<223> n = A,T,C or G

```

```

<400> 97
cctttttttt tttttttgta gattttttgc tatgttactc aggcgtgtct tggactcctg    60
ggctcaagcg atcctccac cttggcttcc caaagtgccg ggattatagg catgagccac    120
catgctcggc ctgctccttt tcttgaaaca cctcctctgt ggttagatt ccaggagact    180
ggaatggtct gccctggtgg gctgctgagt cagggacctg aggtgtttgt tcaactggga    240
ggcgggttca gatcaggaat gtaaggatga tggaaagaag ggagtcactc tggtttggtg    300
ggactgggga gcaatcttga tcacggccac ttacagcttc tgccattgtc cttcaccact    360

```

atctcagcat	ctcgggtccct	cacgatgtcc	ctccagtcaa	ttgtgtccat	gtgacaaagc	420
ttatcgttct	tctcaatata	aacaccccct	gacagaatct	cggtagagctg	agtcaagcgg	480
agctggcgca	nagcgtggct	ggagttgggtg	ttatagttca	acatgacgaa		530

<210> 98
 <211> 479
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(479)
 <223> n = A,T,C or G

<400> 98	
gtcgacgggtt	agtttctgcg
atgctaaaaat	tgggcaccct
agtttaaaga	tatcagcctg
ctcttgactt	cacctttgtg
aatttaagaa	actcaactgc
tagcatgggt	caatacacct
tatcagaccc	gaagcgcacc
tctcgttcag	ggggcctttt
actttgtgtg	ggactgctga
gcccccaact	tcaaagccac
tctgactaca	aaggaaaata
tgccccacgg	agatcattgc
caagtgtattg	gtgcttctgt
aagaaacaag	gaggactggg
attgctcang	attatgggggt
tatcattgat	gataagggta
	ttcttcggca
	gatcactgt
	60
	120
	180
	240
	300
	360
	420
	479

<210> 99
 <211> 502
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(502)
 <223> n = A,T,C or G

<400> 99	
cctttttttt	tttttttgta
agacaatcaa	ctctatgagg
acagtaggta	ctcaataaat
ggatggaagg	atgaatggag
tcttattatg	ggtggaggaa
gagcatcatt	tttgggtgtc
tttacctctt	tcaatgcccc
actttcccaa	catttgcttt
tgccatggca	tcattaaaag
agtttaaatt	tattttttta
gcagagacta	tgaccact
aggtggatg	gaggtaatgg
gacctgctg	aagtgtgagt
gattgagaaa	ataagataaa
gactaataaa	tctagagaat
ttgaaaccga	aaaagtaaat
taaaaatgca	atctctangt
	60
	120
	180
	240
	300
	360
	420
	480
	502

<210> 100
 <211> 537
 <212> DNA
 <213> Homo sapien

<400> 100	
gtcgaccctt	tccataaatc
caagagtttt	gggagttgta
gccaaggacc	tgagacctga
tgtccccctt	tagatctctg
tttttctctt	ccacacagtg
aaggaaaaga	cacatttttt
ttaaagaccc	ctaacccttt
cttgatgatt	gacaacaccc
gttaatcatc	aagagaat
aggggttgact	ttaccattt
aagccacaaa	taggatgctt
ctcaaggcca	gcttatagtc
aggaaatgtt	tttaataaaa
gtgtgctctc	cattctgctc
	atttttcctt
	ggggcttcca
	gggtgggagt
	gggaagactc
	atatatatca
	gaaaattaca
	cttccccatc
	60
	120
	180
	240
	300
	360
	420

tttctgaggt	gcactgggag	gtcccccctt	tatttggggc	ttgatgactt	ttctttttgt	480
agctggggct	ttgatgttcc	tttccagtgt	cattttctcat	ccacataccc	tgacctg	537

<210> 101
 <211> 611
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(611)
 <223> n = A,T,C or G

<400> 101						
gtcgacctaa	aatgaagtgt	ttgaaatcag	aaatctat	ctaagtctc	atagctttaa	60
aactattttt	gtccttatac	tcatacttgt	tattttat	tattcatcct	atatagccat	120
ttgactgaaa	tgtagaaaat	aatttattaa	attgagaaaa	tatgcaggca	ttgaacaatc	180
tttcaagtat	tttgaataaa	aattcaaatt	attatagatt	gcctggaatt	gttaagactg	240
tcagaaggtc	agctcattga	tagctaagta	gtatacactc	tgaaaaacag	aatgtagaaa	300
tgggttttat	aaaagctgac	ctctagagta	aaggaggacc	cagcatgtgt	aattcttcct	360
cttaatactt	taagaccact	aatttgagga	cttatggttt	ctcaccactg	cactcttgca	420
gctttcaaga	aagtacttaa	gttttaaatt	cccagggtgat	ttctaagact	cttgaataga	480
attggttggg	ttcttctgat	attgcatttt	catgagaaaa	aatttcagtg	gtacattaat	540
ttttattttt	ccttttgctt	atagacttcg	catatcattt	aaagtgatgg	ttogagcttn	600
ctctggatac	t					611

<210> 102
 <211> 498
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(498)
 <223> n = A,T,C or G

<400> 102						
cctttttttt	ttttttttta	acgcataatt	gtttttat	ataggtaact	accacatgaa	60
ttataaagac	aacaaaggat	gtcagaatga	acatggatag	gtgtatgcat	actacggcta	120
aggagaaaca	atgttcctac	atattatggg	tagtgagaac	attatctgta	taacagggaa	180
ctgtgattat	ttaaaaaatat	gcagaactta	tttcatctgt	gctttanaaa	taactgtata	240
cagtgttata	agttgaaaag	aactcaaaat	aactaatacc	aatatacac	ctatgtatta	300
naattcaaaa	aagctgcttt	ctgtgaagtc	aatcagctat	attaaaaaat	gacacaaatc	360
caaaacaaga	tgcattgttat	atataaaggg	acattgtaag	tttccttgct	gcattaaacc	420
catggtttta	tccatgaaat	ttccttttaa	ttatcattta	gacagaagca	tgcaaatagt	480
ctcaggatct	acttaaga					498

<210> 103
 <211> 446
 <212> DNA
 <213> Homo sapien

<400> 103						
gtcgactctt	ggtgtttttg	tattttccacc	tcacccccag	cacatagccc	agtctcttgc	60
acaaattaag	tacttaatgt	gtgttgagct	aaattgaata	aaggattatt	agcattagca	120
tattttgtgc	cttggttgta	taagctgggt	gtttgttttg	ttaccttgc	aaatattttat	180
gattatcacc	ccccacata	ctaaattgtt	tttaaaagtt	ttgcctttcc	ttcagatact	240
accccaggca	atttgctgta	gataatgtga	ttgcttccaa	tgacataatt	atcccaaact	300

ctctgccccg gatatacttt gccaaacgaa atttgaattc tctgaataaa ttggtcatgt	360
cctaaaaaaa aaaaaaaaaa aaaaaaaggg gcggccgctc gagtctagag ggccccgttt	420
taaaccccgc tgatcagcct cgactg	446

<210> 104
 <211> 286
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(286)
 <223> n = A,T,C or G

<400> 104	
gtcgaccttc gttatccgcg atgcgtntcc tggcagctac attcctgctc ctggcgctca	60
gcaccgctgc ccaggccgaa ccggtgcagt tcaaggactg cggttctgtg gatggagtta	120
taaaggaagt gaatgtgagc ccatgcccca cccaacctg ccagctgagc aaaggacagt	180
cttacagcgt caatgtcacc ttcaccagca atattcagtc taaaagcagc aaggccgtgg	240
tgcatggcat cctgatgggc gtcccanttc ctttcccat tctga	286

<210> 105
 <211> 406
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(406)
 <223> n = A,T,C or G

<400> 105	
gtcgacgcgt agcagagtgg tcgttgtctt tctaggtctc agccggctgt cgcgacgttc	60
gcccgtctgc tctgaggctc ctgaagccga aaccagctag actttcctcc ttcccgcctg	120
cctgtagcgg cgttggttgc actccgccac catgttcgag gcgcgcctgg tccagggctc	180
catcctcaag aagggtgttg aggcactcaa ggacctcatc aacgaggcct gctgggatat	240
tagctccagc ggtgtaaacc tgcagagcat ggactcgtcc cagctctctt tgggtcagct	300
cacctgcggg tctgagggtc tcgacacctc ccgctgcgac cgcaacctgg ccatgggcgt	360
gaacctcacc agtatgtnc aataactaaa atgcgccggc aatgaa	406

<210> 106
 <211> 258
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(258)
 <223> n = A,T,C or G

<400> 106	
gtcgacgatt tttttgtac attttggctg cagtattggt ggtagaatat actataatat	60
ggatcatctc tacttctgta tttatttatt tattactaga cctcaaccac agtcttcttt	120
ttccccttcc acctctcttt gcctgtagga tgtactgtat gtagtcatgc actttgtatt	180
aatatattan aaatctacag atctgttttg nactttttat actgttggat acttataatc	240
aaaactttta ctagggtg	258

<210> 107

<211> 369
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(369)
 <223> n = A,T,C or G

<400> 107
 gtcgacgtaa aatagaaaca gaaggggact ttatcaacct gattaacttt ctcaacatgt 60
 taaccctaca gttaacatta taatcaatgg tgaatcattg agtactttcc ttctaagatc 120
 agaaacagtt caaagtccac tctcaccatt tctattcaac attgtactgg aatcccagcc 180
 agtgcagtaa taccaataat aaaaaattaa agtcataaag attgaaaagg atgaagtaaa 240
 gctattttcaa ttntatttag aagtatttag aaaccccaaa gaatctacaa aaaactaata 300
 gaaataagtg aatatatgaa ggtcttacta tacaagatca acatatcaaa agcagtggtg 360
 tttaagaaa 369

<210> 108
 <211> 289
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(289)
 <223> n = A,T,C or G

<400> 108
 gtcgacattg catccttgaa atcctgggct cagggtgatcc tcccgctga gcctcctgag 60
 tatctgggac tacagatgcg tgccaccaag cctggctaatt tttgtctcat gtctttctaaa 120
 aattattttg tgaagcccct tcacaaaaaa ccttaaggga aatctgatgg tgctcaggaa 180
 tctaactctc cctaaaccat cctctttaac tgctttctaaa atatctctgt tggcctttct 240
 tanccttttt ctgtttccat tcagtgetcc aagcgctttt tgtttctaa 289

<210> 109
 <211> 444
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(444)
 <223> n = A,T,C or G

<400> 109
 gtcgacctgg cgttggcacc gctgaggaat gggcctgggc ggggaggagac atctctacac 60
 cgttcccacg cggaacagg gcaacatcta caagcccaac aacaaggcca tggcagacga 120
 gctgagcgag aagcaagtgt acgacgcgca caccaaggag atcgacctgg tcaaccgcga 180
 ccctaaacac ctcaacgatg acgtggtcaa gattgacttt gaagatgtga ttgcagaacc 240
 agaagggaca cacagttttg acggcatttg gaaggccagc ttaccacact tcaactgtgac 300
 naaatactgg ttttacogct tgctgtctgc cctctttggc atcccgatgg cactcatctg 360
 gggcatttaa cttcgccatt ctctctttcc tgcacatntg ggcagttgta accatgcatt 420
 aagagcttcc tgattgagat tcag 444

<210> 110
 <211> 196
 <212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(196)

<223> n = A,T,C or G

<400> 110

cctttttttt	ttttttcatt	aaataancca	tcacacacatt	agtacaatac	aattttatat	60
tttttaaata	tactatata	gttaaggata	aggggtgaag	ttttcttcct	ttgtaatacc	120
tggtcaagag	tttaattgat	taggagatta	gngttaacct	tgaggataaa	agtncaaatt	180
tgtctcatta	ggacac					196

<210> 111

<211> 544

<212> DNA

<213> Homo sapien

<400> 111

gtcgacctca	gceggctcgc	gcgacgttcg	cccgtctcgt	ctgaggctcc	tgaagccgaa	60
accagctaga	ctttctcct	tcccgcctgc	ctgtagcggc	gttggttcca	ctccgccacc	120
atgttcgagg	cgcgcctgg	ccagggtctc	atcctcaaga	aggtgttgga	ggcactcaag	180
gacctcatca	acgaggcctg	ctgggatatt	agctccagcg	gtgtaaacct	gcagagcatg	240
gactcgtccc	acgtctcttt	ggtgcagctc	accctgcggg	ctgagggtct	cgacacctac	300
cgctgcgacc	gcaacctggc	catgggcgtg	aacctcacca	gtatgtccaa	aataactaaaa	360
tgcgccggca	atgaagatat	cattacacta	agggccgaag	ataacgcgga	taccttggcg	420
ctagtatttg	aagcaccaaa	ccaggagaaa	gtttcagact	atgaaatgaa	gttgatggat	480
ttagatgttg	aacaacttgg	aattccagaa	caggagtact	gctgtgtagt	aaagatgcct	540
tctg						544

<210> 112

<211> 378

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(378)

<223> n = A,T,C or G

<400> 112

gtcgacacgg	cttccgcacg	gtcatccgcc	ccttctacct	gaccaactcc	tcagggtgtg	60
actagacggc	gtggcccaag	ggtggtgaga	accggagaa	cccaggacgc	cctcactgca	120
ggctccccct	ctcggttcc	ttctctctg	caatgacctt	caacaaccgg	ccaccagatg	180
tcgccctact	cacctgagcg	ctcagcttca	agaaattact	ggaaggcttc	cactagggtc	240
caccaggagt	tctcccacca	cctcaccagt	ttccagggtg	taagcaccag	gacgccctcg	300
aggttgctct	gggatcccc	cacagccct	ggncagtctg	cccttgnac	tggtctgaag	360
gtcattaaaa	ttacattg					378

<210> 113

<211> 530

<212> DNA

<213> Homo sapien

<400> 113

gtcgacgtcg	ttgtctttct	aggtctcagc	cggtcgtcgc	gacgttcgcc	cgtcgtctct	60
gaggctcctg	aagccgaaac	cagctagact	ttctctcttc	ccgcctgcct	gtagcggcgt	120
tggtgccact	ccgccaccat	gttcgaggcg	cgcttggtcc	agggctccat	cctcaagaag	180

gtgttgagg	cactcaagga	cctcatcaac	gaggcctgct	gggatattag	ctccagcgg	240
gtaaacctgc	agagcatgga	ctogtcccac	gtctctttgg	tgagctcac	cctgcggtct	300
gagggcttcg	acacctaccg	ctgagaccgc	aacctggcca	tgggcgtgaa	cctcaccagt	360
atgtccaaaa	tactaaaatg	cgccggcaat	gaagatatca	ttacactaag	ggccgaagat	420
aacgcggata	ccttggcgct	agtatttgaa	gcaccaaacc	aggagaaagt	ttcagactat	480
gaaatgaagt	tgatggattt	agatgttgaa	caacttgga	ttccagaaca		530

<210> 114

<211> 178

<212> DNA

<213> Homo sapien

<400> 114

gtcgacattt	cttcctaata	ttctataatc	tccaactcct	gaaaacccct	ctctcaacta	60
atactttgct	gttgaaatgt	tgtgaaatgt	taagtgtctg	gaaatttttt	ttttctaaga	120
aaaactatta	aagtacttcc	tagtagggca	aaaaaaaaaa	aaaaaaaaaa	aaaaaaaaaa	178

<210> 115

<211> 211

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(211)

<223> n = A,T,C or G

<400> 115

cctttttttt	ttttttttng	gntcaatctt	ttatttgga	caaaggaaaa	aaggactgac	60
accagtttag	cotttgagtg	tgcaaagctc	tgccctccct	cccacccctn	agccccaat	120
ccaanatttc	atagccctaa	caccaccca	agcagnttcc	ctcacacatg	ccctttgntt	180
tcttcctctc	ttctatgggt	ccttaggnaa	a			211

<210> 116

<211> 439

<212> DNA

<213> Homo sapien

<400> 116

gtcgacctgt	cactcactac	atgaataagc	aaatattgtc	ttcaaaagaa	tgacaagaa	60
ccacaattaa	gatgtcatat	tattttgaaa	gtacaaaata	tactaaaaga	gtgtgtgtgt	120
attcacgcag	ttactcgctt	ccatttttat	gacctttcaa	ctataggtaa	taactcttag	180
agaaattaat	ttaatattag	aattttctatt	atgaatcatg	tgaaagcatg	acattcggtc	240
acaatagcac	tatttttaa	aaattataag	ctttaaggta	cgaagtattt	aatagatcta	300
atcaaatatg	ttgattcatg	gctataataa	agcaggagca	attataaaat	cttcaatcaa	360
ttgaactttt	acaaaacca	cttgagaatt	tcatgagcac	tttaaaatct	gaactttcaa	420
agcttgctat	taaatcatt					439

<210> 117

<211> 357

<212> DNA

<213> Homo sapien

<400> 117

gtcgactoca	aattgacttt	gcagcaggg	ggcaggggtca	ggagagtctg	gtcctgccta	60
gtcagattt	catggcacct	gcacttgaag	caagtcaact	ctttatcaca	gggtgtctga	120
aacattagct	tcttttacca	acctgagaaa	attaggatga	cctggcaa	aagatcttga	180
ataggccaaa	agcaagtatc	ttgctgtgtg	tagtctcttg	gttaaagtga	agaaacagta	240

ctgttcacac	ctttcttcac	tgagattcca	gtgtacatga	gaacatatat	ttattgcatg	300
attttctaga	tacacagtct	atgcattatt	catatacatt	tatttttagcc	taaagtg	357

<210> 118
 <211> 431
 <212> DNA
 <213> Homo sapien

<400> 118						
cctcccctgag	gaaattagga	acctgttggc	agatgttgaa	acatttgtag	cagatatatact	60
gaaaggagaaa	aatttatcca	agaaagcaaa	ggaaaagaga	gaatccctta	ttaagaagat	120
aaaagatgta	aagtctatct	atcttcagga	atttcaagac	aaaggtgatg	cagaagatgg	180
ggaagaatat	gatgaccctt	ttgctggggc	tccagacact	atttcattag	cctcagaacg	240
atatgataaaa	gacgatgaag	ccccctctga	tggagcccag	tttcctccaa	ttgcagcaca	300
agaccttctt	tttgttctaa	aggctgggta	ccttgaaaaa	cgcagaaaag	atcacagctt	360
tctgggattt	gaatggcaga	aaacgggtgg	gtgctctcag	taaaacggta	ttctattatt	420
atggaagtga	t					431

<210> 119
 <211> 131
 <212> DNA
 <213> Homo sapien

<400> 119						
cccctcgccc	gtcacgcacc	gcacgttcgt	ggggaacctg	gcgctaaacc	attcgtagac	60
gacctgcttc	tgggtcgggg	tttcgtacgt	agcagagcag	ctccctcgct	gcgatctatt	120
gaaaggtcga	c					131

<210> 120
 <211> 409
 <212> DNA
 <213> Homo sapien

<400> 120						
gtcgacgtaa	aagccacaca	gaaatcaaaa	gataagaata	tagtttcagc	taccaaaaag	60
cagcctcaga	ataaaaagtc	atttcagaag	acaggaccca	gctccttgaa	gtctcctggc	120
cgtaacccac	tgtccatcgt	gagcctaccc	cagtcttcta	ccaaaacaca	aactgcaccg	180
aagtcagcac	agactgtcgc	taagagccag	cattcaacta	aagggcctcc	cagaagtggc	240
aaaaccccag	cttcaatcag	gaaaccaccc	tcatctgtta	aggatgcaga	tagtggagat	300
aaaaaaccta	ctgcaaagaa	aaaggaagat	gatgaccatt	attttgtcat	gactggaagt	360
aagaaaccta	gaaaataaat	acatactcat	tataaaaaaa	aaaaaaaag		409

<210> 121
 <211> 131
 <212> DNA
 <213> Homo sapien

<400> 121						
cccctcgccc	gtcacgcacc	gcacgttcgt	ggggaacctg	gcgctaaacc	attcgtagac	60
gacctgcttc	tgggtcgggg	tttcgtacgt	agcagagcag	ctccctcgct	gcgatctatt	120
gaaaggtcga	c					131

<210> 122
 <211> 130
 <212> DNA
 <213> Homo sapien

<400> 122

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gtcgaccttt caatagatcg cagcgagggg gctgctctgc tacgtacgaa accccgaccc    60
agaagcaggt cgtctacgaa tggtttagcg ccagggtccc cacgaacgtg cggtgcggtga    120
cgggcgaggg                                     130

```

```

<210> 123
<211> 424
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(424)
<223> n = A,T,C or G

```

```

<400> 123
gtcgacgaga tgtggagtgg ctaaaagaag cctgtgttcc tgagaactta gaggaccagg    60
acctctattc caggcttggg cacctacatt tagactatta tatgaggaag caatcaactt    120
ctcacttggt tcaaccactt tcacttgcag tcaaacctga attgtaagtg aaattgcttt    180
cctgatagca aacctgttgg attttctcca gaatccctgg gccactttta gcagtcagat    240
tcgtctaata ctcttttaaa gatggtggca gtgaaactgg tacatgggac ctgactgggc    300
tttgtttgca actttctgat aatttataat tatttcaaaa taaaaaaatt ttaaaaataa    360
aaaaaaaaaa aaagggcggc cgctcgaggt ctagaggggc cgtttaaacc cgntgatcag    420
cctc                                     424

```

```

<210> 124
<211> 548
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(548)
<223> n = A,T,C or G

```

```

<400> 124
cctttttttt tttttttctc tagtaatgac tttattcatg aatctataat ggaattcaaa    60
atagcaaaga acatgaaaat gttcanatta atatttatta accaaatgca tcanaaaata    120
catctatttt cacatatcaa aagtgcctaa aatgcatgtg anaatataaa tattctccac    180
tttgnggaac ttcaagataa tgaaaaattg cttaatacac ttgcccacaa aaactcatta    240
cactgcaaat ncagaanaaa taaaataact cattacattg cagatncaa agaaatcaaa    300
tgtaactggc aaaataacca tttcatggct aatctttngg naaagngcta ttttcacact    360
gaaaaaaaga anttagaaaa gattaaaaat tttaaattct gaaccatcat tctnaaagtc    420
tgaagcggtt tcttttagtat tcactatggt catcacattc atgtgtncac aacatgagac    480
taaacactat ctcaaaatct taaaaaatct ttccatncac anattatttc ctggaagnta    540
aaaattat                                     548

```

```

<210> 125
<211> 562
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(562)
<223> n = A,T,C or G

```

```

<400> 125
gtcgacgctc ctaacaaaga agatatcttg aaaatttcag aggatgagcg catggagctc    60

```

agtaagagct	ttcgagtata	ctgtattatc	cttgtaaaac	ccaaagatgt	gagtctttgg	120
gctgcagtaa	aggagacttg	gaccaaacac	tgtgacaaag	cagagttctt	cagttctgaa	180
aatgttaaag	tgtttgagtc	aattaatatg	gacacaaatg	acatgtgggt	aatgatgaga	240
aaagccttaca	aatacgctt	tgataagtat	agagaccaat	acaactgggt	cttccttgca	300
cgcccacta	cgtttgctat	cattgaaaac	ctaaagtatt	ttttgttaaa	aaaggatcca	360
tcacagcctt	tctatctagg	ccacactata	aaatctggag	accttgaata	tgtgggtatg	420
gaaggaggaa	ttgtcttaag	tgtagaatca	atgaaaagac	ttaacagcct	tctcaatatc	480
ccagaaaagt	gtcctgaaca	gggagggatg	atttggaaga	tatctgaaga	taaacagcta	540
cgagnttgcc	tgaaatatgc	tg				562

<210> 126

<211> 131

<212> DNA

<213> Homo sapien

<400> 126

cccctcgccc	gtcacgcacc	gcacgttcgt	ggggaacctg	gcgctaaacc	attcgtagac	60
gacctgcttc	tgggtcgggg	tttcgtacgt	agcagagcag	ctccctcgct	gcgatctatt	120
gaaaggctga	c					131

<210> 127

<211> 512

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(512)

<223> n = A,T,C or G

<400> 127

gtcgacgtcc	ggcttcggag	cgaggagtgt	cgttgtgcc	gcgactaaaa	agagaattaa	60
atatgggtga	tggtgagaaa	ggcaagaaga	tttttattat	gaagtgttcc	cagtgccaca	120
ccgttgaaaa	gggaggcaag	cacaagactg	ggccaaatct	ccatgggtctc	tttgggcgga	180
agacaggtca	ggcccctgga	tactcttaca	cagccgccaa	taagaacaaa	ggcatcatct	240
ggggagagga	tacactgatg	gagtatttgg	agaatcccaa	gaagtacatc	cctggaacaa	300
aaatgatctt	tgtcggcatt	aagaagaagg	aagaaagggc	agacttaata	gcttatctca	360
aaaaagctac	taatgagtaa	taattggcca	ctgccttatt	tattacaaaa	cagaaatgtc	420
tcatgacttt	tttatgtgta	ccatccttta	atagatctca	tacaccagan	tttcagatca	480
tgaatgactg	acagaatatt	ttgttgggca	gt			512

<210> 128

<211> 483

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(483)

<223> n = A,T,C or G

<400> 128

gtcgacgttt	ttgtgatact	gacacatccc	ccctttcaga	acaccctctg	cccttggatt	60
ctgtgcacag	gaagctagtt	gtcccccctga	atacactctt	tcttccttgt	aatacagcct	120
ctgatttttga	gcccaagaat	aaagactaca	gttctcagac	tccttcgcaa	ataaattttg	180
tgactaaact	ctagtcaaca	gtaagtgtca	tgtagcagct	cctgggaatc	tcctttaaaa	240
agagagcttg	tttataccta	ttgtcatctc	tgttcttctg	tgcccccttc	tccatttttg	300
tgccctggaaa	gcagatgtga	tggttggaat	tccagtcacc	attttggacc	atgaggacaa	360

caccctanag atgtggagtg gctaaaagaa gcctgtgttc ctgagaactt anaggaccan	420
gacctctatt ccaggcttgn acacctanat ttanactatt atatgaggaa gcaatcaact	480
tct	483

<210> 129
 <211> 326
 <212> DNA
 <213> Homo sapien

<400> 129	
gtcgaccttt tatctgtcta tccatccatc atcatttgaa ggcctaatat atgccaaagta	60
ctcacatggg atgcattgag acataaaaaa gactgtctat aacctcaata agtattaaaa	120
atcccattat tacccataag gttcatctta tttcattttt agggaataaa attacatgtc	180
tatgaaatth caattttaag cactattgtt tttcatgacc ataatttatt tttaaaaata	240
aattaaagggt taatttatatg catgtatgta tttctaataa ttaaaaatgt gttcaatccc	300
tgaaaaaaa aaaaaaaaaa aaaaaa	326

<210> 130
 <211> 276
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(276)
 <223> n = A,T,C or G

<400> 130	
gtcgacggac accagctgcg gaanttgcgg ctttggcaga ttgaaatcat ggcagggtcca	60
gaaagtgatg cgcaatacca gttcactggg attaaaaaat atttcaactc ttatactctc	120
acaggtagaa tgaactgtgt actggccaca tatggaagca ttgcattgat tgtcttatat	180
ttcaagttaa ggtccaaaaa aactccagct gtgaaagcaa cataaatgga ttttaactg	240
tctacggttc ttaacctcat ctgttaagtt cccatg	276

<210> 131
 <211> 482
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(482)
 <223> n = A,T,C or G

<400> 131	
cctttttttt ttttttttaa attttaaggt tatttttatt tacaactttt gaaaaatgta	60
catttttttt tacatgggtt acttgtgcaa agtttagattt ggaagtgata aatgcataaa	120
aggngacaat agaacattan acaaaacatt tacaagcctt gtcccatact gctacttaaa	180
ggtactatat atctaaaagt ataaatatcc aaaaaaagat cgcanacatt ggctttaagg	240
ttctcanatg ctgaaagggg aanaattaaa gcatgcagca ataactcagg atttgagtgg	300
aaaatagttt gccacanata tgctatgctc ccttccttga attcattaaa actctaaaat	360
aaagatggac aattgagttt attcacttag ggcagcactg atcctttaaa aagattaaag	420
gagctccaac tttccctagc tnaaaaactc acnatngttt ccattcctct gctcccacac	480
ct	482

<210> 132
 <211> 428
 <212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(428)

<223> n = A,T,C or G

<400> 132

cctttttttt	tttttttgtc	taaaaggcaa	aaaactacaa	acagcccaag	tcctgagctc	60
cccaagacct	ggatcctcca	ctgtccccct	gaaaccggc	aggaggcggg	atggggagca	120
caanagggtg	gttcttaaaa	aagtcacccc	tggatgggaa	agctcttcat	cttctgccgc	180
cttctnttgc	ctcccgtgc	tgccgaggag	agagatggan	aggaccgggg	ctatgccggc	240
aaactcaact	tcttccccct	taggactttg	gngatataga	ggtanaanaa	atcgcagtan	300
aggactgtct	ggaccaggcc	tgccacaatg	gcnatgaggt	cgaagaancc	ctcgaaangg	360
taagcgccan	anccagttga	anagatanag	cgtggcggtg	aacgcctagc	gcaaacaagt	420
agnnggctg						428

<210> 133

<211> 537

<212> DNA

<213> Homo sapien

<400> 133

gtcgacccca	aaccactcc	accttactac	cagacaacct	tagccaaacc	atttacccaa	60
ataaagtata	ggcgatagaa	attgaaacct	ggcgcaatag	atatagtacc	gcaagggaaa	120
gatgaaaaat	tataaccaag	cataatatag	caaggactaa	cccctatacc	ttctgcataa	180
tgaattaact	agaaataact	ttgcaaggag	agccaaagct	aagacccccg	aaaccagacg	240
agctacctaa	gaacagctaa	aagagcacac	ccgtctatgt	agcaaaatag	tgggaagatt	300
tatagggtaga	ggcgacaaaac	ctaccgagcc	tggtagatgc	tgggtgtcca	agatagaatc	360
ttagtccaac	tttaaatttg	cccacagaa	cctctaaatc	cccttgtaaa	tttaactggt	420
agtccaaaga	ggaacagctc	tttggacact	aggaaaaaac	cttgtagaga	gagtaaaaaa	480
tttaacaccc	atagtaggcc	taaaagcagc	caccaattaa	gaaagcgttc	aagctca	537

<210> 134

<211> 535

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(535)

<223> n = A,T,C or G

<400> 134

gtcgactcct	ctcacatggt	ggcttttagga	agatccttgg	ccaggagggt	gatgccagct	60
atcttgcttc	tgaaatatct	acctgggatg	gagtgatagt	aacaccttca	gaaaaggctt	120
atgagaagcc	accagagaag	aaggaaggag	aggaagaaga	ggagaataca	gaagaaccac	180
ctcaaggaga	ggaagaagaa	agcatggaaa	ctcaggagtg	acattccctt	cactcctttt	240
cctacccaag	ggggaagact	ggagcctaag	ctgcctgcta	ctgggcttta	catggtgaca	300
gacatttccg	tgggataggg	aagatagcag	gaagaaaagt	aaactccata	gaagtgtcat	360
tccactgggt	tttgatattg	gcttagctgc	cagtctccca	tttgtgacct	atgccatcca	420
tctataatgg	aggataccaa	catttcttcc	taatattcta	taatctccaa	ctcctgaaaa	480
acccctctct	caactaatac	tttgctgttg	aaatgttngn	aaatgttaag	tgtct	535

<210> 135

<211> 114

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(114)

<223> n = A,T,C or G

<400> 135

gtcgacctca	gcgtcattca	gaannnggaa	aagaatcaat	gtaactcaag	aaaggatgaa	60
aatacccttt	cttcccatcc	acgtgtttcc	atctcaatcc	tcacagggtc	ctgg	114

<210> 136

<211> 354

<212> DNA

<213> Homo sapien

<400> 136

agaagcgaga	tgacgaaggg	aacgtcatcg	tttgaaagc	gtcgcaataa	gacgcacacg	60
ttgtgccgcc	gctgtggctc	taaggcctac	caccttcaga	agtcgacctg	tggcaaagt	120
ggctaccctg	ccaagcgcaa	gagaaagtat	aactggagtg	ccaaggctaa	aagacgaaat	180
accaccggaa	ctggtcgaat	gaggcaccta	aaaattgtat	accgcagatt	caggcatgga	240
ttccgtgaag	gaacaacacc	taaacccaag	agggcagctg	ttgcagcatc	cagttcatct	300
taagaatgtc	aacgattagt	catgcaataa	atgttctggt	tttaaaaaat	aaaa	354

<210> 137

<211> 347

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(347)

<223> n = A,T,C or G

<400> 137

gtcgacggcg	agattacgag	gcgaggctcg	cgcgcccgcc	cccgccttg	ccccagtc	60
ccacccggtc	ggcccggcac	agccatgac	aaggcgatcc	taatcttcaa	caaccacggg	120
aagccgcggc	tctccaagtt	ctaccagccc	tacagtgaag	atacacaaca	gcaaatcatc	180
agggagactt	tccatttggg	atctaagaga	gatgaaaatg	tttgtaattt	cctagaagga	240
ggattattaa	ttggaggatc	tgacaacaaa	ctgatttata	gacattatgc	aacgttatat	300
tttgtcttct	gtgnnggatt	cttnanaaag	tgaacttggc	atttttag		347

<210> 138

<211> 434

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(434)

<223> n = A,T,C or G

<400> 138

cctttttttt	tttttttggg	taaatgactt	actgtgtaat	tttatttcat	attacacaaa	60
tgtaaatcaa	atgctgagta	gacatgcaga	tgacaagcag	tatatgacaa	actctgaana	120
aatagttaca	tgtagagttt	ctcanatttt	tagtgtatct	aanaattaac	tgaagagttt	180
gttaagaatg	caggcttaaa	ggccaatcca	cagattataa	tttcatacaa	acaggatgga	240
gcctaanaac	ctgtaaatta	ttaaacaact	gattaaaaat	agagagggtt	ctatgaagtt	300
aggnntgtcc	ttattttctta	tttgaactgg	acaagtagaa	ggataatagg	taggaccaag	360

tgagcattat cagaatcaaa gtagaggcaa taacaagcca aggtgtttta ncctanctaa 420
agaagctcgt cgac 434

<210> 139
<211> 553
<212> DNA
<213> Homo sapien

<400> 139
gtcgacctga ctataacagt gcctactatg ttaacattag atgaacaagt gaattagagg 60
atTTTTTaaat gtgtatccat cagtgtatgg acacactccc tctaacttct tcaaaaaaca 120
aaaatttcctg gtagagctaa gtggTTTTTta gaagtttggg tttggtaact gatttctacg 180
agataattga acactTTTTa aaatagttga tcattatgtc aaacagccct caacagtaaa 240
cttaaattag gtagaattat agtaagctgg aagagaaaat gttcccaaag agcattagtc 300
cctttctggc accttattac agatgaataa attgagactc acagaaatta aatgacttag 360
ccccagttat ccaactaact ccttaatgtg aggccatgat taggaatagg cttctagtat 420
tcagtcccat attattttga ctgtgtaata ccacgtgcc aattgatttt aaagtcaaat 480
ctcggcctga actgtatggg gaaaaaaaa atctccagct ggctctgctg aatccccaga 540
ggggccctcc act 553

<210> 140
<211> 450
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)...(450)
<223> n = A,T,C or G

<400> 140
gtcgacgcog gtgagttggg tgccgggtgga gtcgtgttgg tcctcagaat ccccgcgtag 60
ccgctgcctc ctccctaccct cgccatgttt cttaccoggt ctgagtacga caggggcgtg 120
aatactTTTT ctcccgagg aagattattt caagtggaa atgccattga ggctatcaag 180
cttggttcta cagccattgg gatccagaca tcagaggggtg tgtgcctagc tgtggagaag 240
agaattactt ccccatgat ggagcccagc agcattgaga aaattgtaga gattgatgct 300
cacatagggt gtgccatgag tgggctaatt gctgatgcta agactttaat tgataaagcc 360
agagtggaga cacagaacca ctggttcacc tacaatgaga caatgaacag nggagagtgt 420
gacccaagct gngtccaatc tgnctttgca 450

<210> 141
<211> 140
<212> DNA
<213> Homo sapien

<400> 141
acacacccct ccctcacaca gggtctgacc gccgctggca gttecagggc taaggatttc 60
ctgcaattac ttgtggagaa ggagttcata gctgggctcc tggaggggag atagagcttc 120
tctttcgttc ccgggtcgac 140

<210> 142
<211> 591
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)...(591)

<223> n = A,T,C or G

<400> 142

gtcgacctgg	acttgcagtg	taaacagaga	cgctgcaa	tgcttgtgga	cggtgtaggc	60
cgctgcaggc	caccatgaac	cggttccgg	atgactacga	cccctacg	gttgaagagc	120
ctagcgacga	ggagccggct	ttgagcagct	ctgaggatga	agtggatgtg	cttttacatg	180
gaactcctga	ccaaaaacga	aaactcatca	gagaatgtct	taccggagaa	agtgaatcat	240
ctagtgaaga	tgaatttgaa	aaggagatgg	aagctgaatt	aaattctacc	atgaaaacaa	300
tggaggacaa	gttatcctct	ctgggaactg	gatcttcctc	aggaaatgga	aaagttgcaa	360
cagctccgac	aaggtactac	gatgatatat	atthttgattc	tgattccgag	gatgaagaca	420
gagcagtaca	ggtgaccaag	aaaaaaaaa	agaaacaaca	caagattcca	acaaatgacg	480
aattactgta	tgatcctgaa	aaagataaca	gagatcaggc	ctgggttgat	gcacagnгаа	540
aggggttacc	atggtttggg	ancacaggag	atcacgtcaa	caacagcctg	t	591

<210> 143

<211> 538

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(538)

<223> n = A,T,C or G

<400> 143

gtcgacaaat	aagaagacac	cttcagcatc	ttaaactaga	ataaataaaa	gaaggggtggc	60
ctcctagaat	ttaagtcagg	agggaggtgg	tgggcaatgg	atgacaagct	ctactttgaa	120
gaggttgaat	ttcagctgac	cactactaaa	gcagtacaag	cttttccttt	cagcaagtgt	180
cttcccagaa	atgtgatagc	aatttttagg	aagaatttgg	caaacataat	gtttagcaga	240
tttgcaacaa	atgctataag	ctcaaatttt	tttttttttt	tttttnggca	gcacactcag	300
ccctccaagg	ggaagtggat	tatttttctt	gcaagtgcac	tancanggga	ggtattaagg	360
acagcaacat	tccttcctgt	ataaaaaaat	aaataaataa	aagaagaaag	gattattgag	420
gccctctctg	ctgnatgtaa	tgtacttcan	gatgttggtg	naaaagatat	caacctanaa	480
taagnttcac	aanaatacat	ttggtttcac	ngaaagttta	aagtcaatct	ggacattc	538

<210> 144

<211> 401

<212> DNA

<213> Homo sapien

<400> 144

gtcgacctgt	tccctttttg	ggcctgtctc	cccatgtata	tgttgagggg	ttggacttca	60
gggcctgtga	gaggccttcc	aacttagact	ttctccccag	gagcataaat	tcagtgaatc	120
tacgtgactc	tcagtgatgg	catcattgcc	taatatccac	ccagcttctg	cttgaaaact	180
tcagagact	ggttcacatg	gggtataaaa	agcccaggcc	ccttgcccca	acttgggaca	240
actatgaaga	gtttccagct	ccacagctcc	ctgaggggct	ggccgaggcc	tttgtggggt	300
ttgcctcaca	accgaattta	tccctctggc	caattctgct	tcaatcactc	cctgccagggt	360
gttgaccttg	aatgtactcc	cccaataaac	ctcctgcaag	c		401

<210> 145

<211> 367

<212> DNA

<213> Homo sapien

<400> 145

cctttttttt	tttttttttag	ttagaaatta	caagtttatt	tttatatttt	gaaaaaggca	60
taatagaaaa	caaaaaataa	caaccaggca	tatcaatatt	tgtgacatac	acatacacac	120
aaaaatgaat	ataggaaata	acacgaagaa	aaagcatagt	atgttttgaa	accaacgtgg	180

```

ggcatgaaca gatttttgat gaaatacaac taaaggtttt aagtgtctat gtaatgttcg      240
agataattacg atcactctta tcctactagc aaaaatttagc aaactaggct ttaaaacatg      300
attcctgttg ttttagcagg atttattttg gtaatgatcc tgcttcctta taaacaacta      360
cgtcgac                                           367

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<210> 146
<211> 395
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(395)
<223> n = A,T,C or G

```

```

<400> 146
gtcgacaaga aagccccctt aatgttttta actgatgata tttttttaag cttaccaata      60
taagtatttt taaaggttct atttttcaaa gtcataacaa tgattgttct tgttttctct      120
catagaatag actgccatcg gataaagagt ggtccctagc ttctattttt ccaagtaaat      180
aagtagaaca tgttcttggg attataccat taaatgttaa ttttcttgaa gaagaaagat      240
tgttgtctgc caagatttta tgttagcgct cggattgagg cagaaaacgg aagcaccagg      300
tttaacactg ggatgacttg ggttgtgttc ctggagggtt gaagngggcc ttccccgcct      360
tttgaggggg aaaactgact gnnttgaaca catat                                           395

```

```

<210> 147
<211> 455
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(455)
<223> n = A,T,C or G

```

```

<400> 147
gtcgactaaa aactggaacg gtgaagggtga cagcagtcgg ttggagcgag catcccccaa      60
agttcacaaat gtggccgagg actttgattg cacattgttg tttttttaat agtcattcca      120
aatatgagat gcgttggttac aggaagtccc ttgccatcct aaaagccacc ccacttctct      180
ctaaggagaa tggcccagtc ctctcccaag tccacacagg ggagggtgata gcattgcttt      240
cgtgtaaatt atgtaatgca aaattttttt aatcttcgcc ttaatacttt tttattttgt      300
tttattttga atgatgagcc ttcgtgcccc cccttcccc ttttttgtcc cccaacttga      360
gatgtatgaa ggcttttggg ctccctggga gtgggtggan gcagccaggg cttacctgta      420
cactggactt gagaccagt t gaaataaaaag tgcac                                           455

```

```

<210> 148
<211> 518
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(518)
<223> n = A,T,C or G

```

```

<400> 148
gtcgacctca cgccttcgcc gtagcatott tcgcagcgga ccgaagagaa gaaaagtagg      60
ccagagccga actctcttcc tgccaagatg tctattgttg tgccgattaa agtactgcat      120
gaggccgagg gccacattgt gacatgtgag acgaacaccg gtgaggtata tcgggggaag      180

```

ctcattgaag	cagaggacaa	catgaactgc	cagatgtcca	acatcacagt	cacatacaga	240
gatggccgag	tggcacagct	ggagcaggta	tacatccgtg	gcagcaaaat	ccgctttctg	300
atthtgcctg	acatgctgaa	gaacgcaccc	atgttaaaga	gcatgaaaaa	taaaaaccaa	360
ggctcagggg	ctggccgagg	aaaagctgct	attctcaagg	cccaagtggc	cgcaagagga	420
agaggacgtg	gaatgggacg	tggaaacatc	tttcaaaagc	gaagggataa	ttttctaagt	480
tgaacagaac	tttgtccttt	tttctttcan	ggtatctg			518

<210> 149

<211> 442

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(442)

<223> n = A,T,C or G

<400> 149

cctttttttt	ttttttttct	tttcataaaa	tttttacttt	atgaattaaa	tacattgaga	60
aacagnaaaa	atatatttac	agtcatttga	agnnggcact	actaacatat	ttaattttaa	120
aaaatctttg	ctgtttcttt	gcctgtttct	ttcaaagaga	attttaaata	tgacttttagc	180
ttttaaaaaa	tacaatang	aaataattac	attcttaata	tgaaaacatt	ttacaacgta	240
tcaccatggt	caattaattc	tgaatatcac	ttaaaagttg	atgtttaa	gtaaagngaa	300
tatttccttt	cttggtanaa	aatcaaaaag	attatctcat	taaaaacacc	ttnggnccta	360
agacttatga	tctgaanatg	nccttttgaa	aagnatcttc	catggctaca	actaaaaaan	420
acccggtaac	acttgtgcac	gg				442

<210> 150

<211> 341

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(341)

<223> n = A,T,C or G

<400> 150

gtnnacctat	tattacccca	tgatacagtt	tagaaaacaa	attcatgcac	taagtaa	60
gaccaa	aatc	g	taagtcactg	ccttttgctc	cagagttggc	120
ttaactagtc	aactttaaag	aaaaaaattt	ttttttctgt	gaaggaaatt	aagtcctat	180
tttcanagag	ctaaaagcaa	tcaaggcatc	tactgtgtta	ttttcctatc	catgtngact	240
catgtttaag	gttgactagg	aagacataat	cattggctgc	taataacaaa	tngatttctt	300
ttnataaaaa	attttaaaga	gtntntaatg	cctttatttta	t		341

<210> 151

<211> 459

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(459)

<223> n = A,T,C or G

<400> 151

gtcgaccagg	tottgaccct	ggtcaacaag	agaataggcc	tttaccgtca	ctttgacgag	60
accgtcaata	ggtacaagca	atcccgggac	atctccaccc	tcaacagtgg	caagaagagc	120

```

ctggagactg aacacaaggc cttgaccagt gagattgcac tgctgcagtc caggctgaag 180
acagaggggt ctgatctgtg cgacagagtg agcgaaatgc agaagctgga tgcacaggtc 240
aaggagctgg tgctgaagtc ggcggtggag gctgagcgcc tggaggctgg caagctcaag 300
aaagacacgt acattgagaa tgagaagctc atctcaggaa agcgccagga gctggtcacc 360
aagatcgacc acatcctgga tgccctgtag cccctgcccg catcctncag ggggccagg 420
gtgccctgcac tttgctgtgg gnangcagat tgggtggta 459

```

```

<210> 152
<211> 242
<212> DNA
<213> Homo sapien

```

```

<400> 152
gtcgacccaa ggtcacagga gcattgcgtc gctgatgggg ttgaagtttg gtttggttct 60
tgtttcagcc caatatgtag agaacatttg aaacagtctg cacctttgat acggtattgc 120
atttccaaag ccaccaatcc attttgtgga ttttatgtgt ctgtggctta ataatacatag 180
taacaacaat aatacctttt tctccatttt gcttgcagga aacatacctt aagttttttt 240
tg 242

```

```

<210> 153
<211> 57
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(57)
<223> n = A,T,C or G

```

```

<400> 153
cctttttttt tttttttttt ttccacatca ctccaggttt atngaattta taaaatt 57

```

```

<210> 154
<211> 437
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(437)
<223> n = A,T,C or G

```

```

<400> 154
cctttttttt ttttttttgt aatncagttt taatttattt tcatcacttt ttcttcataa 60
tccagatatt ttaaaatgca aagaaaatta actttcaatg atatgttcag ggactggcac 120
taaaaaaaat tttcagactg caaatgagtt atacaaatga aaatatcaaa tggagatcca 180
gttatcaaaa tgaaagcact caacatatta aaagttcaca agtatttgta ttgagcacat 240
tacaaaagtc agcttgctaa ctggtgtgat tttaaagaac tattgcanaa gtctgaanaa 300
aatanattta ttagttaact tataaagaga ttaaagagcg tgaaacaagt nttaaaaaana 360
aatttgngcc tttattanaa tgttaggcgt cnacgcggcc gctcnngtct anagggccccg 420
tttaaaccgg ctgatca 437

```

```

<210> 155
<211> 518
<212> DNA
<213> Homo sapien

```

```

<400> 155

```


gtcgacgtga	gccacagtca	cgccactgca	ttctatcctg	ggcaacagat	ggagaccttg	60
tctcaaaaaa	aaaaaatcc	tgacatcgct	atgtattccc	aactttatca	tttgtctgcc	120
tgtttagttt	tgacttatgt	tttttttttt	tccccctgt	ggacatgtag	ttgacggaaa	180
tcgtgaagga	actttaatat	tttattttaa	tttcccaaaa	ctaatacatgc	cttatgtgac	240
taatcttcag	tgataatatt	tcatctactg	atatattttc	ttgaggtgtg	taattttcag	300
tataccttaa	tcatttggtg	taaaaaagag	agagggtttt	gatatatgaa	tgctgttctt	360
gtaaaaatca	atcttgacac	tttattttta	actttttatt	ggtaatgaca	gtgggttttg	420
tacatcatga	ttttcaattt	aggatatctg	tctaatttgt	tttttcagag	taactatatt	480
ggaattcaat	aaaaatattc	aaaatttttc	ttaaaaaa			518

<210> 156

<211> 600

<212> DNA

<213> Homo sapien

<400> 156

gtcgacgttt	atttaagttc	atgtttcact	gtttgcactt	tgcatagaac	aatgggttta	60
ttcgctgatg	taaacggttc	gagtgaagaa	ttaatgcagt	aagtatgaca	acacatacac	120
acttgccctc	ccccatctcc	agaagagggg	agcagagtcc	gagcttatct	aaatatgaat	180
gtggccacaa	agctgtggaa	ggtgacaaa	cttaaacacc	tttgccctgg	ctctgcattg	240
tcacctagag	agcaagaggt	ctatagaaac	atcatgtcac	atgaaacgat	tctctgcttt	300
ttggttctga	acttgaagtc	cctaaactgc	aaaatctaag	agttgggtgg	ttattaaaaat	360
gcttttaaaa	agttaactgt	ggcaccaatt	ctaataatga	ccaacttggt	actgtttttt	420
tttggtttgt	tttggttttg	tgtgtgtgtg	tgtgtggcac	tgggaaaagt	ggaaacaaac	480
atgtattgaa	atacatattg	gaaataaaaa	tgggttgagc	gtcagtgata	ttctcccaga	540
atgtacttat	cttacctcgc	atgtactgta	gtcactcagt	atttgtatat	gttgctagaa	600

<210> 157

<211> 542

<212> DNA

<213> Homo sapien

<400> 157

gtcgacggct	gggaagtcag	ttcgttctct	cctctcctct	cttcttggtt	gaacatgggtg	60
cggactaaag	cagacagtgt	tccaggcact	tacagaaaag	tggtggctgc	tcgagccccc	120
agaaaggtgc	ttggttcttc	cacctctgcc	actaatctga	catcagtttc	atcgaggaaa	180
gctgaaaata	aatatgcagg	agggaacccc	gtttgcgtgc	gcccactcc	caagtggcaa	240
aaaggaattg	gagaattctt	taggttgctc	cctaagatt	ctgaaaaaga	gaatcagatt	300
cctgaagagg	caggaagcag	tggcttagga	aaagcaaa	gaaaagcatg	tcctttgcaa	360
cctgatcaca	caaataatga	aaaagaatag	aactttctca	ttcatctttg	aataacgtct	420
ccttggttac	cctggtattc	tagaatgtaa	atttacataa	atgtgtttgt	tccaattagc	480
tttggtgaac	aggcatttaa	ttaaaaaatt	taggtttaaa	tttagatggt	caaaagtagt	540
tg						542

<210> 158

<211> 526

<212> DNA

<213> Homo sapien

<400> 158

cacctcaggc	tgtggctctt	tgggcttctt	cctaatagcag	aagaagttgc	ccagcagcaa	60
aatcaggagg	gaggtgagca	cctcgccccc	cgccaggatg	aacacgtaca	tgtagacgtg	120
ggtcgcatcc	aggagtttgc	ctcccgaagg	gggccgacg	agcacggcca	ccgcctccat	180
cagcagcacc	aggccaatgg	caactggagaa	cttgtaggag	atgccaaaga	agatgcagaa	240
gaccacgagg	ccgcccgtagt	cgcccgcggt	agagcccgc	aggtccgcga	ggccgttgaa	300
gaacatggag	aagctgaaga	ggtagacgga	gtagggccgc	accttcccaa	gccccgccac	360
gaagcccgcg	gcccgcgcg	cgaagatgtc	aatgaagccc	aggatgggtg	gcagggaagg	420
ggccttggtg	tcgggcacgc	ccaggtcctt	ggcgtagctc	accacgaaca	cgggcggggac	480

57

<211> 77
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(77)
 <223> n = A,T,C or G

<400> 162
 cctttttttt tttttttttt ttnntttttt tttttttttt ttttagggaa anaaatctgg 60
 gttcctttta tttttga 77

<210> 163
 <211> 645
 <212> DNA
 <213> Homo sapien

<400> 163
 gtcgacaaac aatgaatagt ttttcattgt accatgaaat atccagaaca tacttatatg 60
 taaagtatta tttatttgaa tctacaaaaa acaacaaata atttttaaat ataaggattt 120
 tcctagatat tgcacgggag aatatacaaa tagcaaaatt gaggccaagg gccaagagaa 180
 tatccgaact ttaatttcag gaattgaatg ggtttgctag aatgtgatat ttgaagcatc 240
 acataaaaaat gatgggacaa taaattttgc cataaagtca aatttagctg gaaatcctgg 300
 atttttttct gttaaatctg gcaaccctag tctgctagcc aggatccaca agtccttggt 360
 ccactgtgcc ttggtttctc ctttatttct aagtggaaaa agtattagcc accatcttac 420
 ctcacagtga tgttgtagg acatgtggaa gcactttaag ttttttcac ataacataaa 480
 ttattttcaa gtgtaactta ttaacctatt tattatttat gtatttattt aagcatcaaa 540
 tatttgtgca agaatttgga aaaatagaag atgaatcatt gattgaatag ttataaagat 600
 gttatagtaa atttatttta ttttagatat taaatgatgt tttat 645

<210> 164
 <211> 434
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(434)
 <223> n = A,T,C or G

<400> 164
 gtcgaccgga cgcggcggca ttaaaccggt gcaggcgtag cagagtggtc gttgtctttc 60
 taggtctcag ccggtcgtcg cgacgttcgc ccgctcgtc tgaggctcct gaagccgaaa 120
 ccagctagac tttcctcctt cccgcctgcc tgtagcggcg ttgttgccac tccgccacca 180
 tgttcgaggc gcgcctggtc cagggtcca tcctcaagaa ggtgttgag gcactcaagg 240
 acctcatcaa cgaggcctgc tgggatatta gctccagcgg tgtaaacctg cagagcatgg 300
 actgtccca cgtctctttg gtgcagctca ccctgcggtc tgagggcttn gacacctacc 360
 gctgcgaccg caacctggcc atgggcgtga acctcaccag tatgtncaaa atactaaaat 420
 gcgcngcaa tgaa 434

<210> 165
 <211> 388
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature

<222> (1)...(388)

<223> n = A,T,C or G

<400> 165

gtcgaccatt	catatatata	tgcatatata	tgtgaagctc	catatttctg	ttgctttaaa	60
gaagtaaaac	cttccattta	aataagatga	catgcntaan	ataacaaagc	ttccttgatt	120
tccttttcct	gtgtaattna	atagatttgt	tgactagtgc	ttgggcacat	tataaatcag	180
ngttatttgc	tcttggagcc	attttttaaa	aaaaattttg	gcagtgagca	gttgaattta	240
tcttgaattt	atcatgtgtg	tgtatttctg	aagcagctac	atagcagaac	attttaagag	300
attctgttag	cccacatgtt	catgttgggt	gctgctgaat	ggtaaatt	aaataaaatt	360
accagattaa	tcttaaaaaa	aaaaaaaa				388

<210> 166

<211> 443

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(443)

<223> n = A,T,C or G

<400> 166

gtcgaccttg	ctttcttaaa	aaacaaaaaa	actactgtca	gtattaatac	tgagccagac	60
tggcatctac	agatttcaga	tctatcattt	tattgattct	taagcttgta	ttaaaaacta	120
ggcaatatca	tcatggatac	ataggagaag	acacatttac	aatcattcat	tgggcctttt	180
atctgtctat	ccatccatca	tcatttgaag	gcctaataata	tgccaagtac	tcacatggta	240
tgcattgaga	cataaaaaag	actgtctata	acctaataa	gtattaaaaa	tcccattatt	300
acccataagg	ntcatcttat	ttcattttta	gggaataaaa	ttacatgtct	atgaaatttc	360
aattttaagc	actattgnnt	ttcatgacca	taattttatt	ttaaaaataa	attaaaggtt	420
aattataaaa	aaaaaaaaaa	aag				443

<210> 167

<211> 608

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(608)

<223> n = A,T,C or G

<400> 167

gtcgactgcg	cctctccgaa	cgcaacatga	aggtgctcct	tgcgcgcgc	ctcatcgcg	60
ggtccgtctt	cttctgtctg	ctgcccggac	cttctgcggc	cgatgagaag	aagaaggggc	120
ccaaagtcac	cgtcaagggtg	tattttgacc	tacgaattgg	agatgaagat	gtaggccggg	180
tgatctttgg	tctcttcgga	aagactgttc	caaaaacagt	ggataatttt	gtggccttag	240
ctacaggaga	gaaaggattt	ggctacaaaa	acagcaaatt	ccatcgtgta	atcaaggact	300
tcatgatcca	gggcccggagac	ttcaccaggg	gagatggcac	aggaggaaag	agcatctacg	360
gtgagcgctt	ccccgatgag	aacttcaaac	tgaagcacta	cgggcctggc	tgggtgagca	420
tggccaacgc	aggcaaagac	accaacgggt	cccagttctt	catcacgaca	gtcaagacag	480
cctggctaga	tggcaagcat	gtggtgtttg	gcaaagttct	agagggcatg	gangtggtgc	540
ggaangtgga	gagcaccaag	acagacagcc	gggataaacc	cntgaangat	gtgatcatcg	600
cagactgc						608

<210> 168

<211> 569

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(569)

<223> n = A,T,C or G

<400> 168

gtcgacgcgg	ncggccggac	agactgacgt	gtgagctgca	tcgcgggagg	cgcatggngg	60
ggatggcgct	ggcgcgggcc	tggaagcaga	tgctctggtt	ctactaccag	tacctgctgg	120
tcacggcgct	ctacatgctg	gagccctggg	agcggacggt	gttcaattcc	atgctgggtt	180
ccattgtggg	gatggcacta	tacacaggat	acgtcttcat	gccccagcac	atcatggcga	240
tattgcacta	ctttgaaatc	gtacaatgac	caagatgcga	ccaggatcag	aggtttcttg	300
gggaagaccc	accctacgaa	gttggaatga	gaccatcaga	tgtgataaga	aactcttcta	360
gatgtcaaca	taaccaacct	tataaagact	aaaattcatg	agtagaacag	gaaaatcatc	420
ctgactcatg	tgttgtgttc	tttattttta	attttncaaa	gaggctcttg	tatagcagtt	480
ttttgtctat	tttaacattg	taagtcattt	tgttctttga	natcantatt	ttcttaacct	540
ttgtgactgt	ttcaatatta	cccccgnga				569

<210> 169

<211> 216

<212> DNA

<213> Homo sapien

<400> 169

gtcgaccggg	aaccatcta	taaagtaagg	cacactcgta	atggttgaat	tgtgttctgg	60
ttaatttcct	aaaggacttc	acagttgcac	ttatgaaaat	gattttatat	tgaaatgata	120
tttgcataag	aaaaagcatg	tgatttaattg	catattgctt	gagtgttcat	ctgtgaatgt	180
gaaaaataag	ctgttttttt	ttattagata	tttgca			216

<210> 170

<211> 284

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(284)

<223> n = A,T,C or G

<400> 170

cctttttttt	tttttttgaa	atggancttc	tgaatcgaaa	agtttttcac	tttaaagtgt	60
ggatgagtgc	tacaaaaaca	ctnngcatct	tagggcaagt	gtcgctgagc	acctgcttcc	120
ccatattctc	agcannatca	tttcagttct	tagcaatctg	gcaggcaaaa	ggaaagtctg	180
attttgntng	aattngcatt	ttcctgatta	ccancaaact	antttaagct	taatgggcac	240
ntnntatttc	tattctctga	actgcccatt	tttctaccat	tcag		284

<210> 171

<211> 541

<212> DNA

<213> Homo sapien

<400> 171

cagacagcac	tgtgttggcg	tacaggctctt	tgcggatgtc	cacgtcacac	ttcatgatgg	60
agttgaaggt	agtttctgtg	atgccacagg	actccatgcc	caggaaggaa	ggctggaaga	120
gtgcctcagg	gcagcggaac	cgctcattgc	caatggtgat	gacctggccg	tcaggcagct	180
cgtagctctt	ctccaggagg	gagctggaag	cagccgtggc	catctcttgc	tcgaagtcca	240
gggcgacgta	gcacagcttc	tccttaatgt	cacgcacgat	ttcccgtctg	gccgtgggtg	300

tgaagctgta	gccgcgctcg	gtgaggatct	tcatgaggta	gtcagtcagg	tcccggccag	360
ccaggccag	acgcaggatg	gcatggggga	gggcataccc	ctcgtagatg	ggcacagtgt	420
gggtgacccc	gtcaccggag	tccatcacga	tgccagtggg	acggccagag	gcgtacaggg	480
atagcacagc	ctggatagca	acgtacatgg	ctgggggtgt	gaaggctctca	aacatgatct	540
g						541

<210> 172
 <211> 573
 <212> DNA
 <213> Homo sapien

<400> 172						
gtcgactttc	aacaaatcct	gaagtctttc	tgtgaagtga	ccagttctga	actttgaaga	60
taaataattg	ctgtaaattc	cttttgattt	tctttttcca	ggttcatggg	ccttggtaat	120
ttcattcatg	gaaaaaaatc	ttattataat	aacaacaaag	atttgatat	ttttgacttt	180
atatttcctg	agctctcctg	actttgtgaa	aaaggggtga	tgaaaatgca	ttccgaatct	240
gtgagggccc	aaaacagaat	ttaggggtgg	gtgaaagcac	ttgtgcttta	gctttttcat	300
attaaatata	tattatattt	aaacattcat	ggcatagatg	atgatttaca	gacaatttaa	360
aagttcaagt	ctgtactgtt	acagtttgag	aattgtagat	aacatcatac	ataagtcatt	420
tagtaacagc	ctttgtgaaa	tgaacttggt	tactattgga	gataaccaca	cttaataaag	480
aagagacagt	gaaagtacca	tcataattaa	cctaaatttt	tgttatagca	gagtttcttg	540
tttaaaaaaa	aataaaatca	tctgaaaagc	aaa			573

<210> 173
 <211> 545
 <212> DNA
 <213> Homo sapien

<400> 173						
gtcgacctgg	gctggacgtg	gttttgtctg	ctgcgcccgc	tcttcgcgct	ctcgtttcat	60
tttctgcagc	gcgccagcag	gatggcccac	aagcagatct	actactcgga	caagtacttc	120
gacgaacact	acgagtaccg	gcatgttatg	ttaccagag	aactttccaa	acaagtacct	180
aaaactcatc	tgatgtctga	agaggagtgg	aggagacttg	gtgtccaaca	gagtctaggc	240
tgggttcatt	acatgattca	tgagccagaa	ccacatatte	ttctcttttag	acgacctctt	300
ccaaaagatc	aacaaaaatg	aagtttatct	ggggatcgtc	aaatcttttt	caaatttaat	360
gtatatgtgt	atataaggta	gtattcagtg	aatacttgag	aaatgtacaa	atctttcoatc	420
catacctgtg	catgagctgt	attcttcaca	gcaacagagc	tcagttaaat	gcaactgcaa	480
gtaggttact	gtaagatgtt	taagataaaa	gttcttccag	tcagtttttc	tcttaagtgc	540
ctgtt						545

<210> 174
 <211> 469
 <212> DNA
 <213> Homo sapien

<400> 174						
gtcgacaaag	aatcacagct	ttctctccat	gttttattaa	cacacagaaa	aatactttga	60
aaaatatacc	atttctcaaa	aatgaaatgt	atgatttgct	acaaatggcc	atatggaaaa	120
tatgatacct	gcttatTTTT	gactcagggg	gcattcaatt	tttatactaa	ctgaaaatta	180
catgattgcg	ttttgtttta	aaagtgaaaa	aaagtaataa	ctgcttttag	ccttgtaata	240
ttgaatgcgt	caattggctc	cccttgtaga	atgttgaatg	gctatcactg	gtgacagatg	300
ttctgtacat	cgcagtaata	ctgcttatat	aattgtgata	attttccgct	tcttatttgt	360
catttttagt	gatttaaaaa	tcccttgatg	actccctgaa	aaatgactga	tgtttttctt	420
atattaagta	atttctgctg	gtaaagtgtg	agtcctttta	taatttctt		469

<210> 175
 <211> 108
 <212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(108)

<223> n = A,T,C or G

<400> 175

cctttttttt	ttttttttng	aaattnaagt	aacttnatnn	aaattcaaaa	acaatnctta	60
aaactgnntt	tagagtcaag	acccttttgt	attataaaaa	tcacaagt		108

<210> 176

<211> 426

<212> DNA

<213> Homo sapien

<400> 176

gtcgactgtt	tagaagttat	acacagagag	aaggggaaaa	gaaactccat	caatcaagct	60
aaaggcagca	aaggaaaatt	tgaaaagaag	caacgagact	gtttaacaaa	gaacatcaaa	120
taagatgatg	gaactagaag	aaaaacacca	atgtccttaa	ttatataaaa	acatcaatgt	180
ccttaattat	ataaattttt	aaccctcaat	tgggttaaaa	aatcagattt	gtactaagag	240
atgtatcttt	aaaagcaaaa	gaaagaataa	aaagatcaac	aagtaaaaca	aagtaggagt	300
cagaattaat	attagacaaa	ataaagggtga	aaaatactaa	atgcaagaaa	taatatttta	360
gatgacaaaa	atgtatgagc	cataaaaaag	tcatgagttt	ttataaacct	aaaatatagc	420
gtcgac						426

<210> 177

<211> 538

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(538)

<223> n = A,T,C or G

<400> 177

cctttttttt	tttttttttt	tttttttgga	ngnattnгаа	attttttcta	tatanatcat	60
gtgtgacttc	cataaagaaa	aataaacacc	tatacacagt	ttacctaata	tgtgtaatgt	120
taatgaaaag	aatcaaagaa	agatgttcgt	tcattaactc	tntaaatcaa	attgtttttc	180
cattttttacc	aacttgatac	cttaatcaag	tcactcttgt	tcttccttaa	gtgcaaatga	240
attttttggt	tgggttgggg	gacaacacaa	aatacaaac	tgggttgat	tcactgaaag	300
gccaanaaaa	gggccttant	ctaggaagta	nagngtgana	tgatacaccc	acaggctggn	360
gcattctggn	ccacacaaan	acgtgctgnt	ccccgccta	ctgntnaaaa	cagntctggt	420
ttgctnanat	gctgctgntg	caacctgcag	gtccatgana	agaacaactc	cctggttggt	480
tacancccg	gagtgttttg	ngaatttgca	cctacatttc	ccatgtgata	tggactca	538

<210> 178

<211> 566

<212> DNA

<213> Homo sapien

<400> 178

gtcgacttgg	aagcagggtt	atttattata	tacttgcaat	tgaatataag	atacagacat	60
atatatgtgt	tatgtatttc	tagaaatgca	cataacatat	atttgcctat	tgtttaatgt	120
ttttccaga	tatttattac	agaagggcat	ggagggatac	ctacttattc	ttcattatga	180
gaacaattaa	aggcatttat	tagataggaa	attaacagat	catctgcttc	tataacttta	240
ttagctacat	taaataggca	gtgagcaata	atttaaaac	tcaccattat	ataaaataat	300

aaataacaaa	gtaaaagtta	atgttataaa	aataaactga	tagtaaggaa	aatctaaatg	360
ggcatgatcc	cattttagaa	gaccaaata	ttaatagggt	tgtcatgtta	taatagacaa	420
ttgtctaatt	atttctgtgt	ttttatttag	tgggtagcag	aagttgttca	gaagagcaga	480
aatatgtaga	aaacatctct	aaatttttgg	caatttgaaa	tagcaattct	gaggcacaca	540
gctcatctac	aaaaatcttt	tgcaga				566

<210> 179

<211> 277

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(277)

<223> n = A,T,C or G

<400> 179

gncgacggga	aaggaatatt	atggcannaa	gctgagcaag	caattctggt	ggaaagtcaa	60
acctgtcagt	gctccacacc	agggctgtgg	tcctcccaga	catgcatagg	aatggccaca	120
ggtttacact	gccttcccag	caattataag	cacaccagat	tcagggagac	tgaccaccaa	180
gggatagtg	aaaaggacat	tttctcagtt	gggtccatca	gcagtttttc	ttcctgcatt	240
tattgnngaa	aactatngtt	tcatttcttc	ttttata			277

<210> 180

<211> 349

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(349)

<223> n = A,T,C or G

<400> 180

cctttttttt	tttttttttt	tttttttttt	tttttttttt	ttagnataag	gaaaagctac	60
aaacctcaag	gntgttttat	ttaaaccaa	taanttgagc	aagacatatn	tacattaaaa	120
acaaatgaac	acattaaaa	ttcactat	ttcactat	attctagcaa	catatacaaa	180
tactgagnga	ctacagtaca	tgccgnggta	ananaagtac	attntgggan	aatatnactg	240
acnctcaaac	cattttttat	ttccaatat	atttcaatac	atgtttgttt	ccactttttc	300
cagngccaca	cacacncnca	cacaaaaaca	aaacaaaaca	aaaaaaaaac		349

<210> 181

<211> 435

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(435)

<223> n = A,T,C or G

<400> 181

cctttttttt	ttttttttga	cattttacagg	tattttatttg	agtaagagct	cataaaatat	60
attttttata	tatgcacaag	aaaaaatata	tttgaatgaa	taaaaaataa	aatgacagga	120
ggtgacagaa	tttagtggtt	ataaatgagg	tcataaagaa	ctttaataat	tcanagaana	180
agttcaaagt	gtatttataa	gttgagaccc	tgctttacaa	tattttataa	ttttaaaaaa	240
aggcggttaa	aggtgatagg	tgacttaata	attttccact	ttcaaaatgg	gtttctagac	300
actgttatga	agctgctatg	tactaataat	actttgcttg	ccaaagtgtt	tgggttttgt	360

tgttgtttgt ttgtttgttt gtttttggtt catgaacaac agtgtctaga aaccacttt 420
caaaatgggg tcgac 435

<210> 182
<211> 328
<212> DNA
<213> Homo sapien

<400> 182
gtcgaccatt gtatcttttt cttttctatc cttttacatt tactctttca gaatccttat 60
gttttactgt tttcagaaaa cttagttttt aaaatattct gctaatacatt ttcatataa 120
gtttacatta aataagtctt ttaaagttta ttataattaa ataaagttta ttttcacatg 180
tgttttcata tctactgtct cagaactttc tccttgccct atttttccta ttttatcccc 240
tttttgcatc ttttgagttg actttttatg attttatttt tctctcttta ctagtgttga 300
tattatctac ccactaata ttctttca 328

<210> 183
<211> 491
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)...(491)
<223> n = A,T,C or G

<400> 183
cctttttttt tttttttttt tttttttttt ttacaaacct caaggttggt ttattttaa 60
caaataatct gagcaagaca tatatacatt aaaaacaaat gaacacatta aaatttcact 120
attttacaat ctaaattcta gcaacatata caaatactga gtgactacag tacatgcga 180
ggtaagataa gtacattctg gganaatatc actgacgctc aaaccatttt tatttccaat 240
atgtatttca atacatgttt gtttccactt ttcccagngc cacacacaca cacacaaaaa 300
caaaacaaaa caaaaaaaaa cagtcacaag ttggattaca ttanaattgg ngccacagtt 360
gactttaaaa gcattttaat aaccacccaa ctottanatt ttgcagttta gggacttcaa 420
gttcanaacc aaaaagcana gaatcgtttc atgtgacatg atgtttctat agacctcttg 480
ctctctaggt c 491

<210> 184
<211> 478
<212> DNA
<213> Homo sapien

<400> 184
gtcgacggct gctgttggtt gggggccgtc ccgctcctaa ggcaggaaga tggtgccgc 60
aaagaagacg aaaaagtcgc tggagtcgat caactctagg ctccaactcg ttatgaaaag 120
tgggaagtac gtctgggggt acaagcagac tctgaagatg atcagacaag gcaaagcgaa 180
attggtcatt ctgcgtaaca actgccagc tttgaggaaa tctgaaatag agtactatgc 240
tatgttygct aaaactggtg tccatcacta cagtggcaat aatattgaac tgggcacagc 300
atgcggaaaa tactacagag tgtgcacact ggctatcatt gatccaggtg actctgacat 360
cattagaagc atgccagaac agactggtga aaagtaaac ttttcaccta caaaatttca 420
cctgcaaac ttaaacctgc aaaattttcc ttttaataaaa tttgcttggt ttaaaaaa 478

<210> 185
<211> 596
<212> DNA
<213> Homo sapien

<220>

<221> misc_feature
 <222> (1)...(596)
 <223> n = A,T,C or G

<400> 185
 gtcgacggac gaggagtgcg gcactgatga gtactgcgct agtcccaccc gcggagggga 60
 cgcgggcgtg caaatctgtc tcgcctgcag gaagcgccga aaacgctgca tgcgtcacgc 120
 tatgtgctgc cccgggaatt actgcaaaaa tggaaatatgt gtgtctttctg atcaaaatca 180
 tttccgagga gaaattgagg aaaccatcac tgaaagcttt ggtaatgatc atagcacctt 240
 ggatgggtat tccagaagaa ccaccttgtc ttcaaaaatg tatcacacca aaggacaaga 300
 aggttctgtt tgtctccggt catcagactg tgcctcagga ttgtgttgtg ctagacactt 360
 ctggtccaag atctgtaaac ctgtcctgaa agaaggtcaa gtgtgtacca agcataggag 420
 aaaaggctct catggactag aaatattcca gcgttgttac tgtggagaag gtctgtcttg 480
 ccggatcacag aaagatcacc atcaagccag taattcttct aggcttcaca cttgncagag 540
 acactaaacc agctatccaa atgcagtga ctccttttat ataatagatg ctatga 596

<210> 186
 <211> 314
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(314)
 <223> n = A,T,C or G

<400> 186
 gtcgactgcc tatttaaatgt agctaataaa gttatagaag cagatgatct gttaatttcc 60
 tatctaataa atgcctttta ttgttctcat aatgaagaat aagtaggtat ccctccatgc 120
 ccttctgtaa taaatatctg gaaaaaacat taaacaatag gcaaatatat gttatgtgca 180
 tttctagaaa tacataacac atatatatgt ctgtatctta tattcaattg caagtatata 240
 ataaataaac ctgcttccaa acaacaaaaa aaaaaaaaaa aaaaaaaaaan naaaaaaaaa 300
 aaaaaaaaaa aaaa 314

<210> 187
 <211> 331
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(331)
 <223> n = A,T,C or G

<400> 187
 cctttttttt tttttttatt cctcagngct tttgatttta attcttttgg catatctaaa 60
 tgtcagaaaag tgaatatata catacagaat tcaaaacacc ttctaaaat ggttattatt 120
 ggccantcat tnacatcttt attttgaaag tctgaattgn caaatagttc taaagtgcac 180
 tcttgcagct aataaatagc agcatttgtt tataaaacct taagaaattc agaccagggc 240
 tgganaagtc acaataaaaa atcagacatg atctanatat agtcttcctt aatcatctaa 300
 gacaaacact tgtgtgaatt agtttataag g 331

<210> 188
 <211> 567
 <212> DNA
 <213> Homo sapien

<400> 188

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gtcgacgctg aagaaggaaa agaaatgtgt gaaactcata ggagttcccg ctgacgctga      60
ggccttaagt gaaagaagtg gaaacacccc taactctccc aggttagctg ctgaatcaaa      120
gcttcaaaca gaagttaaag aaggaaaaga aacttcaagc aaattggaaa aagaaacttg      180
taagaaatta caccctattc tatatgtgtc ttctaaatct actccagaga cccagtgtcc      240
tcaacagtaa agacttgtct ttaataagag tacggtgcc a ctgcctcaa aagttactat      300
ggtgcttaag attgtcttga tctgacatat atcaccttct gggttattta ctcatgtgc      360
caggacctgg catthttcatg tgcctttgac caagtgttca gaatttgctt gactctaacc      420
tgagagctt cttaagtgat gcccttcat ggagcttcta tgacagtga taaactatta      480
attgaaggaa aatgttataa ttaatgtatc tatttgctgc attgtatatg gattaaatga      540
taaaaaacaa gtaatctacc ctcagag                                     567

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<210> 189
<211> 130
<212> DNA
<213> Homo sapien

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<220>
<221> misc_feature
<222> (1)...(130)
<223> n = A,T,C or G

```

```

<400> 189
cctttttttt tttttttttt tttttttttt tttttttttt tttttatcnc ctaagnanat      60
tttaataataa attttgaaca gttataaaaa anaaanangg cctttgggtc aataacanaa      120
cataacaaaa                                     130

```

```

<210> 190
<211> 426
<212> DNA
<213> Homo sapien

```

```

<400> 190
gtcgaccaac ttcccacata tatttactaa gatgattaag acttacattt tctgcacagg      60
tctgcaaaaa caaaaattat aaactagtcc atccaagaac caaagtttgt ataaacagg      120
tgctataagc ttggtgaaat gaaaatggaa catttcaatc aaacatttcc tatataacaa      180
ttattatatt tacaatttgg tttctgcaat atttttctta tgtccaccct tttaaaaatt      240
attatttgaa gtaattttatt tacaggaaat gttaatgaga tgtattttct tatagagata      300
tttcttacag aaagctttgt agcagaatat atttgcagct attgactttg taatttagga      360
aaaatgtata ataagataaa atctattaaa tttttctcct ctaaaaactg aaaaaaaaaa      420
aaaaag                                     426

```

```

<210> 191
<211> 550
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1)...(550)
<223> n = A,T,C or G

```

```

<400> 191
cctttttttt tttttttttt tttttttttt ttttagtngg gatatgacct ttattgaact      60
tatccaccan agnggaaata atgtctgtac aaaaccaaact gtttgttact ataacttctg      120
catcacaatt aaaatocaaa cagtttttta aaaacagtca actcaatcaa aaccactac      180
ttcanaatca atagcttntt tgaagccaca gtaacactta aatatggtta anactcgaat      240
gcanaaattt ggttggttgg aaagctaatt aaacttccaa cttgctcaaa tagaattaca      300
aaaaggcaaa attgtgtttt tcacananat acagnccact ggaatcacca acactggaca      360

```

gctgttanag	tatttanagt	cctganataa	caaggaatcc	aggcntcctt	taaacagtct	420
tctgttgnc	tttcttccca	atcananatt	tgtggatgtg	tggaatgaca	cnccaccag	480
caattgtagc	cttgatgann	gaatccaatt	cttcatctcc	acgaatagca	agttgcaagt	540
gacgaggggt						550

<210> 192
 <211> 299
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(299)
 <223> n = A,T,C or G

<400> 192						
cctttttttt	tttttttgaa	attnnaaatt	ttattacaaa	aactttttat	tgctataaga	60
aaaatatgta	ttaattctac	aaaataacat	tcagattatg	ttctaattca	attattcaat	120
acaattttatt	ctcttgtaaa	taagagaaac	ttatttagaa	tataaaatta	taacctaatg	180
acaaagctct	agtaaattgn	gaactacacc	tctacaccgg	gottaaatgc	atcctgatta	240
atgattttctt	catacatgtc	actttattta	tccaaaaaag	gatttgagtt	ctcgtcgac	299

<210> 193
 <211> 536
 <212> DNA
 <213> Homo sapiens

<220>
 <221> misc_feature
 <222> (1)...(536)
 <223> n = A,T,C or G

<400> 193						
tttttttttt	ttttttttat	tctnncaatt	tttatttctc	ttacatgctc	aaagaagcca	60
agcaaatacca	ggtatacatg	tatatgtttt	aatttttacag	gagagagaaa	gaggatataag	120
gcaagaattta	actacatttt	catttcacta	tttctttatg	agctctattt	tgctgctaag	180
ttcaagtttc	aaaaaaatta	ttaattcctc	tgctatgtta	tottgtccca	attcacaaaa	240
taacagggat	ttcccatgt	gactcaaaag	caagaatctt	actcctaaat	aacataaaca	300
gcaatatgtg	tgactactgt	cattcattaa	cttcgatggg	gaagttcatt	aaactgacca	360
ttaaaagaac	atttgaacaa	ttccaaaagg	gagcaaggat	aaatctccaa	atcacccaat	420
agacaaggaa	cccagagatg	acatacagng	tgctcacttc	cacccactgc	cactgagaac	480
actgattgct	ctcttcaaac	acagagcgaa	gaatgggcct	catgtcacat	ggggca	536

<210> 194
 <211> 566
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(566)
 <223> n = A,T,C or G

<400> 194						
gtcgactgca	ctattaccca	gggcagatat	tatgagaaac	tgtttcttct	ctaagggttt	60
atggcagact	ttgctttttt	aacatgtgag	aaatgaattt	tttattttgt	gatttatgtg	120
atttcttttg	ctgagtgaag	gaaaggagaa	attgttgcta	ttgtcagcat	cttaaaggta	180

tttccagtca	aggcaaggct	aagtgtttg	tgatagtatt	aagcaagtca	tgttttgaat	240
ggattacctg	tagtgactca	ttggaatgat	ataattatac	aagtaatgcc	aaaaaccaag	300
tcaaagccta	attaaccaa	gcactcattt	aaaaatcatc	atgtttggac	ctatctggac	360
ctctcagcac	tgtaaaatag	ttttggtttt	gtggcatatg	aatagctgtt	taacaaatca	420
aagttagctn	tttgcttctc	agcttttttg	ggcaatacaa	gttaagttct	taatggggag	480
acattatcat	ggcatgactt	aagggaacat	tggtttgtga	aggaaaaaca	gattatctaa	540
agccatctct	atgtttctgt	tcagat				566

<210> 195
 <211> 217
 <212> DNA
 <213> Homo sapien

<400> 195	
gtcgacataa	ataaatggaa
atgccatttt	ttcctaaatt
ggtttttggg	agacataaac
gaataactaa	agcaacctta
	caaaaaaaaaa
	aaaaaaag
	60
	120
	180
	217

<210> 196
 <211> 391
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(391)
 <223> n = A,T,C or G

<400> 196	
gtcgacggac	agacttagga
ttaacttttg	taaggtactg
gcttacaaatg	tgactgaat
ctttgaaata	tgcatgtact
gttatataaa	aaaattgtaa
caccaaaaaa	aaaaaaaaaa
aaaaaaaaaa	aaaaaaaaaa
	aaaaaaaaaa
	a
	60
	120
	180
	240
	300
	360
	391

<210> 197
 <211> 445
 <212> DNA
 <213> Homo sapien

<400> 197	
gtcgactgga	tctttatgtc
ttaaaaccac	acactgccct
gtagtgtcac	tttctcagct
aaaacctttt	tgacccaaaa
tgtgccaagt	cagaggcttt
gctgtttgac	tcaacagtct
cagagttcaa	ggtgtctgtt
ggtggctgga	aacagatttt
	tgctg
	60
	120
	180
	240
	300
	360
	420
	445

<210> 198
 <211> 463
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(463)
 <223> n = A,T,C or G

<400> 198
 gtcgacgtca gtattaatac tgagccagac tggcatctac agatttcaga tctatcattt 60
 tattgattct taagcttgta ttaaaaacta ggcaatatca tcatggatac ataggagaag 120
 acacatttac aatcattcat tgggcctttt atctgtctat ccatccatca tcatttgaag 180
 gcctaataata tgccaagtac tcacatggta tgcattgaga cataaaaaag actgtctata 240
 acctcaataa gtattaaaaa tcccattatt acccataagg ttcatcttat ttcattttta 300
 gggaataaaa ttacatgtct atgaaatttc aattttaagc actattgttt ttcatgacca 360
 taattttattt ttaaaaataa attaaagggt aattatatgc atgtatgtat ttctaataat 420
 taaaaatgtg ttcaatccct ganaaaaaaa aaaaaaaaaa aaa 463

<210> 199
 <211> 129
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(129)
 <223> n = A,T,C or G

<400> 199
 gtcgaccggc gggcagctgc agctttctgct gctgaggccg ggattgctac gactgggact 60
 gaagactcag acgatgccct gctgaagatg accatcagcc ancaagagtt tggccgnact 120
 gggcttcct 129

<210> 200
 <211> 523
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(523)
 <223> n = A,T,C or G

<400> 200
 cctttttttt tttttttttt ttttttnaat ctttatttaa aagtccatgc taataatgng 60
 ttacattttt tacagttaca ttatgataga aactgttgga tttttttaat atctaaaaca 120
 atggcccaact gaanaaagga acaattaact ctttaattaa ttccttagga taaataccca 180
 naaattttaac agctagggca gacttntaat acaataccga aagtccttcc aaaaaccaag 240
 nggttgccaa cttatgtccc ttagcattat aacattcttg agccaatagt gtaaaaaatac 300
 gctgacaatt ttataggcaa acattactca aggtatctta ctttcactt attactaaag 360
 taattaacccc ctaaacagat gtcctcaac agngggacta catcctggta aacctatcat 420
 aagttgaaac tatcaagttg aaatgcattt agtacctga taaacctatc ataaagttga 480
 aaatttgtaa attgaaccag tgtaaatacag aggccatntt act 523

<210> 201
 <211> 532
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature

<222> (1)...(532)

<223> n = A,T,C or G

<400> 201

```

cctttttttt tttttttaca cttgagctta gccaaaaggc tgagaagcga ttttttttta      60
aaagctgttc tttaccatgg tttaaacgct aaaatgcata gctataaaaa caaaacactg      120
agctaactctg attacatcca gcttttgac tcaatagccc ttgaccctcc agtcataagc      180
aagcctgtca ttcgcccagc cctgctatac attctcatta tagtttcgtt tcaaataccag      240
tgttacagaa acaaaacacc aagccctcaa tcatgctatg cgtatcttta tgtgtgcatg      300
tcttatgtat gtttaaaata aacattttta aatgttttag gccaggcttg gnggctcatt      360
cagtttttagt ttgctttttt tttgccattc tttgttattt tngaaataag taaaacattt      420
aaatacttaa gtcacatctg tataaaaagt atattcatag gaaggaaattt aacaatttta      480
ataaaaactta tttagcatatc aatgagtttc aagatacacc tgaaactaaa tt              532

```

<210> 202

<211> 114

<212> DNA

<213> Homo sapien

<400> 202

```

ctccttggtg tggctttctc tgagtgaatg tcacaaggcc ggtgacagga ggggggtggag      60
gtgagggggac aaagtagagg ccgaggggtca gtgccttttg agaaagtcca gaga          114

```

<210> 203

<211> 304

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(304)

<223> n = A,T,C or G

<400> 203

```

gtcgaccttt ttttcccaac ttcttgcttt ctattggatt gttagggatt tctgtttttc      60
actttatttc tctctgctta tttgaaagct atacagcatg gttttctttc tttagggatc      120
actcttcac tttacttttt aaagatggat aaattttata catttaaaaa atttaactctg      180
tatttgatc ttcttcctga gtggacctta gcatgttata aatgctcact gaataattct      240
cattgttaat tagagtttgg ttttatnttt ttaaanncaa tgtacttact tattcttagn      300
gtaa                                              304

```

<210> 204

<211> 581

<212> DNA

<213> Homo sapiens

<220>

<221> misc_feature

<222> (1)...(581)

<223> n = A,T,C or G

<400> 204

```

cngcgttggt aggtgagcnn tttcagaagc gcatgccag gacacgtcgg gaagcaagca      60
tccnttttagc tgcttggaag gaggaccaa gacggctaaa anntcatttg gaaatatctc      120
taaataatttg ttaccatgta taagctgcta aagagaaatt gggcccaaca aaactaattg      180
aataattgag gcagatttgt gtgtatcatc aaattctatc cagaagttga agaactctgaa      240
tttaaagatt gtgtgcattt aataagagga tgacctttca gtttaatttc actatagaag      300

```

```

accatctgga aaatgaatta acacccatta gagatggagc tttgaccctg gattcctcaa 360
aagagctgtc agtctcagaa agtcaaaaag gagaagagag ggacagaaaa tggtctgcag 420
aacaatttga cttgcctcag gatcacttgt gggaacataa gtcaatggaa aatgcagctc 480
cctctcaaga cacagacagt ccactcagtg cagccagcag ttcaaggaac ttggagccac 540
atggaaaaca gccctccttg agagctgcca aagagcatgc t 581

```

```

<210> 205
<211> 409
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(409)
<223> n = A,T,C or G

```

```

<400> 205
gccctgaaga acagtgcctg gatgtggtga cccactggat ccaggaaggt gaagaagggc 60
gtccaaagga tgaccgccac ctccgtggct gtggctacct tcccggctgc ccgggctcca 120
atggtttcca caacaacgac accttccact tctgaaatg ctgcaacacc accaaatgca 180
acgagggccc aatcctggag cttgaaaatc tgccgcagaa tggccgccag tgttacagct 240
gcaaggggaa cagcaccat ggatgctcct ctgaagagac tttcctcatt gactgccggg 300
gccccatgaa tcaatgtctg gtagccacgc gngcgacgtc acagagacnc ggaaaaacca 360
aagctatatn ggtaaagagg ctgtgcaacc cgctctcaat gtgccaaca 409

```

```

<210> 206
<211> 561
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(561)
<223> n = A,T,C or G

```

```

<400> 206
gtntcatggg aaaggacatg tctctcgaag aaaggttata aaccctgaga tatgaggggtt 60
tttttgagac atccgagcct gtttcgttcc gggntgggan caggaataac octgacttct 120
gagctttcat aaccccagga tcctccagaa aatttgcggc gcgctgaggg aaaaccttgc 180
tgaagctgta cattggaatg cgtttacagt cattgtaatg gaagcaaaat acatgaagga 240
aaaactgtta tttgtatccc tgcttattgc acctgacgac tagttgcaga tggttttgtt 300
tacctaagaa aacttgtgat ataaatgaaa aaaacacctg ttttcctaga gtcattgggtt 360
acaaatatgc ttcgtctaag agctatttgt ccattctcct ggagagtgtt tcaatttcga 420
cccatcagtt gtgaaccact aattattcag atgaataagt gtacagatga ggagcaaatg 480
tttggtttta ttgaaagaaa caaagccata ctttcagaaa agcaagtggg atgtgcattt 540
gatatgcttt ggaagcttca a 561

```

```

<210> 207
<211> 461
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(461)
<223> n = A,T,C or G

```

```

<400> 207

```



```

ggtntttcca gccaatgtga cctttaaaac ctatgaaggt ntnatgcaca gttcgtgtca 60
acaggaaatg atggatgtca agcaattcat tgataaactc ctacctcaa ttgattgacg 120
tcactaagag gccttgtgta gaagtacacc agcatcattg tagtagagtg taaacctttt 180
cccatgccca gtcttcaa atttctaatgtt ttgcagtgtt aaaatgtttt gcaaatatcat 240
gccgataaca cagatcaa ataatatctcct catgagaaat ttatgatctt ttaagtttct 300
atacatgtat tcttataaga cgaccagga tctactatat tagaatagat gaagcaggta 360
gcttcttttt tctcaa atgt aattcagcaa aataatcacg tactgccacc agatttttta 420
ttacatcatt tgaaaattag cagtatgctt aatgaaaatt t 461

```

```

<210> 208
<211> 296
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(296)
<223> n = A,T,C or G

```

```

<400> 208
gatgaacatc catccnaatt ncgaagagcc tatattatac cctcttcaag aatttgcattg 60
gcatcaatat ctacaggaga aaaaaaggga actcaaaaat gaaacctggg aatattcttc 120
ctctgtgatt tcttttggtta atggtcagtt tctgggtgat gcattggatc tgcagaaatg 180
ggcccacgag gtgtgggata tagttgacat taaaccctct gcactttatg acgcactcac 240
tgaggatttt tccgctaagt tcttaagaga caccaagcat gatttcgtgt ttttgg 296

```

```

<210> 209
<211> 282
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(282)
<223> n = A,T,C or G

```

```

<400> 209
gcataataaa tgctttgagc ttcttgacta tcatatacct aaagaaagtg catcagagaa 60
tnatatctct gacttttnnc tgactggcaa aaagcnagct ttatcttctc ttataggatg 120
cttagtttgc cactncaactt caaaccaatg ggacagtcnt anatggngng acagtgttna 180
ancncaccaa aaggntncnt ttcentgggg ccancnctgt cntnancctc nctaancat 240
ttgnanaatt ttaancncnn gttaantaaa aaaaaaaaaa aa 282

```

```

<210> 210
<211> 1445
<212> DNA
<213> Homo sapiens

```

```

<400> 210
ggcgttgtga ggtgagcttt ttcagaagcg cgatcccagg acacgtcggg aagcaagcat 60
ccccagagct gcttggaag aggaccaaag acgtctaaaa agtcatttgg aaatatctct 120
aaatatattg taccatgtat aagctgctaa agagaaattg ggccaacaa aactaattga 180
ataattgagg cagatttgtg tgtatcatca aattctatcc agaagttgaa gaatctgaat 240
ttaagattg tgtgcattta ataagaggat gacctttcag ttaatttca ctatagaaga 300
ccatctggaa aatgaattaa caccattag agatggagct ttgacctgg attcctcaa 360
agagctgtca gtctcagaaa gtcaaaaagg agaagagagg gacagaaaat gttctgcaga 420

```

```

acaatttgac ttgcctcagg atcacttgtg ggaacataag tcaatggaaa atgcagctcc 480
ctctcaagac acagacagtc cactcagtc agccagcagt tcaaggaaact tggagccaca 540
tggaataacag ccctccttga gagctgccaa agagcatgct atgcctaaag atttaaagaa 600
gatgttagaa aataaagtca tagaaacatt accagggtttc cagcatgtta agttatcagt 660
agtgaataacc atcttgttga aagagaactt ccctggagaa aacatagttt caaaaagctt 720
ttcttctcac tctgatctga ttacaggtgt ttatgaggga ggcttaaaaa tctgggaatg 780
taoctttgac ctcttggtt atttcacaaa ggccaaagtg aaatttgctg ggaaaaaagt 840
cttgatctt gggttggtt caggtttact aggtataact gcattcaagg gaggggtccaa 900
agaaattcac tttcaagatt ataacagtat ggtgattgat gaagtaacct tacctaattg 960
agtagctaac tccactttgg aagatgaaga aaatgatgta aatgagccag atgtgaaaag 1020
atgcaggaaa ccaaaagtaa cacaactata taaatgccga ttttttctg gtgagtgggtc 1080
tgagttttgt aagcttgtac taagtagtga aaaacttttt gtaaaatatg atctcattct 1140
cacctcagaa accattttaca acccagatta ttatagtaat ttgcaccaga ctttccttag 1200
actgttaagt aaaaatggac gtgtactttt ggccagcaaa gcacattatt ttgggtgtagg 1260
tggaggtgtt catctctttc agaagtttgt agaagaaaga gatgttttta agaccagaat 1320
actcaaaata attgatgaag gattgaagag gttcataatt gaaataactt ttaagtttcc 1380
tggttaatta acattcactg agtatccaaa atgaaataaa cagaaggacc aaaaaaaaaa 1440
aaaaa 1445

```

<210> 211
 <211> 414
 <212> DNA
 <213> Homo sapiens

```

<400> 211
aaaaaggga ggaaggagag acagataact ctacgtcatt taaaaaacta caataaaata 60
ttatgaatta tcaattagat caaagttcct cacagctata tttatatagg taataaaaaa 120
ttaaataggc taaatgccca aaaatttaag actggcaaaa tatacttggc taaatactgt 180
gcgtctctat taaataccat gtttcagaag aattattaat gacatgagaa tatgtcaca 240
atacatattg atatgtgcaa atacatattg caaagtaaga ttatagaatg atcctagttc 300
aaaaatgtca catatatatg tatttaaaaa aaaaggcagt taagatttac aacaaaatgt 360
tagtggtggg accttctggt aggaatacag attttttttt attcagaagt tttt 414

```

<210> 212
 <211> 720
 <212> DNA
 <213> Homo sapiens

```

<400> 212
gtcgacgtaa aatagaaaca gaaggggact ttatcaacct gattaacttt ctcaacatgt 60
taaccctaca gttaacatta taatcaatgg tgaatcattg agtactttcc ttctaagatc 120
agaaacagtt caaagtcac tctcaccatt tctattcaac attgtactgg aatcccagcc 180
agtgcagtaa taccaataat aaaaaattaa agtcataaag attgaaaagg atgaagtaaa 240
gctatttcaa ttctatttag aagtatttag aaaccccaaa gaatctacaa aaaactaata 300
gaaataagtg aatatatgaa ggtcttacta tacaagatca acatatcaaa agcagtggta 360
tttaagaaaa ggttgagac tatttataat aaacagtggg tgaattttgt taatgctttt 420
tctgtatttt ttgaaatgat cttattattt ttctctttgc taaaaatgtg agtaaccttg 480
agttgacttt ctgtgtaaat caaccttgtg tcccaggaaa aaactccaat tgatcatgat 540
gtgttatcct ttttatacat tgctgtattc aatatgctaa tatattttatt ttttgtgtct 600
atttcatgag ggatcagtc atgtaattgt tttttcttgt tatatctttg ttgggtttat 660
taatcaacat tatgctaact tcatacaata tattggaaca tgctccctcc ttttattttc 720

```

<210> 213
 <211> 1114
 <212> DNA
 <213> Homo sapiens

<400> 213

```

gctcctaaca aagaagatat cttgaaaatt tcagaggatg agcgcatgga gctcagtaag 60
agctttcgag tatactgtat tatccttgta aaacccaaag atgtgagtct ttgggctgca 120
gtaaaggaga cttggaccaaa acactgtgac aaagcagagt tcttcagttc tgaaaatgtt 180
aaagtgtttg agtcaattaa tatggacaca aatgacatgt ggtaaagat gatgaaaagct 240
tacaaatacg cctttgataa gtatagagac caatacaact ggttcttcct tgcacgcccc 300
actacgtttg ctatcattga aaacctaag tttttttgt taaaaaagga tccatcacag 360
cctttctatc taggccacac tataaaatct ggagacctg aatatgtggg tatggaagga 420
ggaattgtct taagtgtaga atcaatgaaa agacttaaca gccttctcaa tatccagaa 480
aagtgtcctg aacagggagg gatgatttgg aagatatctg aagataaaca gctagcagtt 540
tgcctgaaat atgctggagt atttgcagaa aatgcagaag atgctgatgg aaaagatgta 600
tttaatacca aatctgttgg gctttctatt aaagaggcaa tgacttatca cccaaccag 660
gtagtagaag gctgtgttgc agatatggct gttactttta atggactgac tccaaatcag 720
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<210> 214

<211> 1495

<212> DNA

<213> Homo sapiens

<400> 214

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<210> 215

<211> 838

<212> DNA

<213> Homo sapiens

<400> 215

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gtgcttggtt ctccacctc tgccactaat tcgacatcag tttcatcgag gaaagctgaa 180
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<210> 216

<211> 938

<212> DNA

<213> Homo sapiens

<400> 216

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cttgctgaag tagcggttca gcatgatgag cgagggctgg aagttgagtg ccaaacccaa 840
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<210> 217

<211> 1982

<212> DNA

<213> Homo sapiens

<400> 217

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1982

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<210> 218

<211> 592

<212> DNA

<213> Homo sapiens

<400> 218

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gtttggtttt attgaaagaa acaaagccat actttcagaa aagcaagtgg gatgtgcatt 540
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<210> 219

<211> 650

<212> DNA

<213> Homo sapiens

<400> 219

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<210> 220

<211> 782

<212> DNA

<213> Homo sapiens

<400> 220

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aa

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<210> 221

<211> 2417

<212> DNA

<213> Homo sapiens

<400> 221

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<210> 222

<211> 1466

<212> DNA

<213> Homo sapiens

<400> 222

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